

Chapter 4

Relationship Between Community Niche and Succession

The stability of biodiversity and community structure results from the interaction between the environment and the organism, and the interactions between organisms and the environment, as well as among organisms, are balanced over large temporal scales. Therefore, changes in the environment of a region inevitably rebalance community composition and the environment. The time required to achieve equilibrium and reconstitute the community corresponds to the time required for certain species to die out or for populations to change, as well as that required for new species to emerge (Bertness and Leonard, 1997). River ecosystems are banded, with large geographic spans and large vertical environmental variability; due to the small range of cross-sectional observations, it is difficult both to characterize the full range of river features and to study biological communities (Kareiva and Washington, 1995). The biodiversity in a given community is influenced by the spatial scale, and larger spatial scales include greater levels of environmental heterogeneity (Cavender-Bares *et al.*). Therefore, it is necessary to consider the boundaries of the biological community (Swenson *et al.*, 2006) in niche studies. The species chosen by the researchers can delineate the species boundaries of the community or can determine the geographical boundaries of species distributions. It is necessary to compare types of community composition at different geographic and taxonomic scales to reveal the mechanisms underlying the relationships between species and the environment in a heterogeneous habitat. Fausch *et al.* (2002) proposed the concept of an ovipositing field and a fattening field, but it was unclear how to determine the ovipositing field and fattening field as a research unit. We believe that the drift range from spawning ground to fattening ground as a spatial boundary is a solution to the problem. Because research goals differ, the data collection objects also vary. In this book, we generally continue to use the geographical names of simple habits to divide the geographic ranges of fish communities (some artificial river dams divide the areas), combining the distance between spawning grounds and the natural drift by an early fish resource at the river as an ecological unit for study. We also determined the “geographical boundary” using a “random” selection of species. Then, cross-analyses at different levels were performed to better understand the

classifications and ecological characteristics of ecological communities, to study the mechanism of species coexistence and diversity, and to determine the common characteristics and mechanisms that maintain species diversity.

Species diversity and species richness are related to the niche, and the niche includes the competition and mutual benefits among species in the community. Niche is not a simple biological species structure, and community species composition is a product of the dynamic adaptation of species to environmental change (YosiakiItô *et al.*, 1981). The biome is the main body of the ecosystem and is in relative equilibrium among species and with the environment; the biome has stable characteristics and can maintain dynamic stability despite small disturbances (Yodzis, 1981). The biological community can resist disturbance and adapt to environmental change. Species composition, including interspecies relationships, is the basis of ecosystem stability.

Both species and biological traits are influenced by the environment, and habitat and niche differentiation together shape the patterns of species coexistence in the community (Kraft and Ackerly, 2010). Species distributions have geographical boundaries, and species composition has regional distribution characteristics, reflecting the constraints of environmental boundary factors (Convertino 2011; Glor and Warren, 2011). Environmental and spatial characteristics influence species distribution patterns (Henriques-Silva *et al.*, 2013), indicating that the characteristics of species communities can also reflect environmental characteristics. For example, fish with suckers on the abdomen must inhabit alpine and canyon rivers. The number of species in a community is related to the size of the geographical range, and there is a positive correlation between the size of the range and species abundance (Otso Ovaskainen *et al.*, 2016). At a regional scale, more species in the community imply higher biomass abundance and a larger adaptive space, while fewer species imply lower biomass abundance and a smaller adaptive space. With the exception of isolated island biotas, the persistent negative relationship found on Indo-Pacific islands represents a significant departure from the accepted positive relationship between range size and abundance (Reeve *et al.*, 2016). The mechanisms of community species construction are a focus of ecological research, and the ability to derive community characteristics from taxonomic characteristics is critical for an understanding of ecosystem evolution and community species construction. Morphological data are included in the process of community change (Erös *et al.*, 2012).

In this chapter, the relationships leading to species change described in the previous chapter were analyzed with respect to species succession. The niche formation of the “subunit community” was analyzed using the model, and the influences of interspecific morphological factors on the niche were explored. It should be noted that no specific data on the communities and ecological niche occupancy of the 104 species of fish from the Pearl River system, which represent 56 fish genera, are available due to a lack of historic data.

The “subunit community” mainly included the dominant species in the middle and lower reaches of the Pearl River. From 104 primitive communities to 25 fish “subunit communities,” the total niche of four big fish reached more than 40%, and the catch of these four fish in the Pearl River System in the 1980s accounted for about 40% of the total catch, or about 50%–60% of the samples of the four major

species collected (The fishery resources survey and compilation committee of the Pearl River System, 1985). It was thus difficult to use “standard” data to measure changes in community niches. The “fish community morphological model” can be used to calculate the “spatial niche” proportion of various species in the simulated community, and the data seem to reflect the data for certain historical periods (or specific spatiotemporal ranges). For example, the results for the four big fish seem to be close to the historical data (*i.e.*, “the four big fish represented about 40% of total catch or about 50%–60% of the early resource samples”). From 2016 to 2018, our laboratory estimated that the catch of the four big fish in the Pearl River at Guangdong and Guangxi was about 10%. As a “standard” measure, the results of the model showed that the actual fish community was disturbed. Interestingly, the model results might thus reflect a decline in the function of the Pearl River ecosystem from 50% to 10% in recent decades. This type of result exemplifies the goals of the model construction.

4.1 Characteristics of Community Stability

In communities, species coexistence depends on mechanisms of equilibrium and stability. Stability mechanisms are essential for species coexistence and include resource allocation, and predation, as well as other mechanisms that depend on spatial and temporal environmental factors that change with population density fluctuations. Equilibrium mechanisms reduce large disparities in fitness to stabilize species coexistence and diversity. One of the key aims of community research is to quantify niche relationships among species. Community construction methods must consider the ways in which species interconnect in localized communities, including the effects of species, populations, patchiness, and other ecological spatial relationships (Leibold *et al.*, 2004). The analysis of community succession using morphological models may provide insights into interspecific species associations based on specialized morphological information.

Based on the fish community morphological model, the fish niche succession model was used to simulate the “primitive community” composed of 56 genera and 104 species of fish in the Pearl River system. The modeling result showed that the niche loss of any one fish in the community led to the niche rearrangement of other species and that this rearrangement process was more complex: the niches of some fish species increased, those of others decreased, and still, others remained unchanged. The model analysis divided the interspecific relationship in the community into two main categories: competitive and mutual benefit.

Linear river ecosystems span a large geographic area. Upstream and downstream fish become “central communities.” That is, closer relationships were formed within each group (upstream or downstream). In the analysis of niche rearrangement, some upstream (or downstream) fish were found to be absent, and these absences had little effect on the niches of the fish downstream (or upstream). Furthermore, the experimental communities were classified according to the distribution characteristics of fish in the upper, middle, and lower reaches of the Pearl River. The “large

community” of fish was decomposed into several “small communities” more closely related with respect to morphology and function. The results helped to clarify some mechanisms of community stability.

The rates of change in the various niches in the simulated community in response to the absence of single fish species were weighted by the average rate of change, and the average rate of change in the community was calculated as the comprehensive rate of change across the community. The comprehensive rate of change reflected the roles of the species in community stability. In the community, the species with low comprehensive change rates had a little decisive effect on community stability, while the species with high comprehensive change rates had substantial decisive effects on the stability of the community. For example, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, *Puntius semifasciolatus*, *Misgurnus anguillicaudatus*, and *Luciobrama macrocephalus* (see table 4.1) are the five species that were most important for the stability of the simulated community, while *Cyprinus chilia*, *Procypris merus*, *Ochetobius elongatus*, *Cyprinus carpio*, and *Onychostoma ovalis rhomboides* were the five species with the least effect on the stability of the community. These results suggested that the relative stability effects of each species can be used to predict key species and their roles in community construction. Examination of the stability effects of species in the community sequentially may also provide methods and means for the optimization of community species composition.

TAB. 4.1 – Simulation of the effects of different community species on community stability.

Fish	Absolute value of the average change rate of the community species (%)
<i>Hypophthalmichthys molitrix</i>	592.5
<i>Hypophthalmichthys nobilis</i>	360.0
<i>Puntius semifasciolatus</i>	322.0
<i>Misgurnus anguillicaudatus</i>	316.5
<i>Luciobrama macrocephalus</i>	307.4
<i>Discocheilus wui</i>	307.2
<i>Pelteobagrus vachellii</i>	291.8
<i>Rasbora steineri</i>	289.4
<i>Pelteobagrus fulvidraco</i>	284.3
<i>Lateolabrax japonicus</i>	280.1
<i>Rasborinus lineatus</i>	275.6
<i>Schizothorax meridionalis</i>	273.3
<i>Simocyclocheilus macrocephalus</i>	263.7
<i>Ctenopharyngodon idella</i>	255.3
<i>Simocyclocheilus yangzongensis</i>	255.2
<i>Elopichthys bambusa</i>	253.1
<i>Discogobio tetrabarbatus</i>	244.8
<i>Channa maculata</i>	242.1
<i>Acrossocheilus clivosius</i>	241.0
<i>Acheilognathus tonkinensis</i>	240.7

TAB. 4.1 – (continued).

Fish	Absolute value of the average change rate of the community species (%)
<i>Mylopharyngodon piceus</i>	234.9
<i>Sinocyclocheilus grahami tingi</i>	234.6
<i>Siniperca kneri</i>	234.5
<i>Xenocypris argentea</i>	232.5
<i>Leptobotia pellegrini</i>	229.8
<i>Pseudolaubuca sinensis</i>	229.2
<i>Acrossocheilus labiatus</i>	228.3
<i>Hemibagrus macropterus</i>	228.2
<i>Tor sinensis</i>	224.5
<i>Squalidus argentatus</i>	224.0
<i>Garra pingi pingi</i>	223.7
<i>Discogobiolongibarbus</i>	220.7
<i>Parasinilabeo assimilis</i>	219.4
<i>Takifugu ocellatus</i>	217.5
<i>Cyprinus multitaeniata</i>	215.5
<i>Sinilabeo discognathoides wui</i>	214.6
<i>Rhinogobius giurinus</i>	214.6
<i>Acrossocheilts elongatus</i>	209.2
<i>Spinibarbus denticulatus</i>	208.5
<i>Lucosoma chinensis</i>	207.5
<i>Cyprinus rabaudi</i>	205.8
<i>Carassius auratus auratus</i>	204.4
<i>Acrossocheilus beijiangensis</i>	204.3
<i>Sinocyclocheilus anophthalmus</i>	203.8
<i>Cyprinus yilongensis</i>	203.3
<i>Sinocyclocheilus macrolepis</i>	202.7
<i>Carassioides cantonensis</i>	202.6
<i>Garra pingi hainanensis</i>	198.4
<i>Carassius Auratus gibelio</i>	196.7
<i>Pseudogyriinocheilus prochilus</i>	196.2
<i>Onychostoma sima</i>	195.5
<i>Garra pingi yiliangensis</i>	193.7
<i>Sinocyclocheilus microphthalmus</i>	193.2
<i>Rectoris posehensis</i>	191.9
<i>Tachysurus argentivittatus</i>	191.2
<i>Pelteobagrus intermedius</i>	190.0
<i>Tor zonatus</i>	189.8
<i>Anabas testudineus</i>	189.7
<i>Parabramis pekinensis</i>	189.7
<i>Acrossocheilts hemispinus hemispinus</i>	189.0
<i>Osteochilus salsburyi</i>	187.7
<i>Hemiculter leucisculus</i>	185.9

TAB. 4.1 – (continued).

Fish	Absolute value of the average change rate of the community species (%)
<i>Onychostoma barbatulum</i>	184.2
<i>Cyprinus longzhouensis</i>	184.2
<i>Zacco platypus</i>	183.4
<i>Luciocyprinus langsoni</i>	179.7
<i>Cyprinus pellegrini</i>	178.8
<i>Onychostoma macrolepis</i>	178.1
<i>Sinibotia pulchra</i>	176.8
<i>Bangana decora</i>	176.0
<i>Leiocassis crassilabris</i>	175.9
<i>Pseudocrossocheilus bamaensis</i>	175.2
<i>Acrossocheilus fasciatus</i>	173.8
<i>Acrossocheilus paradoxus</i>	173.3
<i>Discogobio brachyphysallidos</i>	173.3
<i>Cirrhinus molitorella</i>	173.1
<i>Spinibarbus hollandi</i>	171.6
<i>Opsariichthys bidens</i>	170.2
<i>Semilabeo obscurus</i>	164.8
<i>Squaliobarbus curriculus</i>	163.9
<i>Ancherythroculter lini</i>	160.0
<i>Mystacoleucus marginatus</i>	158.4
<i>Ptychidio macrops</i>	156.2
<i>Sinilabeo rendahli</i>	154.0
<i>Hemibagrus guttatus</i>	153.9
<i>Cyprinus fuxianensis</i>	151.3
<i>Folifer brevifilis</i>	151.3
<i>Sinilabeo discognathoides N</i>	149.9
<i>Micronemacheilus pulcher</i>	148.9
<i>Spinibarbus sinensis</i>	148.4
<i>Semilabeo notabilis</i>	146.9
<i>Balitora kwangsiensis</i>	141.3
<i>Rhodeus ocellatus</i>	132.6
<i>Megalobrama skolkovii</i>	128.0
<i>Garra orientalis</i>	114.5
<i>Onychostoma lini</i>	110.7
<i>Acrossocheilus iridescens iridescens</i>	109.3
<i>Ptychidio jordani</i>	101.9
<i>Bostrichthys sinensis</i>	85.7
<i>Cyprinus carpio</i>	76.8
<i>Ochetobius elongatus</i>	70.2
<i>Onychostoma ovalis rhomboides</i>	54.6
<i>Procypris merus</i>	49.2
<i>Cyprinus chilia</i>	40.5

4.1.1 Species Substitutability

In the simulated community of 104 species of fish, the niche of the community of 103 species (without species “X”) changed after the removal of species “X.” The species substitutability index of species “X” to the community was reflected by the average value of various niche change rates. In some cases, the average niche change rate of the “104-X” community was high, which shows that “X” is very important to community stability and is irreplaceable. In other cases, the average niche change rate of the “104-X” community was low after the removal of species “X”, suggesting that the species in the community were not important (“X” substitutable). Based on the characteristics of the species composition of the simulated communities, the species substitutability indices for fish in the main-stream, middle reaches, and lower reaches were small, which indicates that species “X” is substitutable by other fish in the middle and lower reaches of the river ecosystem and that some species can fill in the niche vacated by species “X” after its removal. The high substitutability indices of fish species in the tributaries, middle reaches, and upper reaches showed that species “X” was not substitutable and that the fish in these communities do not complement one other. The results of this analysis were consistent with the general scientific principles of evolution in a riverine biological community: that is, from the ocean into rivers, fish first establish widely adaptive functional communities in estuaries and downstream, then expand upstream and into tributaries, which they evolve into functional-specific communities. From the perspective of systemic community function, the fish communities of the upstream areas and tributaries are simpler (function specialized to adapt to the specialized environment) and the species in the community are less replaceable as compared to the communities of downstream areas and the middle reaches.

In the analysis, the species loss of the community was reflected in the niche change rate of the community. The niche change rate varied from 3% to 2894.3% (see table 4.2), while the substitutability index of species “X” ranged from 1 to 964.8. The index value reflected the relationship between species “X” and other species in the community. Small index values indicated that the corresponding fish was more easily substituted. That is, other fish could fill the niche of species “X.”

4.1.2 Community Cohesion

The niches of the simulated community of 104 fish changed after the removal of species “X.” The average niche change rate of the observed species in each community (*i.e.*, the community of “104 species – species X”) was used to represent the effects of each specific species on community stability. Without species “X,” the average niche change rate of a given species in the n (103) “104-X” communities reflected the degree of association among species in the community. In highly cohesive communities, the contribution of a particular species to community formation is high, and *vice versa*.

TAB. 4.2 – Index of species substitutability in the simulated community.

Species (“X”)	Alternative indicators
<i>Hypophthalmichthys molitrix</i>	1.0
<i>Hypophthalmichthys nobilis</i>	1.3
<i>Ctenopharyngodon idella</i>	1.3
<i>Lateolabrax japonicus</i>	1.5
<i>Hemibagrus guttatus</i>	1.6
<i>Elopichthys bambusa</i>	1.9
<i>Pelteobagrus fulvidraco</i>	2.2
<i>Pelteobagrus vachellii</i>	2.2
<i>Onychostoma sima</i>	2.5
<i>Misgurnus anguillicaudatus</i>	3.3
<i>Sinocyclocheilus grahami tingi</i>	3.5
<i>Mylopharyngodon piceus</i>	3.6
<i>Squalidus argentatus</i>	4.1
<i>Semilabeo notabilis</i>	4.3
<i>Puntius semifasciolatus</i>	4.6
<i>Takifugu ocellatus</i>	4.8
<i>Channa maculata</i>	4.9
<i>Lucosoma chinensis</i>	5.4
<i>Acrossocheilus paradoxus</i>	5.6
<i>Rasbora steineri</i>	5.8
<i>Acrossocheilts hemispinus hemispinus</i>	6.0
<i>Anabas testudineus</i>	6.2
<i>Balitora kwangsiensis</i>	6.3
<i>Siniperca kneri</i>	6.4
<i>Cyprinus carpio</i>	6.5
<i>Tor sinensis</i>	7.0
<i>Squaliobarbus curriculus</i>	7.3
<i>Megalobrama skolkovii</i>	7.5
<i>Ptychidio jordani</i>	7.6
<i>Hemibagrus macropterus</i>	8.0
<i>Spinibarbus sinensis</i>	8.3
<i>Hemiculter leucisculus</i>	8.4
<i>Cyprinus rabaudi</i>	8.6
<i>Ptychidio macrops</i>	8.9
<i>Micronemacheilus pulcher</i>	9.0
<i>Spinibarbus denticulatus denticulatus</i>	9.0
<i>Leptobotia pellegrini</i>	9.9
<i>Acrossocheilus iridescens iridescens</i>	9.9
<i>Pseudolaubuca sinensis</i>	11.1
<i>Sinibotia pulchra</i>	11.4
<i>Cyprinus longzhouensis</i>	11.7
<i>Mystacoleucus marginatus</i>	12.6
<i>Osteochilus salsburyi</i>	13.8

TAB. 4.2 – (continued).

Species (“X”)	Alternative indicators
<i>Zacco platypus</i>	14.0
<i>Sinocyclocheilus microphthalmus</i>	14.2
<i>Discogobio tetrabarbatus</i>	14.4
<i>Acrossocheilus labiatus</i>	14.4
<i>Discogobiolongibarbatus</i>	14.4
<i>Pelteobagrus intermedius</i>	14.9
<i>Luciocyprinus langsoni</i>	14.9
<i>Ancherythroculter lini</i>	15.3
<i>Spinibarbus hollandi</i>	15.6
<i>Acrossocheilus fasciatus</i>	15.9
<i>Tachysurus argentivittatus</i>	17.3
<i>Rasborinus lineatus</i>	17.8
<i>Acrossocheilts elongatus</i>	18.9
<i>Rhinogobius giurinus</i>	19.1
<i>Semilabeo obscurus</i>	19.1
<i>Carassius Auratus gibelio</i>	19.1
<i>Discocheilus wui</i>	19.3
<i>Bangana decora</i>	19.5
<i>Bostrichthys sinensis</i>	19.8
<i>Discogobio brachyphysallidos</i>	19.8
<i>Procypris merus</i>	20.0
<i>Cirrhinus molitorella</i>	20.2
<i>Opsariichthys bidens</i>	20.6
<i>Cyprinus pellegrini</i>	20.8
<i>Garra pingi yiliangensis</i>	21.2
<i>Parasinilabeo assimilis</i>	22.2
<i>Sinocyclocheilus anophthalmus</i>	22.5
<i>Folifer brevifilis brevifilis</i>	23.1
<i>Rhodeus ocellatus</i>	23.6
<i>Onychostoma macrolepis</i>	26.7
<i>Garra pingi hainanensis</i>	26.7
<i>Cyprinus yilongensis</i>	27.3
<i>Parabramis pekinensis</i>	30.7
<i>Acrossocheilus beijiangensis</i>	37.9
<i>Acheilognathus tonkinensis</i>	41.1
<i>Pseudocrossocheilus bamaensis</i>	41.1
<i>Ochetobius elongatus</i>	41.7
<i>Sinocyclocheilus macrocephalus</i>	42.1
<i>Carassius auratus auratus</i>	42.5
<i>Tor zonatus</i>	46.9
<i>Leiocassis crassilabris</i>	48.2
<i>Schizothorax meridionalis</i>	49.1
<i>Cyprinus multitaeniata</i>	60.8

TAB. 4.2 – (continued).

Species (“X”)	Alternative indicators
<i>Cyprinus fuxianensis</i>	91.9
<i>Garra pingi pingi</i>	111.6
<i>Onychostoma lini</i>	142.6
<i>Acrossocheilus clivosius</i>	149.9
<i>Onychostoma barbatulum</i>	151.2
<i>Sinilabeo discognathoides wui</i>	171.5
<i>Pseudogyriinocheilus prochilus</i>	173.6
<i>Xenocypris argentea</i>	180.7
<i>Sinilabeo discognathoides</i>	194.1
<i>Rectoris posehensis</i>	199.7
<i>Luciobrama macrocephalus</i>	204.8
<i>Carassioides cantonensis</i>	237.8
<i>Sinilabeo rendahli</i>	266.7
<i>Onychostoma ovalis rhomboides</i>	268.8
<i>Sinocyclocheilus yangzongensis</i>	389.6
<i>Sinocyclocheilus macrolepis</i>	719.3
<i>Cyprinus chilia</i>	894.3
<i>Garra orientalis</i>	964.8

The treatment of common numbers in table 4.1 with an absolute value of 40.5%, showed that the cohesion of the simulated community was 1–15 (see table 4.3). The species-binding capacity reflects the effects of each species on niche composition and community stability. Higher species associations imply more stable community structures, suggesting that individual species make smaller contributions to the stability of the community structure.

4.1.3 Community Integration

The internal stability of the community structure is determined by species compatibility (or exclusion), and the compatibility between one fish species and other fish species can be determined using the model of morphological parameters established by us (see table 4.4). In the analysis of single species loss in the simulated community, qualitative increases and decreases in the niches of specific species were observed. If the niche of observed species (G) increased, this implied that G was in competition with the removed species. Next, the proportion of the competing species in the community of 103 simulated species (g) was calculated and treated according to the greatest common divisor in order to obtain the repulsion index of g in the simulated community. Species with large exclusion index values competed with the large inner niche of the community, whereas the species with small exclusion index values reflected the small mutual restrictions in the community to some extent.

TAB. 4.3 – Cohesion of the simulated community.

Specific species	Community cohesion
<i>Hypophthalmichthys molitrix</i>	14.8
<i>Hypophthalmichthys nobilis</i>	8.8
<i>Puntius semifasciolatus</i>	8.0
<i>Misgurnus anguillicaudatus</i>	7.9
<i>Luciobrama macrocephalus</i>	7.7
<i>Discocheilus wui</i>	7.6
<i>Pelteobagrus vachellii</i>	7.3
<i>Rasbora steineri</i>	7.2
<i>Pelteobagrus fulvidraco</i>	7.1
<i>Lateolabrax japonicus</i>	6.9
<i>Schizothorax meridionalis</i>	6.8
<i>Rasborinus lineatus</i>	6.8
<i>Sinocyclocheilus macrocephalus</i>	6.6
<i>Sinocyclocheilus yangzongensis</i>	6.4
<i>Elopichthys bambusa</i>	6.3
<i>Ctenopharyngodon idella</i>	6.3
<i>Discogobio tetrabarbatus</i>	6.1
<i>Channa maculata</i>	6.0
<i>Acheilognathus tonkinensis</i>	6.0
<i>Acrossocheilus clivosus</i>	6.0
<i>Siniperca kneri</i>	5.9
<i>Mylopharyngodon piceus</i>	5.9
<i>Sinocyclocheilus grahami tingi</i>	5.9
<i>Leptobotia pellegrini</i>	5.8
<i>Pseudolaubuca sinensis</i>	5.7
<i>Hemibagrus macropterus</i>	5.7
<i>Xenocypris argentea</i>	5.7
<i>Acrossocheilus labiatus</i>	5.7
<i>Tor sinensis</i>	5.6
<i>Squalidus argentatus</i>	5.6
<i>Discogobiolongibarbatus</i>	5.5
<i>Garra pingi pingi</i>	5.5
<i>Parasinilabeo assimilis</i>	5.5
<i>Takifugu ocellatus</i>	5.4
<i>Cyprinus multitaeniata</i>	5.4
<i>Rhinogobius giurinus</i>	5.4
<i>Sinilabeo discognathoides wui</i>	5.3
<i>Acrossocheilts elongatus</i>	5.2
<i>Spinibarbus denticulatus denticulatus</i>	5.2
<i>Lucosoma chinensis</i>	5.2
<i>Cyprinus rabaudi</i>	5.1
<i>Carassius auratus auratus</i>	5.1
<i>Acrossocheilus beijiangensis</i>	5.1

TAB. 4.3 – (continued).

Specific species	Community cohesion
<i>Cyprinus yilongensis</i>	5.1
<i>Sinocyclocheilus anophthalmus</i>	5.1
<i>Carassioides cantonensis</i>	5.1
<i>Sinocyclocheilus macrolepis</i>	5.1
<i>Garra pingi hainanensis</i>	5.0
<i>Carassius Auratus gibelio</i>	4.9
<i>Pseudogyrinocheilus prochilus</i>	4.9
<i>Onychostoma sima</i>	4.9
<i>Sinocyclocheilus microphthalmus</i>	4.8
<i>Garra pingi yiliangensis</i>	4.8
<i>Rectoris posehensis</i>	4.8
<i>Pelteobagrus intermedium</i>	4.8
<i>Tachysurus argentivittatus</i>	4.8
<i>Parabramis pekinensis</i>	4.7
<i>Anabas testudineus</i>	4.7
<i>Tor zonatus</i>	4.7
<i>Acrossocheilts hemispinus hemispinus</i>	4.7
<i>Osteochilus salsburyi</i>	4.7
<i>Hemiculter leucisculus</i>	4.7
<i>Cyprinus longzhouensis</i>	4.6
<i>Onychostoma barbatulum</i>	4.6
<i>Zacco platypus</i>	4.6
<i>Luciocyprinus langsoni</i>	4.5
<i>Cyprinus pellegrini</i>	4.5
<i>Onychostoma macrolepis</i>	4.4
<i>Bangana decora</i>	4.4
<i>Leiocassis crassilabris</i>	4.4
<i>Sinibotia pulchra</i>	4.4
<i>Pseudocrossocheilus bamaensis</i>	4.4
<i>Cirrhinus molitorella</i>	4.3
<i>Acrossocheilus paradoxus</i>	4.3
<i>Acrossocheilus fasciatus</i>	4.3
<i>Discogobio brachyphysallidos</i>	4.3
<i>Spinibarbus hollandi</i>	4.3
<i>Opsariichthys bidens</i>	4.3
<i>Squaliobarbus curriculus</i>	4.1
<i>Semilabeo obscurus</i>	4.1
<i>Ancherythroculter lini</i>	4.0
<i>Mystacoleucus marginatus</i>	4.0
<i>Ptychidio macrops</i>	3.9
<i>Semilabeo rendahli</i>	3.9
<i>Hemibagrus guttatus</i>	3.8
<i>Folifer brevifilis brevifilis</i>	3.8

TAB. 4.3 – (continued).

Specific species	Community cohesion
<i>Sinilabeo discognathoides</i>	3.8
<i>Cyprinus fuxianensis</i>	3.7
<i>Micronemacheilus pulcher</i>	3.7
<i>Semilabeo notabilis</i>	3.7
<i>Balitora kwangsiensis</i>	3.5
<i>Rhodeus ocellatus</i>	3.3
<i>Megalobrama skolkovii</i>	3.2
<i>Garra orientalis</i>	2.9
<i>Onychostoma lini</i>	2.8
<i>Acrossocheilus iridescens iridescens</i>	2.8
<i>Ptychidio jordani</i>	2.6
<i>Cyprinus carpio</i>	1.9
<i>Ochetobius elongatus</i>	1.7
<i>Onychostoma ovalis rhomboides</i>	1.4
<i>Procypris merus</i>	1.3
<i>Cyprinus chilia</i>	1.0

If the niche of the observed species (G) decreased after the removal of another species, then G and the removed species were in a mutually beneficial relationship. The proportion of mutually beneficial species out of the 103 remaining species was determined statistically and treated according to the greatest common divisor to obtain the compatibility index of G in the simulated community. Species with large compatibility index values reflected less competition in the inner niche of the community, whereas the species with small compatibility index values indicated a somewhat higher degree of competition in the community. The compatibility index may reflect the fusion degree of species in the community.

The stability of the community structure is determined by species compatibility, and the compatibility between one fish and all others can be obtained using the model of morphological parameters established by us. For example, *Sinocyclocheilus* sp. is closely related but highly morphologically differentiated, because each species inhabits limestone caves. Table 4.5 shows the results of the model analysis of six species of *Sinocyclocheilus*. In this table, “*” represents a mutual beneficial relationship, and “■” represents a competitive relationship. Using the percentage of mutually beneficial species as an index of compatibility, *Sinocyclocheilus microphthalmus* was the most compatible (86.4%) and *Sinocyclocheilus macrolepis* was the least compatible (1.9%). Although species in the genus *Sinocyclocheilus* are closely related to other species in same family, the compatibility results obtained by the model show that there are great differences among the species of *Sinocyclocheilus* within the community. The taxonomic properties of the morphological parameters, including the compatibility of species among communities, suggested that closely related fish might respond differently to the niche changes associated with the removal of a single species.

TAB. 4.4 – The indices of species exclusion and compatibility in the simulated communities.

Fish	Species exclusion index	Compatibility index
<i>Cyprinus carpio</i>	3.3	1.0
<i>Procypris merus</i>	3.0	1.3
<i>Spinibarbus hollandi</i>	2.7	1.8
<i>Pseudolaubuca sinensis</i>	2.6	1.9
<i>Sinilabeo discognathoides</i>	2.5	2.1
<i>Luciobrama macrocephalus</i>	2.3	2.3
<i>Bangana decora</i>	2.3	2.4
<i>Tachysurus argentivittatus</i>	2.2	2.5
<i>Sinocyclocheilus microphthalmus</i>	2.1	2.6
<i>Mylopharyngodon piceus</i>	2.1	2.6
<i>Ptychidio jordani</i>	2.1	2.6
<i>Ancherythroculter lini</i>	2.0	2.7
<i>Bostrichthys sinensis</i>	2.0	2.7
<i>Hemibagrus guttatus</i>	2.0	2.7
<i>Ctenopharyngodon idella</i>	2.0	2.7
<i>Ochetobius elongatus</i>	2.0	2.7
<i>Acheilognathus tonkinensis</i>	2.0	2.8
<i>Hypophthalmichthys nobilis</i>	1.9	2.8
<i>Megalobrama skolkovii</i>	1.9	2.8
<i>Elopichthys bambusa</i>	1.9	2.8
<i>Siniperca kneri</i>	1.9	2.8
<i>Opsariichthys bidens</i>	1.9	2.8
<i>Sinocyclocheilus grahami tingi</i>	1.9	2.8
<i>Cyprinus longzhouensis</i>	1.9	2.8
<i>Puntius semifasciolatus</i>	1.9	2.8
<i>Hypophthalmichthys molitrix</i>	1.9	2.9
<i>Sinocyclocheilus macrolepis</i>	1.9	2.9
<i>Onychostoma barbatulum</i>	1.9	2.9
<i>Acrossocheilus clivosius</i>	1.9	2.9
<i>Pseudocrossocheilus bamaensis</i>	1.9	2.9
<i>Spinibarbus denticulatus denticulatus</i>	1.9	2.9
<i>Carassius auratus auratus</i>	1.8	2.9
<i>Pelteobagrus fulvidraco</i>	1.8	2.9
<i>Parasinilabeo assimilis</i>	1.8	2.9
<i>Discogobio tetrabarbatus</i>	1.8	2.9
<i>Acrossocheilts elongatus</i>	1.8	2.9
<i>Discogobiolongibarbatus</i>	1.8	2.9
<i>Discocheilus wui</i>	1.8	2.9
<i>Rasbora steineri</i>	1.8	2.9
<i>Leiocassis crassilabris</i>	1.8	2.9
<i>Pelteobagrus vachellii</i>	1.8	2.9
<i>Squalidus argentatus</i>	1.8	3.0
<i>Schizothorax meridionalis</i>	1.8	3.0

TAB. 4.4 – (continued).

Fish	Species exclusion index	Compatibility index
<i>Acrossocheilus iridescens iridescens</i>	1.8	3.0
<i>Discogobio brachyphysallidos</i>	1.8	3.0
<i>Micronemacheilus pulcher</i>	1.8	3.0
<i>Sinibotia pulchra</i>	1.8	3.0
<i>Rhodeus ocellatus</i>	1.8	3.1
<i>Sinocyclocheilus macrocephalus</i>	1.8	3.1
<i>Sinocyclocheilus anophthalmus</i>	1.8	3.1
<i>Cyprinus yilongensis</i>	1.8	3.1
<i>Rectoris posehensis</i>	1.8	3.1
<i>Zacco platypus</i>	1.8	3.1
<i>Parabramis pekinensis</i>	1.7	3.1
<i>Rhinogobius giurinus</i>	1.7	3.1
<i>Sinocyclocheilus yangzongensis</i>	1.7	3.1
<i>Acrossocheilus fasciatus</i>	1.7	3.1
<i>Cyprinus pellegrini</i>	1.7	3.1
<i>Pelteobagrus intermedius</i>	1.7	3.1
<i>Takifugu ocellatus</i>	1.7	3.1
<i>Lateolabrax japonicus</i>	1.7	3.1
<i>Lucosoma chinensis</i>	1.7	3.2
<i>Acrossocheilus beijiangensis</i>	1.7	3.2
<i>Ptychidio macrops</i>	1.7	3.2
<i>Rasborinus lineatus</i>	1.7	3.2
<i>Balitora kwangsiensis</i>	1.7	3.2
<i>Anabas testudineus</i>	1.7	3.2
<i>Channa maculata</i>	1.7	3.2
<i>Tor sinensis</i>	1.7	3.2
<i>Hemibagrus macropterus</i>	1.7	3.2
<i>Carassioides cantonensis</i>	1.7	3.2
<i>Leptobotia pellegrini</i>	1.7	3.2
<i>Cirrhinus molitorella</i>	1.6	3.3
<i>Tor zonatus</i>	1.6	3.3
<i>Mystacoleucus marginatus</i>	1.6	3.3
<i>Semilabeo notabilis</i>	1.6	3.3
<i>Carassius Auratus gibelio</i>	1.6	3.3
<i>Garra pingi pingi</i>	1.6	3.3
<i>Xenocypris argentea</i>	1.6	3.3
<i>Misgurnus anguillicaudatus</i>	1.6	3.3
<i>Luciocyprinus langsoni</i>	1.6	3.3
<i>Cyprinus multitaeniata</i>	1.6	3.3
<i>Pseudogyrinocheilus prochilus</i>	1.5	3.4
<i>Osteochilus salsburyi</i>	1.5	3.4
<i>Hemiculter leucisculus</i>	1.5	3.4
<i>Sinilabeo discognathoides wui</i>	1.5	3.5

TAB. 4.4 – (continued).

Fish	Species exclusion index	Compatibility index
<i>Sinilabeo rendahli</i>	1.4	3.5
<i>Garra pingi hainanensis</i>	1.4	3.5
<i>Squaliobarbus curriculus</i>	1.4	3.6
<i>Cyprinus rabaudi</i>	1.4	3.6
<i>Acrossocheilus paradoxus</i>	1.4	3.6
<i>Onychostoma macrolepis</i>	1.3	3.6
<i>Onychostoma ovalis rhomboides</i>	1.3	3.6
<i>Acrossocheilus labiatus</i>	1.3	3.6
<i>Cyprinus fuxianensis</i>	1.3	3.7
<i>Garra orientalis</i>	1.3	3.7
<i>Garra pingi yiliangensis</i>	1.2	3.8
<i>Cyprinus chilia</i>	1.2	3.8
<i>Folifer brevifilis brevifilis</i>	1.2	3.8
<i>Acrossocheilts hemispinus hemispinus</i>	1.2	3.8
<i>Onychostoma lini</i>	1.2	3.8
<i>Spinibarbus sinensis</i>	1.2	3.8
<i>Semilabeo obscurus</i>	1.1	4.0
<i>Onychostoma sima</i>	1.0	4.1

4.1.4 Characteristics of the Geographic Distribution of the Ecological Niche Response

The 104 fish species in the simulated community were divided into two groups: the mainstream species and the tributary stream species. There were 47 mainstream species and 57 tributary-stream species (see table 4.6). The average change rates of the tributary-stream fish were larger than those of the mainstream fish, suggesting that the niche variability and degree of disturbance of the tributary-stream species were higher, and community stability was lower. To some extent, variations in fish community niches were larger in small-scale streams than in large-scale streams. The results of this analysis were consistent with the characteristics of fish communities in tributaries.

During the analysis of the niche changes associated with the removal of species from the simulated community, each species' rates of niche change in the "104-X" community were calculated, and a weighted average of each species' niche change rates after missing "X" species was expressed as integrated niche change rates. The fish species with small integrated niche rates in the communities reflect high stability in the community; the fish species with large integrated niche rates in the communities reflect low stability in the community.

Although the "primitive community" constructed by the model had some factors of "random" selection, this community was also selected based on the fish species recorded in the real-world Pearl River system. The average niche variations in the "subunit community" fish species in the upper, middle, and lower reaches of the river were analyzed. When the average niche change was small, the community was highly

TAB. 4.5 – Compatibility differences among fish in the genus *Sinocyclocheilus* within the community. (“*” indicates a mutually beneficial relationship, and “■” indicates a competitive relationship).

Fish	<i>Sinocyclocheilus grahami tingi</i>	<i>Sinocyclocheilus microphthalmus</i>	<i>Sinocyclocheilus macrolepis</i>	<i>Sinocyclocheilus anophthalmus</i>	<i>Sinocyclocheilus macrocephalus</i>	<i>Sinocyclocheilus yangzongensis</i>
<i>Sinocyclocheilus yangzongensis</i>	*	*	■	■	■	*
<i>Ptychidio macrops</i>	■	*	■	*	■	*
<i>Cirrhinus molitorella</i>	■	*	■	*	■	*
<i>Carassius auratus auratus</i>	■	*	■	*	■	*
<i>Cyprinus pellegrini</i>	■	■	■	*	■	*
<i>Acrossocheilus iridescens</i>	■	*	*	*	■	*
<i>Leiocassis crassilabris</i>	■	*	■	*	■	*
<i>Carassius Auratus</i>	■	■	■	*	■	*
<i>Hemibagrus macropterus</i>	■	*	■	*	■	*
<i>Megalobrama skolkovii</i>	■	*	■	*	■	*
<i>Tor sinensis</i>	■	*	■	*	■	*
<i>Mystacoleucus marginatus</i>	■	*	■	*	*	*
<i>Onychostoma barbatulum</i>	■	*	■	*	■	■
<i>Pseudocrossocheilus bamaensis</i>	■	*	■	*	■	■
<i>Osteochilus salsburyi</i>	■	*	■	*	■	■
<i>Carassioides cantonensis</i>	■	*	■	*	■	■
<i>Acrossocheilus fasciatus</i>	■	*	■	*	■	■
<i>Rhodeus ocellatus</i>	■	*	■	*	■	■
<i>Opsariichthys bidens</i>	■	*	■	*	■	■
<i>Bostrichthys sinensis</i>	■	■	■	*	■	■
<i>Zacco platypus</i>	■	*	■	*	■	■
<i>Siniperca kneri</i>	*	*	■	*	*	■
<i>Ptychidio jordani</i>	■	*	■	■	■	■
<i>Spinibarbus hollandi</i>	■	*	■	■	■	■
<i>Sinocyclocheilus microphthalmus</i>	■	*	■	■	■	■
<i>Procypris merus</i>	■	*	■	■	■	■
<i>Ancherythroculter lini</i>	*	*	■	■	*	■
<i>Pseudolaubuca sinensis</i>	■	*	■	■	■	■
<i>Luciobrama macrocephalus</i>	*	*	■	■	*	■
<i>Cyprinus carpio</i>	■	■	■	■	■	■
<i>Cyprinus longzhouensis</i>	■	■	■	*	■	■
<i>Tor zonatus</i>	■	*	■	*	■	■
<i>Acheilognathus tonkinensis</i>	■	*	■	*	*	■

TAB. 4.5 – (continued).

Fish	<i>Sinocyclocheilus grahami tingi</i>	<i>Sinocyclocheilus microphthalmus</i>	<i>Sinocyclocheilus macrolepis</i>	<i>Sinocyclocheilus anophthalmus</i>	<i>Sinocyclocheilus macrocephalus</i>	<i>Sinocyclocheilus yangzongensis</i>
<i>Tachysurus argentevittatus</i>	■	*	■	*	■	■
<i>Parasinilabeo assimilis</i>	■	*	■	*	■	■
<i>Rhinogobius giurinus</i>	■	*	■	*	■	■
<i>Pelteobagrus intermedius</i>	■	■	■	*	■	■
<i>Lucosoma chinensis</i>	■	*	■	*	■	■
<i>Rectoris posehensis</i>	■	*	■	*	■	■
<i>Sinilabeo rendahli</i>	■	■	■	*	■	■
<i>Garra pingi hainanensis</i>	■	■	■	*	■	■
<i>Discogobiolongibaratus</i>	■	*	■	■	■	■
<i>Semilabeo obscurus</i>	■	*	■	*	■	■
<i>Sinibotia pulchra</i>	■	■	■	*	■	■
<i>Sinocyclocheilus macrocephalus</i>	*	*	■	*	*	■
<i>Cyprinus multitaeniata</i>	■	*	■	*	■	■
<i>Cyprinus yilongensis</i>	*	*	■	■	■	■
<i>Luciocyprinus langsoni</i>	■	■	■	*	■	■
<i>Micronemacheilus pulcher</i>	■	■	■	*	■	■
<i>Acrossocheilus paradoxus</i>	■	*	■	*	■	■
<i>Acrossocheilus beijiangensis</i>	■	*	■	*	■	■
<i>Squalidus argentatus</i>	■	*	■	*	■	■
<i>Acrossocheilts hemispinus hemispinus</i>	■	*	■	*	■	■
<i>Garra orientalis</i>	■	*	■	*	■	■
<i>Schizothorax meridionalis</i>	*	*	■	*	■	■
<i>Spinibarbus denticulatus denticulatus</i>	■	*	■	*	■	■
<i>Onychostoma lini</i>	■	*	■	*	■	■
<i>Acrossocheilts elongatus</i>	■	*	■	*	■	■
<i>Onychostoma macrolepis</i>	■	*	■	*	■	■
<i>Pseudogyrinocheilus prochilus</i>	■	*	■	*	■	■
<i>Sinocyclocheilus macrolepis</i>	*	*	*	■	■	■
<i>Hemiculter leucisculus</i>	■	*	■	*	■	■
<i>Balitora kwangsiensis</i>	*	*	■	*	■	■
<i>Squaliobarbus curriculus</i>	■	*	■	*	■	■
<i>Cyprinus chilia</i>	*	*	■	*	■	■
<i>Parabramis pekinensis</i>	■	*	■	*	■	■
<i>Onychostoma ovalis rhomboides</i>	*	*	■	*	■	■
<i>Ochetobius elongatus</i>	*	*	■	■	■	■

TAB. 4.5 – (continued).

<i>Mylopharyngodon piceus</i>	*	*	■	*	*	■
<i>Elopichthys bambusa</i>	*	*	■	*	*	■
<i>Bangana decora</i>	■	*	■	■	■	■
<i>Discogobio brachyphysallidos</i>	■	*	■	*	■	■
<i>Hemibagrus guttatus</i>	*	■	*	*	■	■
<i>Sinocyclocheilus anophthalmus</i>	*	*	■	*	■	■
<i>Leptobotia pellegrini</i>	■	*	■	*	■	■
<i>Sinocyclocheilus grahami tingi</i>	*	*	■	*	■	■
<i>Discogobio tetrabarbatas</i>	■	*	■	*	■	■
<i>Anabas testudineus</i>	■	*	■	*	■	■
<i>Channa maculata</i>	■	*	■	*	■	■
<i>Cyprinus rabaudi</i>	■	*	■	*	■	■
<i>Spinibarbus sinensis</i>	■	*	■	*	■	■
<i>Folifer brevifilis brevifilis</i>	*	*	■	*	■	■
<i>Semilabeo notabilis</i>	*	*	■	*	■	■
<i>Similabeo discognathoides</i>	*	■	■	*	■	■
<i>Ctenopharyngodon idella</i>	*	*	■	*	■	■
<i>Takifugu ocellatus</i>	■	■	■	*	■	■
<i>Misgurnus anguillicaudatus</i>	■	*	■	*	■	■
<i>Garra pingi yiliangensis</i>	■	*	■	■	■	■
<i>Onychostoma sima</i>	■	*	■	*	■	■
<i>Cyprinus fuxianensis</i>	■	*	■	*	■	■
<i>Acrossocheilus clivosius</i>	*	*	■	*	■	■
<i>Rasbora steineri</i>	■	*	■	■	■	■
<i>Discocheilus wui</i>	*	*	■	*	■	■
<i>Acrossocheilus labiatus</i>	■	*	■	■	■	■
<i>Pelteobagrus vachellii</i>	*	*	■	■	■	■
<i>Similabeo discognathoides wui</i>	*	*	■	■	■	■
<i>Pelteobagrus fulvidraco</i>	*	*	■	■	■	■
<i>Puntius semifasciolatus</i>	■	*	■	■	■	■
<i>Lateolabrax japonicus</i>	*	*	■	■	*	■
<i>Garra pingi pingi</i>	*	*	■	■	■	■
<i>Hypophthalmichthys molitrix</i>	*	*	■	■	*	■
<i>Rasborinus lineatus Parabramis pekinensis</i>	■	*	■	■	■	■
<i>Xenocypris argentea</i>	*	*	■	*	■	■
<i>Hypophthalmichthys nobilis</i>	*	*	■	■	■	■
Compatibility (%)	30	86	3	76	10	12

TAB. 4.6 – Mean differences in the of niche change rates of various fish in the simulated communities.

Fish	The average change rate of the niche of the mainstream type fish (%)	Fish	The average change rate of the niche of the tributary fish (%)
<i>Hypophthalmichthys molitrix</i>	3.0	<i>Onychostoma sima</i>	7.4
<i>Hypophthalmichthys nobilis</i>	3.8	<i>Sinocyclocheilus grahami tingi</i>	10.5
<i>Ctenopharyngodon idella</i>	4.0	<i>Semilabeo notabilis</i>	12.8
<i>Lateolabrax japonicus</i>	4.5	<i>Acrossocheilus paradoxus</i>	16.7
<i>Hemibagrus guttatus</i>	4.7	<i>Rasbora steineri</i>	17.5
<i>Elopichthys bambusa</i>	5.7	<i>Acrossocheilts hemispinus hemispinus</i>	17.9
<i>Pelteobagrus fulvidraco</i>	6.6	<i>Balitora kwangsiensis</i>	18.8
<i>Pelteobagrus vachellii</i>	6.6	<i>Spinibarbus sinensis</i>	24.8
<i>Misgurnus anguillicaudatus</i>	10.0	<i>Cyprinus rabaudi</i>	25.8
<i>Mylopharyngodon piceus</i>	10.8	<i>Acrossocheilus iridescens iridescens</i>	29.7
<i>Squalidus argentatus</i>	12.3	<i>Cyprinus longzhouensis</i>	35.0
<i>Puntius semifasciolatus</i>	13.7	<i>Mystacoleucus marginatus</i>	37.8
<i>Takifugu ocellatus</i>	14.4	<i>Osteochilus salsburyi</i>	41.4
<i>Channa maculata</i>	14.6	<i>Discogobio tetrabarbatus</i>	43.1
<i>Lcucosoma chinensis</i>	16.2	<i>Acrossocheilus labiatus</i>	43.1
<i>Anabas testudineus</i>	18.6	<i>Discogobio longibarbatus</i>	43.3
<i>Siniperca kneri</i>	19.3	<i>Luciocyprinus langsoni</i>	44.8
<i>Cyprinus carpio Linnaeus, 1758</i>	19.6	<i>Sinocyclocheilus microphthalmus</i>	42.7
<i>Tor sinensis</i>	21.0	<i>Spinibarbus hollandi</i>	46.7
<i>Squaliobarbus curriculus</i>	22.0	<i>Acrossocheilus fasciatus</i>	47.7
<i>Megalobrama skolkovii</i>	22.4	<i>Acrossocheilts elongatus</i>	56.7
<i>Ptychidio jordani</i>	22.8	<i>Semilabeo obscurus</i>	57.3
<i>Hemibagrus macropterus</i>	24.0	<i>Discocheilus wui</i>	57.8
<i>Hemiculter leucisculus</i>	25.3	<i>Bangana decora</i>	58.5
<i>Ptychidio macrops</i>	26.6	<i>Discogobio brachyphysallidos</i>	59.4

TAB. 4.6 – (continued).

<i>Micronemacheilus pulcher</i>	27.0	<i>Procypris merus</i>	60.1
<i>Spinibarbus denticulatus denticulatus</i>	27.1	<i>Opsariichthys bidens</i>	61.8
<i>Leptobotia pellegrini</i>	29.6	<i>Cyprinus pellegrini</i>	62.5
<i>Pseudolaubuca sinensis</i>	33.4	<i>Garra pingi yiliangensis</i>	63.7
<i>Sinibotia pulchra</i>	34.3	<i>Parasinilabeo assimilis</i>	66.6
<i>Zacco platypus</i>	42.1	<i>Sinocyclocheilus anophthalmus</i>	67.4
<i>Pelteobagrus intermedius</i>	44.6	<i>Folifer brevifilis brevifilis</i>	69.4
<i>Ancherythroculter lini</i>	45.9	<i>Onychostoma macrolepis</i>	80.0
<i>Tachysurus argentivittatus</i>	51.8	<i>Garra pingi hainanensis</i>	80.2
<i>Rasborinus lineatus</i>	53.4	<i>Cyprinus yilongensis</i>	81.9
<i>Rhinogobius giurinus</i>	57.2	<i>Acrossocheilus beijiangensis</i>	113.6
<i>Carassius Auratus gibelio</i>	57.4	<i>Acheilognathus tonkinensis</i>	123.3
<i>Bostrichthys sinensis</i>	59.3	<i>Pseudocrossocheilus bamaensis</i>	123.4
<i>Cirrhinus molitorella</i>	60.7	<i>Sinocyclocheilus macrocephalus</i>	126.2
<i>Rhodeus ocellatus</i>	70.9	<i>Tor zonatus</i>	140.6
<i>Parabramis pekinensis</i>	92.0	<i>Leiocassis crassilabris</i>	144.7
<i>Ochetobius elongatus</i>	125.2	<i>Schizothorax meridionalis</i>	147.2
<i>Carassius auratus auratus</i>	127.4	<i>Cyprinus multitaeniata</i>	182.5
<i>Garra pingi pingi</i>	334.7	<i>Cyprinus fuzianensis</i>	275.7
<i>Xenocypris argentea</i>	542.2	<i>Onychostoma lini</i>	427.9
<i>Luciobrama macrocephalus</i>	614.3	<i>Acrossocheilus clivosius</i>	449.7
<i>Garra orientalis</i>	2894.3	<i>Onychostoma barbatulum</i>	453.7
		<i>Sinilabeo discognathoides wui</i>	514.4
		<i>Pseudogyriinocheilus prochilus</i>	520.8
		<i>Sinilabeo discognathoides</i>	582.3
		<i>Rectoris posehensis</i>	599.0
		<i>Carassioides cantonensis</i>	713.4
		<i>Sinilabeo rendahli</i>	800.1

TAB. 4.6 – (continued).

Fish	The average change rate of the niche of the mainstream type fish (%)	Fish	The average change rate of the niche of the tributary fish (%)
		<i>Onychostoma ovalis rhomboides</i>	806.5
		<i>Sinocyclocheilus yangzongensis</i>	1168.7
		<i>Sinocyclocheilus macrolepis</i>	2157.8
		<i>Cyprinus chilia</i>	2682.9
Average rate of change (%)	122.9	Average rate of change (%)	260.4

stable and variability was low. Conversely, when the average niche change was large, community stability was low and variability was high.

4.1.5 Differences

4.1.5.1 Species Size Types

Species vary in size. It is generally believed that large species occupy more niches and have a greater impact on the community, while small species occupy fewer niches and have no impact on the community. The 104 species of fish were divided into four classes based on individual size. The results showed that the species with larger individuals had a substantial influence on the composition of the community niche. The average change rate of the community niche was greatest for large species but was greater for small species than for medium species (see table 4.7). This suggested that the composition of the community niche is related to the functional composition. Small fish are more sensitive to environmental changes. In a changing environment, fish are in a highly variable state. Because the species niche pattern is still adapting to the evolution of the ecosystem, the positional relationships among species “blending” into the community are more indispensable. This phenomenon was to some extent consistent with the results of the community deconstruction model.

Therefore, the effects of fish loss on community niche change are not entirely determined by the species size. Further analysis showed that, during ecological deletion and the rearrangement of species niches, niche allocation was not based on the size of the species in the community, and species of the same size had different niches. Similar results were obtained using both the “primitive communities” and the “104-X” communities. *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, and *Elopichthys bambusa* are all the same size. The niches of these five fish exhibited “random” differences in response to fish removal from the simulated community, similar to *Megalobrama skolkovii* and *Parabramis pekinensis* (see table 4.8). This suggested that differences in the niche are not determined by species size, but by the implicit functional division of the community. This phenomenon was observed in large fish (see figure 4.1), medium fish (see figure 4.2), and small fish (see figure 4.3).

4.1.5.2 Affinities

The effects of fish with the same classification on the community niche differed. As shown in the graph below, the niche changes of different species of fish in the community were analyzed by selecting two closely related species of fish. In the model analysis of us, no similar niche effects were found among the cyprinids (seven species of *Cyprinus* and two species of *Procyprinus*; see figure 4.4) or the light-lipped fish (eight species; see figure 4.5). That is, fish with similar classification attributes differed with respect to the rate of niche change. Niche overlaps are common in multi-species communities, and the relationships among species in the food chain are a network. Close relatives of similar species or species with similar functional attributes exist simultaneously in the system, and different niche functions exist in

TAB. 4.7 – Average niche change rates and individual weight differences among community species.

Species ≥10 kg	Change rate (%)	Species ≥kg	Change rate (%)	Species ≥100 g	Change rate (%)	Species ≥g	Change rate (%)
<i>Hypophthalmichthys molitrix</i>	591	<i>Pelteobagrus vachellii</i>	291	<i>Pelteobagrus fulvidraco</i>	284	<i>Puntius semifasciolatus</i>	320
<i>Hypophthalmichthys nobilis</i>	353	<i>Channa maculata</i>	241	<i>Acrossocheilus clivosius</i>	238	<i>Misgurnus anguillicaudatus</i>	316
<i>Elopichthys bambusa</i>	251	<i>Lateolabrax japonicus</i>	275	<i>Acrossocheilus labiatus</i>	227	<i>Discocheilus wui</i>	305
<i>Ctenopharyngodon idella</i>	250	<i>Siniperca kneri</i>	236	<i>Tor sinensis</i>	225	<i>Rasbora steineri</i>	288
<i>Mylopharyngodon piceus</i>	235	<i>Hemibagrus macropterus</i>	228	<i>Takifugu ocellatus</i>	216	<i>Schizothorax meridionalis</i>	273
<i>Luciobrama macrocephalus</i>	307	<i>Xenocypris argentea</i>	228	<i>Cyprinus multitaeniata</i>	216	<i>Rasborinus lineatus</i>	271
		<i>Spinibarbus denticulatus</i>	208	<i>Sinilabeo discognathoides wui</i>	212	<i>Sinocyclocheilus macrocephalus</i>	264
		<i>Cyprinus rabaudi</i>	205	<i>Acrossocheilts elongatus</i>	209	<i>Sinocyclocheilus yangzongensis</i>	255
		<i>Cyprinus yilongensis</i>	203	<i>Carassius auratus auratus</i>	204	<i>Discogobio tetrabarbatus</i>	244
		<i>Pelteobagrus intermedius</i>	190	<i>Acrossocheilus beijiangensis</i>	204	<i>Acheilognathus tonkinensis</i>	240
		<i>Tor zonatus</i>	188	<i>Carassius auratus gibelio</i>	197	<i>Sinocyclocheilus grahami tingi</i>	234
		<i>Cyprinus longzhouensis</i>	184	<i>Onychostoma sima</i>	194	<i>Leptobotia pellegrini</i>	231

TAB. 4.7 – (continued).

<i>Luciocyprinus langsoni</i>	180	<i>Parabramis pekinensis</i>	189	<i>Pseudolaubuca sinensis</i>	229
<i>Cyprinus pellegrini</i>	179	<i>Acrossocheilts hemispinus</i>	188	<i>Squalidus argentatus</i>	223
<i>Cirrhinus molitorella</i>	173	<i>Onychostoma barbatulum</i>	184	<i>Discogobiolongibarbus</i>	220
<i>Spinibarbus hollandi</i>	171	<i>Onychostoma macrolepis</i>	177	<i>Garra pingi pingi</i>	220
<i>Squaliobarbus curriculus</i>	164	<i>Bangana decora</i>	177	<i>Parasinilabeo assimilis</i>	219
<i>Semilabeo obscurus</i>	163	<i>Leiocassis crassilabris</i>	176	<i>Rhinogobius giurinus</i>	216
<i>Ancherythroculter lini</i>	160	<i>Acrossocheilus paradoxus</i>	173	<i>Lcucosoma chinensis</i>	206
<i>Ptychidio macrops</i>	156	<i>Acrossocheilus fasciatus</i>	173	<i>Sinocyclocheilus anophthalmus</i>	203
<i>Hemibagrus guttatus</i>	153	<i>Semilabeo notabilis</i>	147	<i>Carassioides cantonensis</i>	202
<i>Folifer brevifilis brevifilis</i>	150	<i>Onychostoma lini</i>	110	<i>Sinocyclocheilus macrolepis</i>	202
<i>Sinilabeo discognathoides</i>	150	<i>Acrossocheilus iridescens</i>	110	<i>Garra pingi hainanensis</i>	198
<i>Cyprinus fuxianensis</i>	149	<i>Ptychidio jordani</i>	102	<i>Pseudogyriinocheilus prochilus</i>	196
<i>Megalobrama skolkovii</i>	128	<i>Onychostoma ovalis</i>	54	<i>Sinocyclocheilus microphthalmus</i>	193
		<i>rhomboides</i>			

TAB. 4.7 – (continued).

Species ≥ 10 kg	Change rate (%)	Species ≥kg	Change rate (%)	Species ≥100 g	Change rate (%)	Species ≥g	Change rate (%)
		<i>Procypris merus</i>	50			<i>Garra pingi yiliangensis</i>	192
		<i>Cyprinus chilia</i>	40			<i>Rectoris posehensis</i>	192
		<i>Cyprinus carpio</i>	77			<i>Tachysurus argentivittatus</i>	190
		<i>Ochetobius elongatus</i>	69			<i>Anabas testudineus</i>	189
						<i>Osteochilus salsburyi</i>	187
						<i>Hemiculter leucisculus</i>	186
						<i>Zacco platypus</i>	183
						<i>Sinibotia pulchra</i>	176
						<i>Pseudocrossocheilus bamaensis</i>	175
						<i>Discogobio brachyphysallidos</i>	173
						<i>Opsariichthys bidens</i>	170
						<i>Mystacoleucus marginatus</i>	158
						<i>Sinilabeo rendahli</i>	154
						<i>Micronemacheilus pulcher</i>	148
						<i>Balitora kwangsiensis</i>	140
						<i>Rhodeus ocellatus</i>	132
						<i>Garra orientalis</i>	114
Community average niche change rate (%)	331		172		183		210

TAB. 4.8 – Community niches were not assigned proportional to species size (%).

Fish	<i>Mylopharyngodon piceus</i>	<i>Ctenopharyngodon idella</i>	<i>Hypophthalmichthys molitrix</i>	<i>Hypophthalmichthys nobilis</i>	<i>Elopichthys bambusa</i>	<i>Megalobrama skolkovii</i>	<i>Parabramis pekinensis</i>
<i>Mylopharyngodon piceus</i>	1.790	1.812	13.055	4.926	1.616	1.745	1.774
<i>Ctenopharyngodon idella</i>	1.346	1.508	1.538	1.885	1.174	1.311	1.335
<i>Hypophthalmichthys molitrix</i>	10.647	10.711	10.708	10.209	10.783	10.377	10.526
<i>Hypophthalmichthys nobilis</i>	3.551	3.737	3.594	0.070	3.742	3.468	3.515
<i>Elopichthys bambusa</i>	1.513	1.133	1.324	4.737	1.547	1.474	1.502
<i>Megalobrama skolkovii</i>	1.045	0.641	0.834	0.605	1.394	0.685	1.034
<i>Parabramis pekinensis</i>	1.560	0.954	0.911	0.331	1.537	0.684	1.979

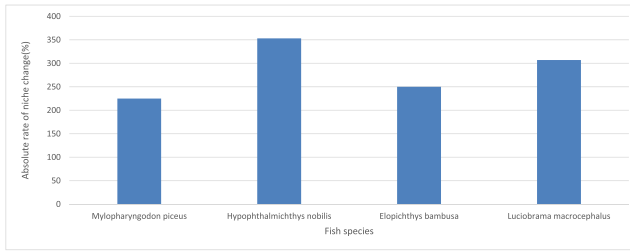


FIG. 4.1 – Variations in the ecological niches of the large fish in simulated communities.

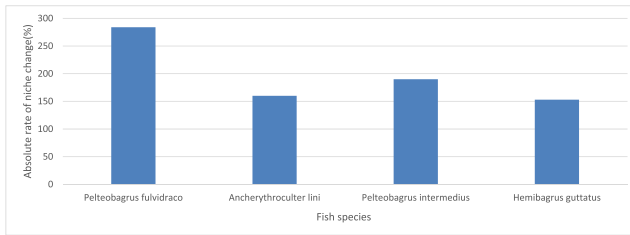


FIG. 4.2 – Variations in the ecological niches of the medium fish in simulated communities.

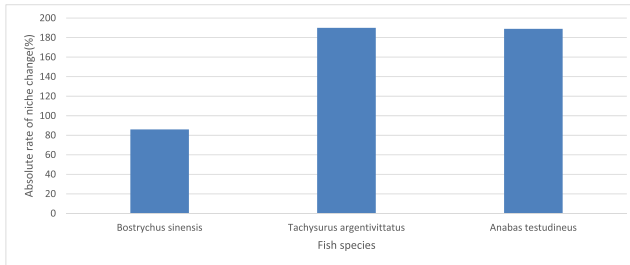


FIG. 4.3 – Variations in the ecological niches of the small fish in simulated communities.

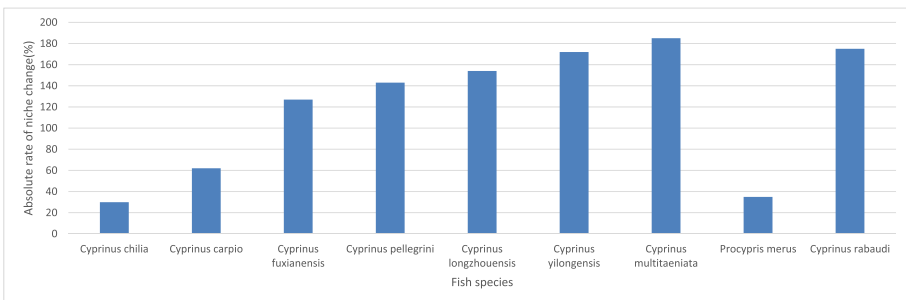


FIG. 4.4 – Variations in ecological niches of the fish similar to *Cyprinus* in simulated communities.

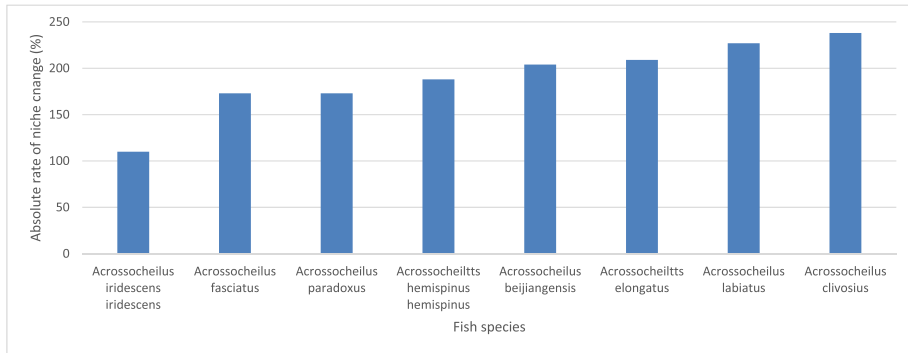


FIG. 4.5 – Variations in ecological niches of the *Acrossocheilus* in simulated communities.

different communities. The different functional division of species in different communities reflects their different effects on the communities. In real-world regional waters, fish of the same origin and function rarely appear together, and the differences identified by the model analysis also suggested that fish of the same origin and function inhabit different communities in the water and play a variety of distinct functional roles.

4.1.6 Feeding Habits

4.1.6.1 Carnivorous Fish

Community species compositions remain stable due to optimal acquired energy states. Under competitive pressure from other fish, the ecological niche of a species cannot be preserved. There are many competitive species in the community and great competitive pressure, so the proportion of competitive species to the greater community represents the competitive pressure. Competitive pressure can characterize the stability of a certain species in a community. In communities, predatory fish compete with one another. Thus, compatible community species are required for the maintenance of a stable niche. *Elopichthys bambusa*, *Lateolabrax japonicus*, *Mylopharyngodon piceus*, *Hypophthalmichthys nobilis*, *Ancherythroculter lin*, *Siniperca kneri*, and *Luciobrama macrocephalus* are all carnivorous fish. The model compatibility analysis showed that the species competing with *Luciobrama macrocephalus* reached 83%, while *Elopichthys bambusa* experienced only 8% of the competitive pressure (see table 4.9). High competitive pressure indicates a difficult niche in a community, or that niche maintenance may be greatly affected by community succession.

In the simulated community, carnivorous fish had various mutually beneficial relationships with other fish, including *Elopichthys bambusa*, 92%; *Lateolabrax japonicus*, 90%; *Mylopharyngodon piceus*, 88%; *Hypophthalmichthys nobilis*, 83%; *Ancherythroculter lini*, 66%; *Siniperca kneri*, 63%; and *Luciobrama macrocephalus*, only 17%. The visible niche of *Luciobrama macrocephalus* was least stable in the simulated community, suggesting that this species was more likely to be lost than the other six fish species. Figure 4.6 shows the response values of community niche

TAB. 4.9 – Differences in the responses of several carnivorous fish species responded to changes in removed niches.*

Fish	<i>Luciobrama macrocephalus</i>	<i>Elopichthys bambusa</i>	<i>Hypophthalmichthys nobilis</i>	<i>Ancherythroculter lini</i>	<i>Lateolabrax japonicus</i>	<i>Mylopharyngodon piceus</i>	<i>Siniperca kneri</i>
<i>Siniperca kneri</i>	39.2	-0.7	-0.2	-1.0	-0.8	-0.5	-100.0
<i>Hypophthalmichthys molitrix</i>	1583.5	213.0	-98.0	-51.9	-7.0	629.3	-34.5
<i>Puntius semifasciolatus</i>	1327.9	-7.8	-7.7	185.8	-7.7	-7.7	-22.5
<i>Lateolabrax japonicus</i>	1100.5	-3.0	-3.8	-26.1	-100.0	-3.1	-22.5
<i>Xenocypris argentea</i>	1258.3	-3.1	-2.9	-25.4	-2.8	-3.0	-21.7
<i>Cyprinus fuxianensis</i>	1408.3	-2.6	-2.6	-25.2	-2.5	-2.5	-21.5
<i>Garra orientalis</i>	824.2	-2.3	-2.2	-25.0	-2.2	-2.3	-21.3
<i>Garra pingi pingi</i>	1122.9	-14.3	3.2	-24.7	-16.8	-12.3	-21.0
<i>Rasbora lineatus</i>	1356.2	-6.2	-3.1	59.0	-5.9	-6.0	-21.0
<i>Parabramis pekinensis</i>	1125.1	-1.8	-1.6	-24.6	-16.6	-1.7	-20.9
<i>Folifer brevifilis brevifilis</i>	793.1	-4.5	-5.6	72.8	-5.5	-5.2	-20.7
<i>Mylopharyngodon piceus</i>	572.3	-25.1	5.2	54.4	-0.1	-100.0	-20.7
<i>Acrossocheilus clivosius</i>	979.9	-14.4	3.8	-24.4	-16.5	-12.1	-20.7
<i>Onychostoma ovalis rhomboides</i>	539.0	-1.0	-0.8	-24.1	-1.0	-1.0	-20.3
<i>Cyprinus chilia</i>	538.5	-1.1	-0.9	-24.1	-1.0	-1.0	-20.3
<i>Elopichthys bambusa</i>	-42.7	-100.0	5.4	-1.9	0.8	-9.7	-20.0
<i>Discocheilus wui</i>	1351.3	-5.0	-4.7	195.1	-4.7	-4.8	-20.0
<i>Ochetobius elongatus</i>	842.9	-0.3	-0.1	-23.6	-0.3	-0.2	-19.8
<i>Mystacoleucus marginatus</i>	1254.3	-2.8	-2.6	100.3	-2.9	-2.8	-18.7
<i>Cyprinus carpio</i>	-52.6	1.5	1.8	-22.3	1.3	1.5	-18.5
<i>Rhinogobius giurinus</i>	1071.3	-6.1	-6.0	-6.0	-6.0	-6.0	-6.0
<i>Takifugu ocellatus</i>	929.7	-5.4	-6.1	93.8	-6.0	-5.8	-6.0
<i>Parasinilabeo assimilis</i>	791.5	-5.9	-5.7	-5.6	-5.7	-5.8	-5.6
<i>Pelteobagrus intermedius</i>	795.5	-4.6	-2.5	-5.2	-5.2	-5.0	-5.2
<i>Rectoris posehensis</i>	511.0	-5.4	-5.2	-5.1	-5.1	-5.3	-5.1
<i>Anabas testudineus</i>	1369.9	-4.5	-5.2	-4.9	-5.0	-4.8	-4.9
<i>Channa maculata</i>	1227.1	-5.0	-5.0	96.0	-5.0	-4.9	-4.9
<i>Discogobio longibarbus</i>	370.8	-5.0	-4.7	-4.7	-4.7	-4.8	-4.7
<i>Squalidus argentatus</i>	229.9	-4.5	-4.2	-4.5	-4.4	-4.4	-4.4
<i>Tachysurus argentivittatus</i>	1608.1	4.3	-12.5	31.8	6.6	2.2	-4.3
<i>Discogobio brachyphysallidos</i>	230.0	-4.6	-4.1	-4.3	-4.3	-4.4	-4.3

TAB. 4.9 – (continued).

<i>Garra pingi hainanensis</i>	948.2	-4.6	-4.4	-4.3	-4.3	-4.5	-4.3
<i>Cyprinus rabaudi</i>	516.6	-4.0	-4.3	-4.2	-4.2	-4.1	-4.2
<i>Acrossocheilus beijiangensis</i>	230.5	-4.3	-4.0	-4.1	-4.1	-4.1	-4.1
<i>Sinibotia pulchra</i>	229.6	-3.6	-4.1	97.9	-4.1	-3.9	-4.0
<i>Cyprinus multitaeniata</i>	373.9	-3.8	-4.1	-4.0	-4.1	-3.9	-4.0
<i>Micronemacheilus pulcher</i>	520.0	-3.3	-3.8	98.3	-3.9	-3.7	-3.9
<i>Similabeo rendahli</i>	663.2	-4.2	-4.0	98.2	-3.9	-4.1	-3.8
<i>Semilabeo obscurus</i>	1243.4	-3.9	-3.9	-3.7	-3.8	-3.9	-3.8
<i>Garra pingi yiliangensis</i>	1391.1	-4.0	-3.8	-3.5	-3.6	-3.9	-3.6
<i>Onychostoma sima</i>	1248.1	-3.6	-3.5	-3.4	-3.5	-3.6	-3.4
<i>Cyprinus longzhouensis</i>	234.7	-2.9	-2.9	99.9	-3.1	-3.0	-3.2
<i>Acrossocheilus labiatus</i>	1251.5	-3.3	-3.2	-3.2	-3.2	-3.2	-3.2
<i>Acrossocheilus paradoxus</i>	524.0	-3.2	-3.2	-3.1	-3.1	-3.1	-3.1
<i>Spinibarbus sinensis</i>	1401.4	-2.9	-3.0	-2.9	-2.9	-2.9	-2.9
<i>Megalobrama skolkovii</i>	237.0	-2.6	-2.3	-2.8	-2.7	-2.5	-2.8
<i>Acrossocheilts hemispinus hemispinus</i>	966.8	-2.7	-2.6	-2.6	-2.6	-2.6	-2.6
<i>Pseudogyrinocheilus prochilus</i>	677.8	-2.4	-2.0	-2.2	-2.2	-2.3	-2.2
<i>Tor zonatus</i>	238.3	-2.3	-1.8	-2.1	-2.0	-2.1	-2.1
<i>Onychostoma barbatulum</i>	238.7	-2.4	-1.7	-2.1	-2.0	-2.2	-2.0
<i>Hemiculter leucisculus</i>	532.6	-2.0	-1.8	-2.0	-2.0	-1.9	-2.0
<i>Onychostoma macrolepis</i>	237.5	-2.1	-1.9	-2.0	-2.0	-2.0	-2.0
<i>Pseudocrossocheilus bamaensis</i>	92.0	-2.2	1.2	-2.0	-1.9	-2.0	-1.9
<i>Onychostoma lini</i>	532.5	-1.9	-1.8	-13.3	-1.9	-1.9	-1.8
<i>Osteochilus salsburyi</i>	92.5	-2.1	-1.3	-13.2	-1.6	-1.8	-1.7
<i>Acrossocheilus fasciatus</i>	-54.0	-1.5	-1.1	-1.6	-1.5	-1.4	-1.5
<i>Opsariichthys bidens</i>	-53.9	-1.2	-0.8	-1.3	-1.2	-1.1	-1.3
<i>Rhodeus ocellatus</i>	-53.6	-1.4	-0.6	-1.2	-1.1	-1.1	-1.1
<i>Luciocyprinus langsoni</i>	232.4	-3.0	-3.2	-3.1	-3.2	-2.8	-0.9
<i>Zacco platypus</i>	-53.7	-0.8	-0.5	-1.0	-0.8	-0.7	-0.9
<i>Cyprinus pellegrini</i>	-54.6	-3.3	-2.4	100.5	-2.9	-3.1	-0.7

TAB. 4.9 – (continued).

Fish	<i>Luciobrama macrocephalus</i>	<i>Elopichthys bambusa</i>	<i>Hypophthalmichthys nobilis</i>	<i>Ancherythroculter lini</i>	<i>Lateolabrax japonicus</i>	<i>Mylopharyngodon piceus</i>	<i>Siniperca kneri</i>
<i>Ptychidio jordani</i>	-52.5	-0.2	0.2	-0.5	-0.3	-0.1	-0.4
<i>Sinocyclocheilus microphthalmus</i>	-53.1	-0.2	3.1	-0.2	-0.1	-0.0	-0.2
<i>Carassius Auratus gibelio</i>	91.1	-2.7	-2.0	101.5	-2.4	-2.5	-0.1
<i>Procypris merus</i>	-51.4	0.1	0.6	-0.2	-0.0	0.2	-0.1
<i>Pseudolaubuca sinensis</i>	-52.0	0.2	0.7	95.2	0.2	0.3	0.1
<i>Hemibagrus macropterus</i>	238.6	-1.2	-1.8	-2.1	-2.0	-1.6	0.2
<i>Parabramis pekinensis</i>	241.0	-0.8	-1.0	-24.3	-1.2	-0.9	1.0
<i>Carassioides cantonensis</i>	-53.7	-1.5	-0.4	104.5	-0.9	-1.2	1.3
<i>Luciobrama macrocephalus</i>	-100.0	0.7	0.7	210.9	0.2	0.6	2.4
<i>Pelteobagrus vachellii</i>	1972.4	-7.3	-2.6	179.1	-17.3	-2.0	19.1
<i>Similabeo discognathoides wui</i>	822.0	-2.8	-2.5	78.4	-9.9	-2.7	19.1
<i>Pelteobagrus fulvidraco</i>	1836.0	-0.7	-2.0	180.7	-16.8	-1.4	19.8
<i>Semilabeo notabilis</i>	541.0	-12.7	-0.5	-23.8	-15.7	-0.5	21.4
<i>Similabeo discognathoides</i>	544.4	-12.5	5.3	82.9	-15.3	-10.6	22.1
<i>Bangana decora</i>	554.6	-11.8	1.8	85.6	-14.1	0.9	23.8
<i>Ctenopharyngodon idella</i>	293.3	-12.5	1.2	18.7	16.3	1.2	31.3
<i>Sinocyclocheilus macrocephalus</i>	1504.2	-4.2	-4.1	-4.1	-4.1	-4.1	35.8
<i>Spinibarbus denticulatus denticulatus</i>	232.8	-3.1	-3.3	-3.5	-3.5	-3.2	36.8
<i>Schizothorax meridionalis</i>	233.1	-3.4	-3.2	-3.4	-3.3	-3.3	37.0
<i>Cirrhinus molitorella</i>	-54.6	-3.3	-2.8	99.6	-3.3	-3.3	37.1
<i>Acrossocheilus iridescens iridescens</i>	-54.8	-2.9	-2.7	-3.2	-3.1	-2.9	37.4
<i>Hemibagrus guttatus</i>	657.6	-1.4	-7.4	190.2	13.3	8.0	37.8
<i>Carassius auratus auratus</i>	237.7	-2.5	-2.2	100.8	-2.7	-2.6	37.9
<i>Leiocassis crassilabris</i>	238.2	-1.6	0.7	-2.6	-2.4	-2.1	38.3
<i>Tor sinensis</i>	238.8	-1.7	-1.8	101.9	-2.2	-1.9	38.6
<i>Sinocyclocheilus macrolepis</i>	1101.0	-2.1	-1.8	192.4	-1.9	-1.9	39.0

TAB. 4.9 – (continued).

<i>Leptobotia pellegrini</i>	238.8	-1.1	4.0	102.5	-1.9	-1.5	39.1
<i>Sinocyclocheilus yangzongensis</i>	1102.9	-1.8	-1.6	-1.7	-1.7	-1.7	39.3
<i>Squaliobarbus curriculus</i>	239.5	-1.4	-1.4	-13.1	-1.6	-1.4	39.4
<i>Sinocyclocheilus grahami tingi</i>	94.9	-0.8	-0.4	207.6	-0.8	-0.6	40.6
<i>Sinocyclocheilus anophthalmus</i>	522.3	-0.9	-0.4	207.7	-0.7	-0.7	40.7
<i>Bostrichthys sinensis</i>	-53.4	-0.6	-0.1	-0.7	-0.6	-0.5	40.9
<i>Spinibarbus hollandi</i>	-52.4	0.6	3.4	-0.1	0.1	0.4	41.9
<i>Balitora kwangsiensis</i>	976.9	-1.5	-1.5	-24.7	-1.7	-1.6	50.9
<i>Discogobio tetrabarbatus</i>	1214.0	-6.3	-6.0	-5.8	-5.9	-6.1	62.8
<i>Rasbora steineri</i>	1221.8	-5.3	-5.3	-5.3	-5.3	-5.3	63.8
<i>Misgurnus anguillicaudatus</i>	804.7	-3.2	-4.4	-4.2	-4.2	-3.8	65.6
<i>Acheilognathus tonkinensis</i>	229.9	-4.5	-4.1	196.6	-4.2	-4.3	65.7
<i>Acrossocheilitts elongatus</i>	234.9	-3.2	-2.7	200.7	-3.0	-3.0	68.0
<i>Ptychidio macrops</i>	382.7	-2.6	-2.3	-2.6	-2.6	-2.5	68.7
<i>Ancherythroculter lini</i>	-51.4	-0.5	0.4	-100.0	-0.2	-0.2	72.8
<i>Hypophthalmichthys nobilis</i>	1257.6	2.2	-100.0	-26.2	29.0	175.2	108.8
<i>Cyprinus yilongensis</i>	-12.4	-4.6	-4.4	20.6	247.8	1181.9	1911.6
Competitive fish species	85	8	18	35	10	12	39
Competitive pressure %	83	8	17	34	10	12	38

*Note: niche reduction %.

change rates after the loss of different carnivorous fish. Community species composition and diversity change are the basis of the formation of community functional diversity, which is a comprehensive embodiment of the ability to adapt to environmental changes.

4.1.6.2 *Phytophagous Fish*

In the simulated community, the mutually beneficial relationships between herbivorous fish and other fish differed. For example, *Ctenopharyngodon idella* was 91.3%, *Hypophthalmichthys molitrix* was 90.3%, *Megalobrama skolkovii* was 90.3%, and *Spinibarbus denticulatus denticulatus* was 85.4%, but *Parabramis pekinensis* was only 2.9%. Figure 4.7 shows that the community species relationships among the bream and the other four fish were not in the same dimension. Thus, the niche performance of *Parabramis pekinensis* was obviously restricted by other types of communities. If other species were removed (eliminated), the niche of *Parabramis pekinensis* would be increased greatly, and the other four phytophagous fish were highly compatible. Phytophagous fish use the primary productivity of the water system to obtain energy and had an overall lower rate of niche change in the simulated community than carnivorous fish. This might indicate that the energy competition pressure among these fish is lower than that among carnivorous fish. More precisely, phytophagous food sources are more widely available than carnivorous food sources. Figure 4.8 shows that five species of fish with the same phytophagic nature are missing the same fish, reflecting the difference in the rate of ecological niche change.

4.1.6.3 *Omnivorous Fish*

There are much omnivorous fish in rivers, such as *Procypris merus*, *Cyprinus longzhouensis*, *Cyprinus pellegrini*, *Cyprinus yilongensis*, *Cyprinus carpio*, *Cyprinus rabaudi*, *Cyprinus multitaeniata*, *Cyprinus fuxianensis*, and *Cyprinus chilia*. Community compatibility analysis showed that the niches of these species varied greatly in response to community change (see figure 4.9). *Cyprinus chilia* was completely incompatible with other species, while *Procypris merus* was 100% compatible with other community species (see table 4.10). Thus, one type of niche was relatively stable, containing fish such as *Cyprinus yilongensis*, *Cyprinus carpio*, and *Cyprinus rabaudi*, while the other type of niche was highly unstable, containing such fish as *Procypris merus*, *Cyprinus longzhouensis*, *Cyprinus pellegrini*, *Cyprinus multitaeniata*, *Cyprinus fuxianensis*, and *Cyprinus chilia*. Across these species, the niche of *Cyprinus carpio* was the most stable, and that of *Cyprinus chilia* was the most unstable.

4.2 “Subunit Community” Succession

Two species can be competitive or symbiotic depending on their mutual interests. Species relationships can be parasitic, partially symbiotic, or mutually beneficial. The longer a community evolves, the more favorable and stable its environment, and

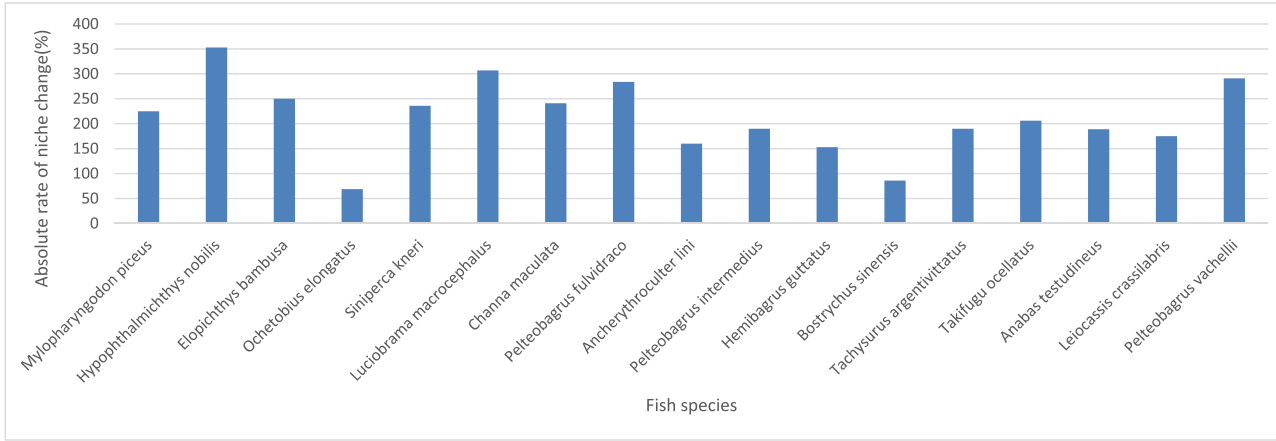


FIG. 4.6 – Variations in ecological niches of several carnivorous fish in simulated communities.

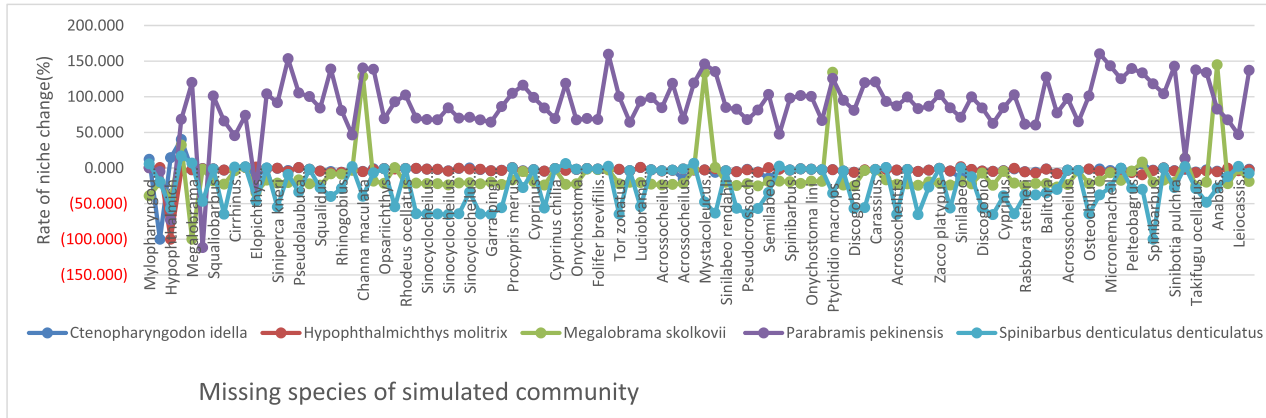


FIG. 4.7 – Differences in the responses of five phytophagous fish to the removal of various species from the community.

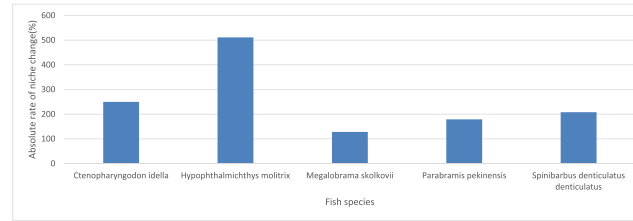


FIG. 4.8 – Variations in the ecological niches of several phytophagous fish in simulated communities.

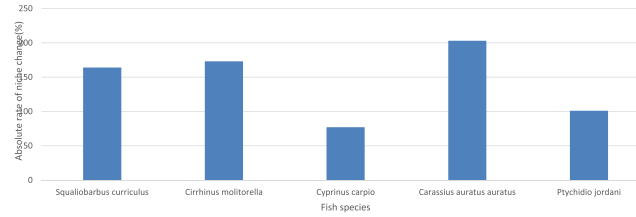


FIG. 4.9 – Variations in ecological niches of several omnivorous fish in simulated communities.

TAB. 4.10 – Niche differences among the *Cyprinus* in the community.

Fish	<i>Procypris merus</i>	<i>Cyprinus longzhouensis</i>	<i>Cyprinus pellegrini</i>	<i>Cyprinus yilongensis</i>	<i>Cyprinus carpio</i>	<i>Cyprinus rabaudi</i>	<i>Cyprinus multitaeniata</i>	<i>Cyprinus fuxianensis</i>	<i>Cyprinus chilia</i>
Consistency (%)	100.0	97.1	83.5	67.0	31.1	29.1	21.4	1.0	0
Niche average rate (%)	- 60.5	- 35.6	- 34.9	14.7	2.0	21.0	178.8	272.1	2656.1

the more species it contains. If two species utilize the same resources (niche overlap), they must compete and one species will be excluded. However, if the resource demands of one of the species change (niche differentiation), the two species may coexist. The trend in biological community development is toward niche differentiation and species increase. The study of the relationships among biological communities needs to go beyond the boundaries of specific taxa or taxon combinations (Bronstein, 1994). Morphological characters can reflect environmental characteristics and can be used to help understand the mechanisms underlying fish community composition in an ecosystem. By modeling community species, we can identify similar species types to better understand the diversity of species with the same functions, as well as the function-buffering ability of the ecosystem.

Although the “primitive community” constructed by the model had some factors of “random” selection, factors were also selected based on fish species recorded in the real-world Pearl River system. Because of the complexity of the river system and the diversity of the river habitats, the distributions of fish in the upper, middle, and lower reaches of the river differ noticeably. Such small communities correspond to small areas and may reflect characteristics that are more appropriate to a particular environment. In this section, we considered 25 dominant species in the middle and lower reaches of the Pearl River the “subunit community” in the simulated community of 104 species of fish (the “primitive community”; table 4.11, all fish) for the model analysis of niche succession by surplus species. The following table shows the species that have evolved into a “subunit community” (the italics in the table indicate hypothetical “subunit community” species).

The order in which the species were removed from the simulated community, was *Sinocyclocheilus macrolepis*, *Sinocyclocheilus macrocephalus*, *Sinocyclocheilus grahami tingi*, and *Pelteobagrus vachellii*. The niches of some species in the “subunit community” fluctuated up and down as the fish were eliminated one by one. If the niche value of the species in the “subunit community” was higher than that of the species before the loss, the removed fish is the competition object of the observed species in the “subunit community.” From this, we concluded that the relationship between the “removed fish” and the “observed fish in the subunit community” was competitive. Alternatively, if the niche value species in the “subunit community” decreased in response to each removed fish as compared to the niche value before the removal, the relationship between the “removed fish” and the “observed fish in the subunit community” was judged to be mutually beneficial. If the niche value of the species in the “subunit community” did not change in response to simulated fish removal, the relationship between the “removed fish” and the “observed fish in the subunit community” was considered neutral.

4.2.1 Niche Changes in the “Subunit Community”

4.2.1.1 *Mylopharyngodon piceus* (Richardson, 1846)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Mylopharyngodon piceus* (Richardson, 1846) varied from 1.6227% and 12.8275%. The maximum extent of each deletion was 11.2049%, and the final niche

TAB. 4.11 – The “original community” (all fish in the table) and the “subunit community*” of the simulated communities.

Fish species						
<i>Mylopharyngodon piceus*</i>	<i>Hemiculter leucisculus*</i>	<i>Acrossocheilus labiatus</i>	<i>Tor zonatus</i>	<i>Discogobiolongibarbus</i>	<i>Puntius semifasciolatus</i>	<i>Takifugu ocellatus</i>
<i>Ctenopharyngodon idella*</i>	<i>Squalidus argentatus*</i>	<i>Acrossocheilts hemispinus hemispinus</i>	<i>Mystacoleucus marginatus</i>	<i>Discogobio brachyphysallidos</i>	<i>Spinibarbus hollandi</i>	<i>Lateolabrax japonicus</i>
<i>Hypophthalmichthys molitrix*</i>	<i>Lucosoma chinensis*</i>	<i>Acrossocheilus beijiagensis</i>	<i>Sinilabeo rendahli</i>	<i>Discogobio tetrabarbus</i>	<i>Spinibarbus sinensis</i>	<i>Balitora kwangsiensis</i>
<i>Hypophthalmichthys nobilis*</i>	<i>Rhinogobius giurinus*</i>	<i>Acrossocheilts elongatus</i>	<i>Bangana decora</i>	<i>Discocheilus wui</i>	<i>Spinibarbus denticulatus denticulatus</i>	<i>Micronemacheilus pulcher</i>
<i>Megalobrama skolkovii *</i>	<i>Carassius auratus auratus *</i>	<i>Acrossocheilus paradoxus</i>	<i>Sinilabeo discognathoides</i>	<i>Schizothorax meridionalis</i>	<i>Sinocyclocheilus macrolepis</i>	<i>Anabas testudineus</i>
<i>Parabramis pekinensis*</i>	<i>Channa maculata*</i>	<i>Acrossocheilus fasciatus</i>	<i>Sinilabeo discognathoides wui</i>	<i>Procypris merus</i>	<i>Sinocyclocheilus macrocephalus</i>	<i>Pelteobagrus vachellii</i>
<i>Squaliobarbus curriculus*</i>	<i>Pelteobagrus fulvidraco*</i>	<i>Acrossocheilus clivosius</i>	<i>Osteochilus salsburyi</i>	<i>Cyprinus rabaudi</i>	<i>Sinocyclocheilus grahami tingi</i>	<i>Bostrichthys sinensis</i>
<i>Xenocypris argentea*</i>	<i>Opsariichthys bidens*</i>	<i>Acrossocheilus iridescens iridescens</i>	<i>Rectoris posehensis</i>	<i>Cyprinus fuzianensis</i>	<i>Sinocyclocheilus yangzongensis</i>	<i>Pelteobagrus intermedius</i>
<i>Cirrhinus molitorella*</i>	<i>Ancherythroculter lini*</i>	<i>Onychostoma macrolepis</i>	<i>Pseudocrossocheilus bamaensis</i>	<i>Cyprinus yilongensis</i>	<i>Sinocyclocheilus anophthalmus</i>	<i>Tachysurus argentivittatus</i>
<i>Cyprinus carpio*</i>	<i>Rhodeus ocellatus</i>	<i>Onychostoma barbatulum</i>	<i>Parasinilabeo assimilis</i>	<i>Cyprinus longzhouensis</i>	<i>Sinocyclocheilus microphthalmus</i>	<i>Luciobrama macrocephalus</i>
<i>Elopichthys bambusa*</i>	<i>Ptychidio jordani</i>	<i>Onychostoma sima</i>	<i>Semilabeo notabilis</i>	<i>Cyprinus chilia</i>	<i>Luciocyprinus langsoni</i>	<i>Rasbora steineri</i>
<i>Ochetobius elongatus*</i>	<i>Ptychidio macrops</i>	<i>Onychostoma lini</i>	<i>Semilabeo obscurus</i>	<i>Cyprinus pellegrini</i>	<i>Hemibagrus guttatus</i>	<i>Zacco platypus</i>
<i>Siniperca kneri *</i>	<i>Garra orientalis</i>	<i>Onychostoma ovalis rhomboides</i>	<i>Pseudogyriinocheilus prochilus</i>	<i>Carassioides cantonensis</i>	<i>Leptobotia pellegrini</i>	<i>Rasborinus lineatus</i>
<i>Misgurnus anguillicaudatus*</i>	<i>Garra pingi pingi</i>	<i>Folifer brevifilis brevifilis</i>	<i>Garra pingi yiliangensis</i>	<i>Carassius Auratus gibelio</i>	<i>Leiocassis crassilabris</i>	<i>Acheilognathus tonkinensis</i>
<i>Pseudolaubuca sinensis Bleeker, 1865*</i>	<i>Cyprinus multitaeniata</i>	<i>Tor sinensis</i>	<i>Garra pingi hainanensis</i>	<i>Sinibotia pulchra</i>	<i>Hemibagrus macropterus</i>	

was 12.5356%. As shown in figure 4.10, there were 48 corresponding competitive fish, 30 reciprocal fish, and one neutral fish. The maximum niche decrease of *Mylopharyngodon piceus* happens after the deletion of *Hemibagrus guttatus*, and the maximum niche increase of *Mylopharyngodon piceus* occurs after the deletion of *Anabas testudineus*.

4.2.1.2 *Ctenopharyngodon idella* (Valenciennes, 1844)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Ctenopharyngodon idella* (Valenciennes, 1844) varied from 1.1720% to 8.6426%. The maximum extent of each deletion was 7.4706%. As shown in figure 4.11, there were 51 corresponding competitive fish and 28 reciprocal fish. The maximum niche decrease of *Ctenopharyngodon idella* happens after the deletion of *Lateolabrax japonicus*, and the maximum niche increase of *Ctenopharyngodon idella* occurs after the deletion of *Takifugu ocellatus*.

4.2.1.3 *Hypophthalmichthys molitrix* (Valenciennes, 1844)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Hypophthalmichthys molitrix* (Valenciennes, 1844) varied from 10.4459% to 16.4285%. The maximum extent of each deletion was 5.9826%. As shown in figure 4.12, there were 45 corresponding competitive fish and 34 reciprocal fish. The maximum niche decrease of *Hypophthalmichthys molitrix* happens after the deletion of *Acrossocheilus fasciatus*, and the maximum niche increase of *Hypophthalmichthys molitrix* occurs after the deletion of *Spinibarbus sinensis*.

4.2.1.4 *Hypophthalmichthys nobilis* (Richardson, 1845)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Hypophthalmichthys nobilis* (Richardson, 1845) varied from 3.4881% to 10.9246%. The maximum extent of each deletion was 7.4365%. As shown in figure 4.13, there were 50 corresponding competitive fish, 29 reciprocal fish. The maximum niche decrease of *Hypophthalmichthys nobilis* happens after the deletion of *Discogo biolongibarbus*, and the maximum niche increase of *Hypophthalmichthys nobilis* occurs after the deletion of *Zacco platypus*.

4.2.1.5 *Megalobrama skolkovii* (Dybowski, 1872)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Megalobrama skolkovii* (Dybowski, 1872) varied from 1.0454% to 3.9841%. The maximum extent of each deletion was 2.9387%. As shown in figure 4.14, there were 43 corresponding competitive fish and 36 reciprocal fish. The maximum niche decrease of *Megalobrama skolkovii* happens after the deletion of *Garra orientalis*, and the maximum niche increase of *Megalobrama skolkovii* occurs after the deletion of *Acrossocheilts elongatus*.

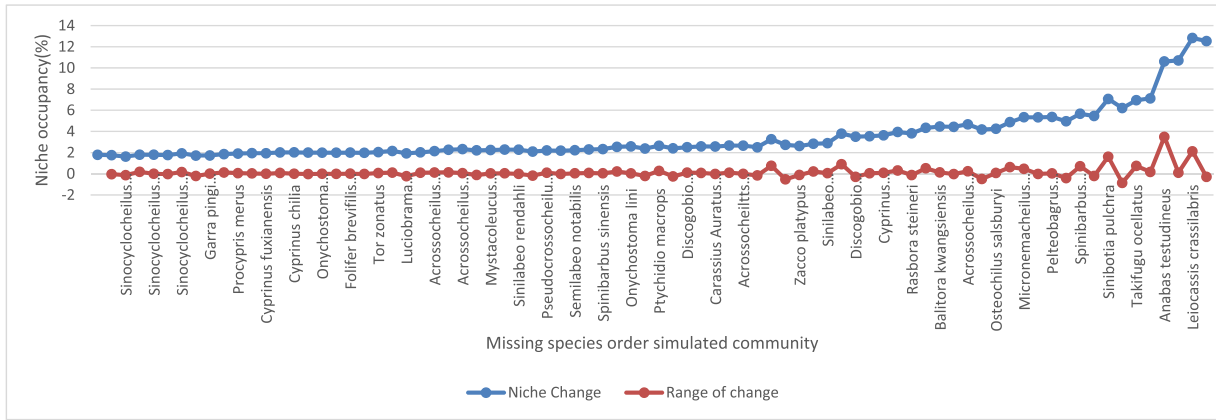


FIG. 4.10 – Amplitude of niche change for *Mylopharyngodon piceus* when species in the “primitive community” were removed one by one.

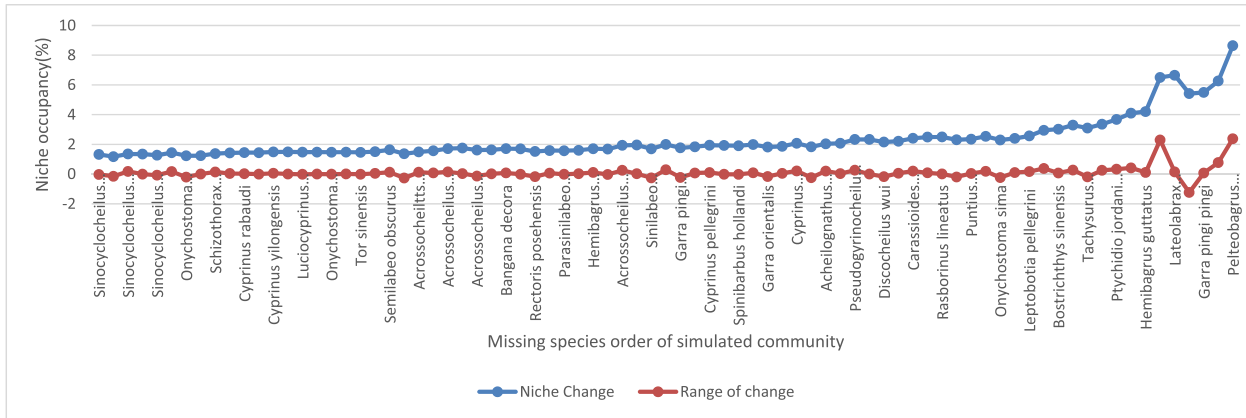


FIG. 4.11 – Amplitude of niche change for *Ctenopharyngodon idella* when species in the “primitive community” were removed one by one.

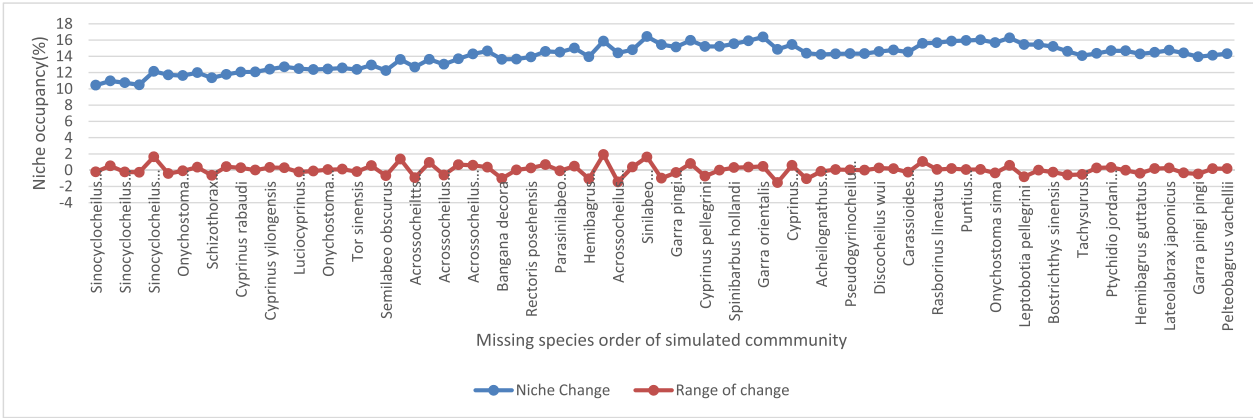


FIG. 4.12 – Amplitude of niche change for *Mhyphthalmichthys molitrix* when species in the “primitive community” were removed one by one.

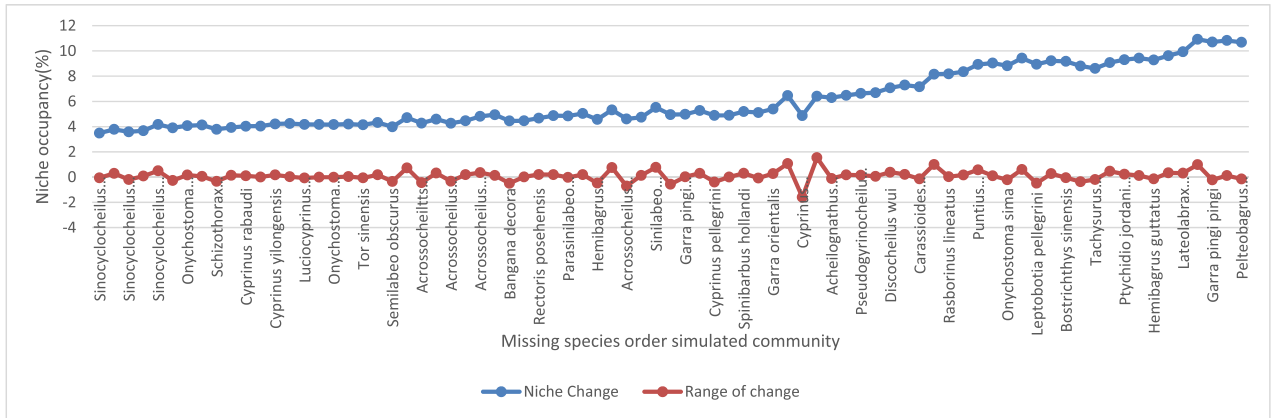


FIG. 4.13 – Amplitude of niche change for *Hypophthalmichthys nobilis* when species in the “primitive community” were removed one by one.

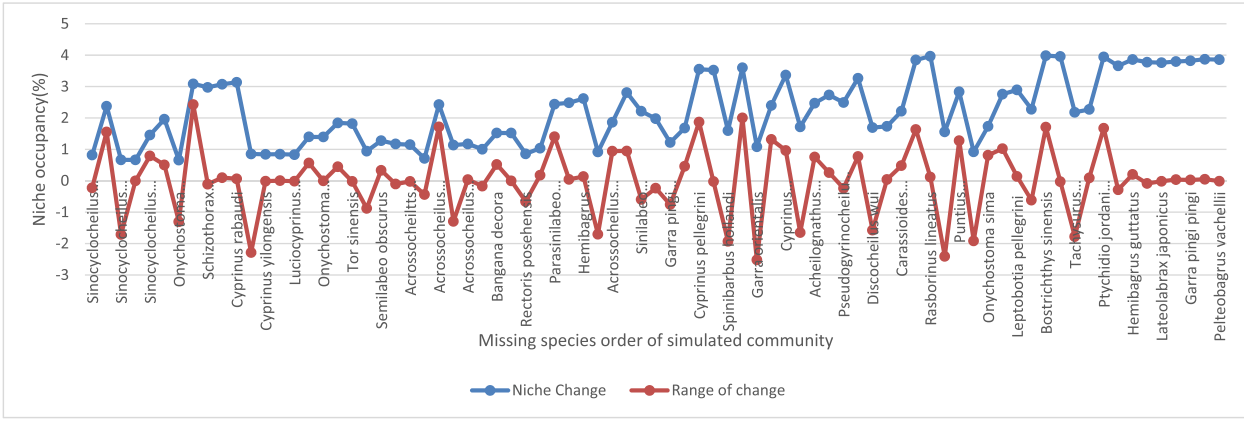


FIG. 4.14 – Amplitude of niche change for *Megalobrama skolkovii* when species in the “primitive community” were removed one by one.

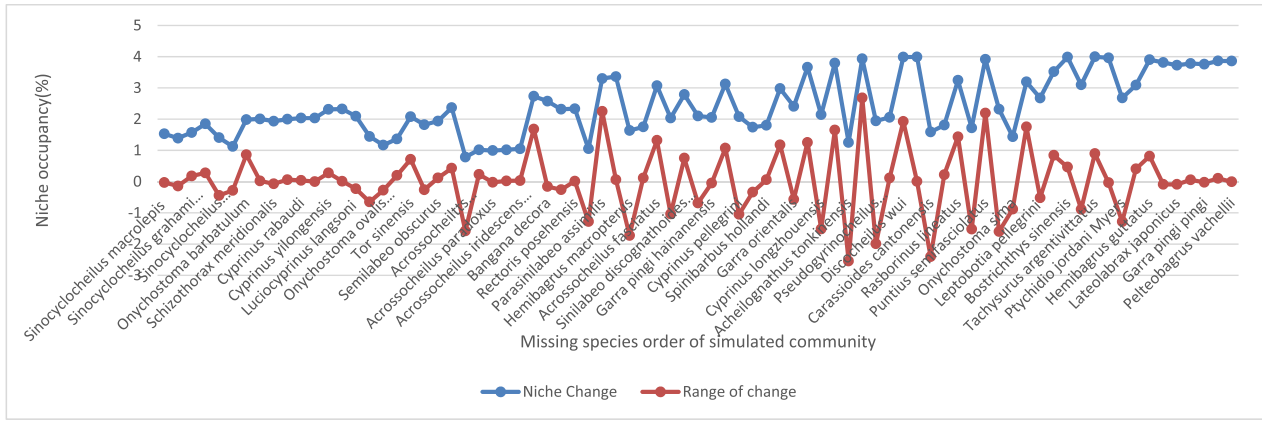


FIG. 4.15 – Amplitude of niche change for *Parabramis pekinensis* when species in the “primitive community” were removed one by one.

4.2.1.6 *Parabramis pekinensis* (Basilewsky, 1855)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Parabramis pekinensis* (Basilewsky, 1855) varied from 0.7864% to 3.9975%. The maximum extent of each deletion was 3.211%. As shown in figure 4.15, there were 43 corresponding competitive fish, and 36 reciprocal fish. The maximum niche decrease of *Parabramis pekinensis* happens after the deletion of *Pseudogyrinocheilus prochilus*, and the maximum niche increase of *Parabramis pekinensis* occurs after the deletion of *Sinilabeo discognathoides*.

4.2.1.7 *Squaliobarbus curriculus* (Richardson, 1846)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Squaliobarbus curriculus* (Richardson, 1846) varied from 0.5375% to 3.9975%. The maximum extent of each deletion was 3.4536%. As shown in figure 4.16, there were 43 corresponding competitive fish and 36 reciprocal fish. The maximum niche decrease of *Squaliobarbus curriculus* happens after the deletion of *Luciobrama macrocephalus*, and the maximum niche increase of *Squaliobarbus curriculus* occurs after the deletion of *Acrossocheilits hemispinus hemispinus*.

4.2.1.8 *Xenocypris argentea* (Günther, 1868)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Xenocypris argentea* (Günther, 1868) varied from 0.0029% to 2.569%. The maximum extent of each deletion was 2.5661%. As shown in figure 4.17, there were 45 corresponding competitive fish and 34 reciprocal fish. The maximum niche decrease of *Xenocypris argentea* happens after the deletion of *Discocheilus wui*, and the maximum niche increase of *Xenocypris argentea* occurs after the deletion of *Cyprinus multitaeniata*.

4.2.1.9 *Cirrhinus molitorella* (Valenciennes, 1844)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Cirrhinus molitorella* (Valenciennes, 1844) varied from 0.0025% to 2.67%. The maximum extent of each deletion was 2.6675%. As shown in figure 4.18, there were 38 corresponding competitive fish and 41 reciprocal fish. The maximum niche decrease of *Cirrhinus molitorella* happens after the deletion of *Acrossocheilus beijiangensis*, and the maximum niche increase of *Cirrhinus molitorella* occurs after the deletion of *Acrossocheilus paradoxus*.

4.2.1.10 *Cyprinus carpio* (Linnaeus, 1758)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Cyprinus carpio* (Linnaeus, 1758) varied from 0.1598% to 3.9503%. The maximum extent of each deletion was 3.7905%. As shown in figure 4.19, there were 38 corresponding competitive fish and 41 reciprocal fish. The maximum niche decrease of *Cyprinus carpio* happens after the deletion of *Mystacoleucus marginatus*,

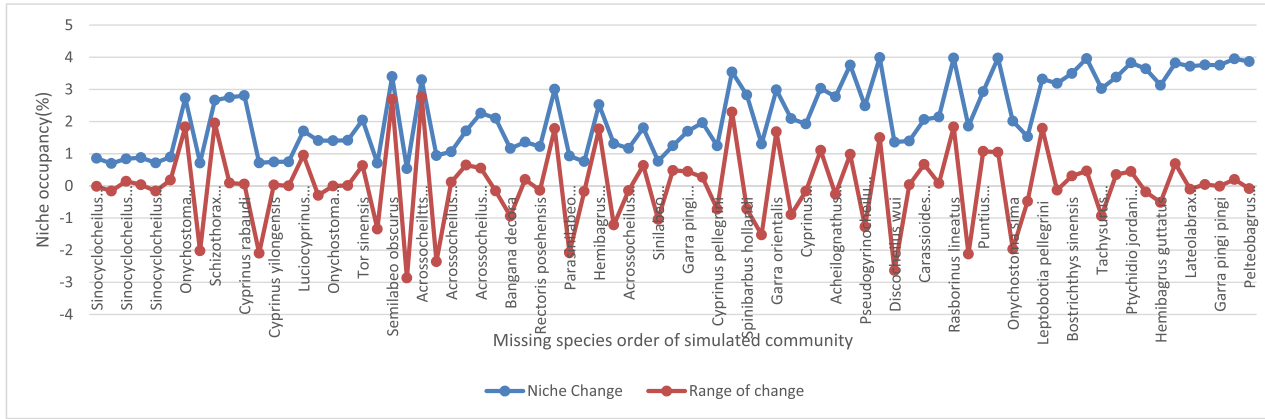


FIG. 4.16 – Amplitude of niche change for *Squaliobarbus curriculus* when species in the “primitive community” were removed one by one.

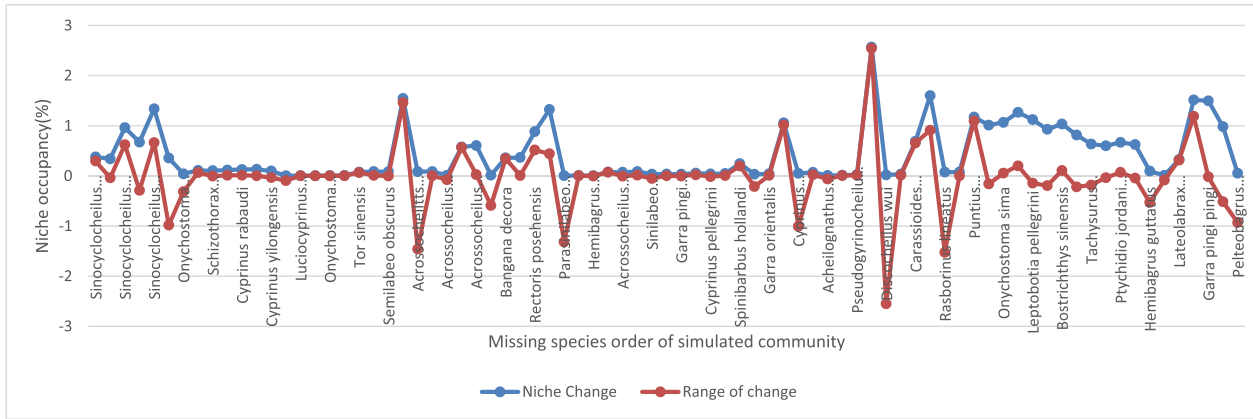


FIG. 4.17 – Amplitude of niche change for *Xenocypris argentea* when species in the “primitive community” were removed one by one.

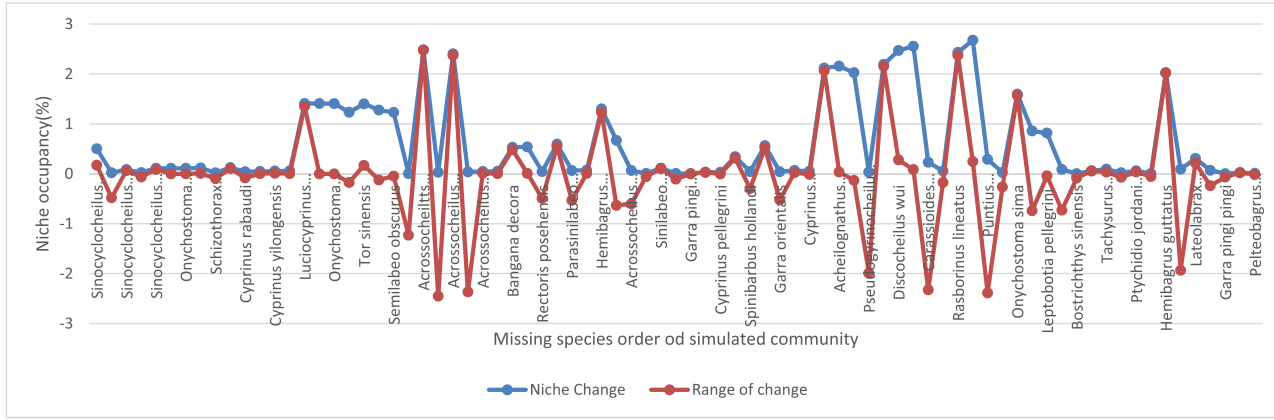


FIG. 4.18 – Amplitude of niche change for *Cirrhinus molitorella* when species in the “primitive community” were removed one by one.

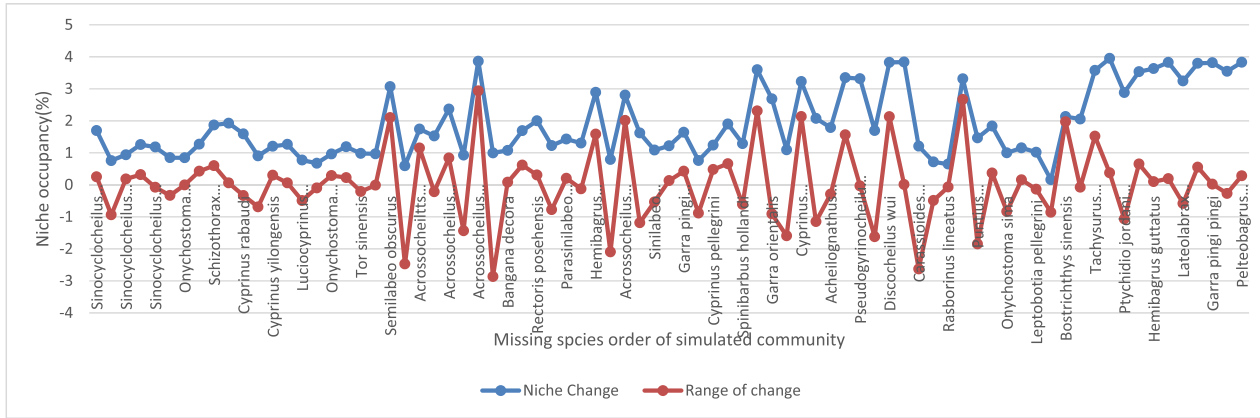


FIG. 4.19 – Amplitude of niche change for *Cyprinus carpio* when species in the “primitive community” were removed one by one.

and the maximum niche increase of *Cyprinus carpio* occurs after the deletion of *Acrossocheilus iridescens iridescens*.

4.2.1.11 *Elopichthys bambusa* (Richardson, 1845)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Elopichthys bambusa* (Richardson, 1845) varied from 1.3382% to 4.3429%. The maximum extent of each deletion was 3.0048%. As shown in figure 4.20, there were 46 corresponding competitive fish and 33 reciprocal fish. The maximum niche decrease of *Elopichthys bambusa* happens after the deletion of *Onychostoma sima*, and the maximum niche increase of *Elopichthys bambusa* occurs after the deletion of *Leptobotia pellegrini*.

4.2.1.12 *Ochetobius elongatus* (Kner, 1867)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Ochetobius elongatus* (Kner, 1867) varied from 0.0060% to 2.8333%. The maximum extent of each deletion was 2.8273%. As shown in figure 4.21, there were 46 corresponding competitive fish and 33 reciprocal fish. The maximum niche decrease of *Ochetobius elongatus* happens after the deletion of *Onychostoma sima*, and the maximum niche increase of *Ochetobius elongatus* occurs after the deletion of *Garra pingi pingi*.

4.2.1.13 *Siniperca kneri* (Garman, 1912)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Siniperca kneri* (Garman, 1912) varied from 0.0199 to 3.9541%. The maximum extent of each deletion was 3.9342%. As shown in figure 4.22, there were 46 corresponding competitive fish and 33 reciprocal fish. The maximum niche decrease of *Siniperca kneri* happens after the deletion of *Leptobotia pellegrini*, and the maximum niche increase of *Siniperca kneri* occurs after the deletion of *Leiocassis crassilabris*.

4.2.1.14 *Misgurnus anguillicaudatus* (Cantor, 1842)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Misgurnus anguillicaudatus* (Cantor, 1842) varied from 0.8690% to 3.9838%. The maximum extent of each deletion was 3.1148%. As shown in figure 4.23, there were 41 corresponding competitive fish and 38 reciprocal fish. The maximum niche decrease of *Misgurnus anguillicaudatus* happens after the deletion of *Puntius semifasciolatus*, and the maximum niche increase of *Misgurnus anguillicaudatus* occurs after the deletion of *Bostrichthys sinensis*.

4.2.1.15 *Pseudolaubuca sinensis* (Bleeker, 1865)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Pseudolaubuca sinensis* (Bleeker, 1865) varied from 0.7101% to 3.9933%.

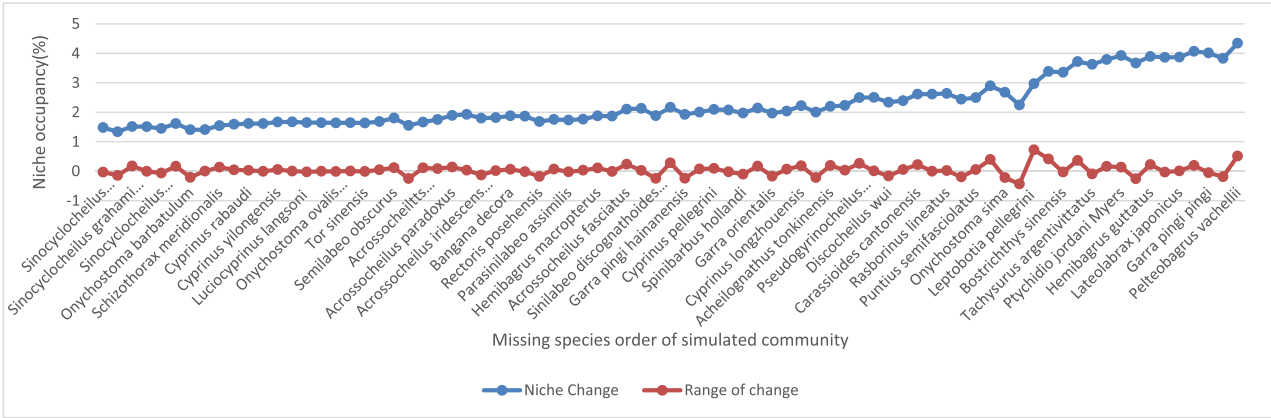


FIG. 4.20 – Amplitude of niche change for *Elopichthys bambusa* when species in the “primitive community” were removed one by one.

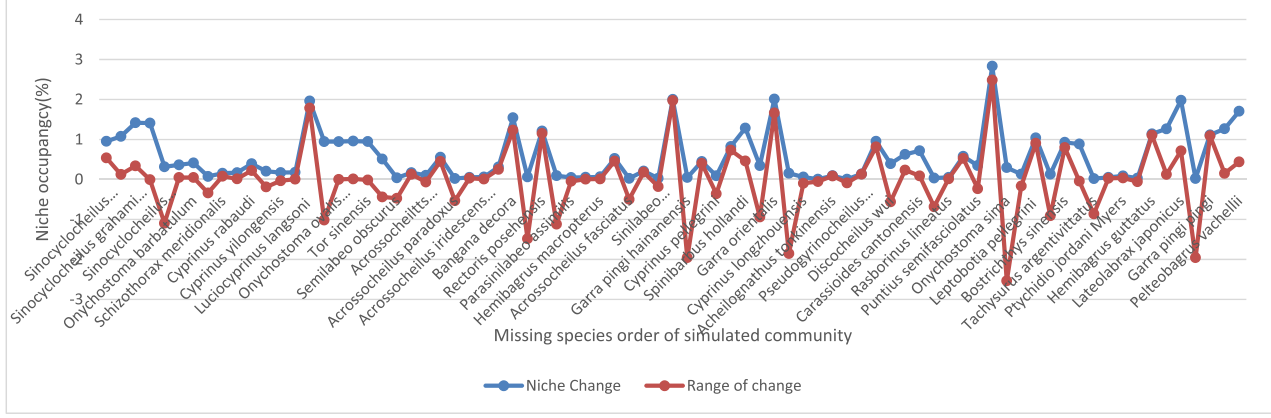


FIG. 4.21 – Amplitude of niche change for *Ochetobius elongatus* when species in the “primitive community” were removed one by one.

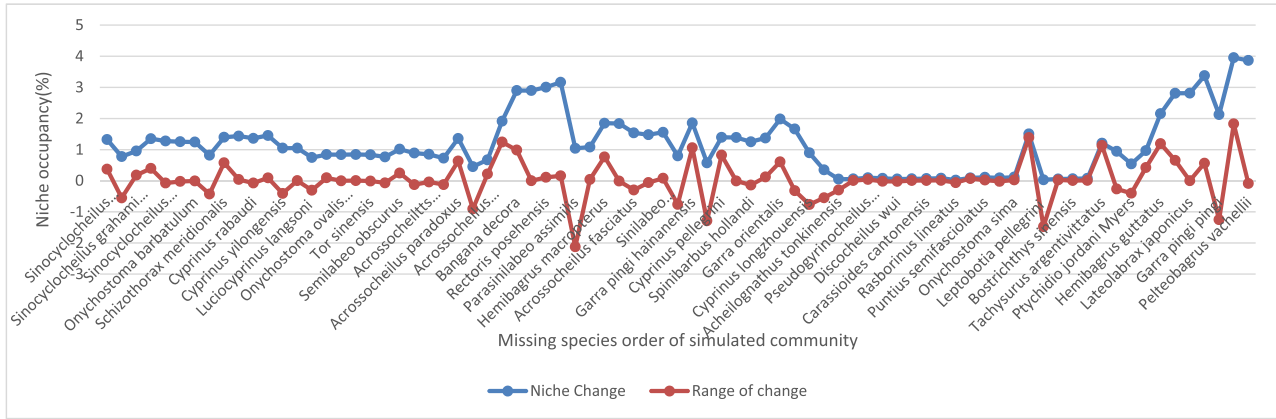


FIG. 4.22 – Amplitude of niche change for *Siniperca kneri* when species in the “primitive community” were removed one by one.

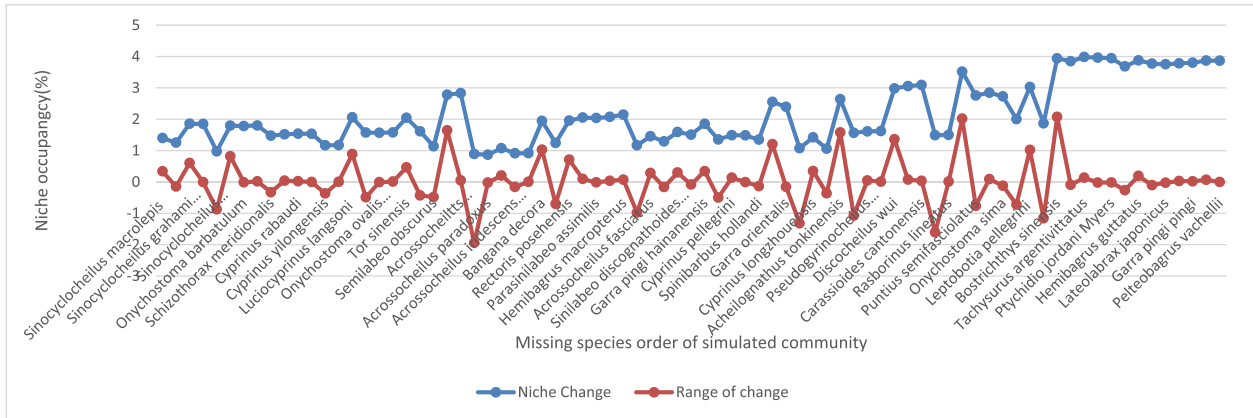


FIG. 4.23 – Amplitude of niche change for *Misgurnus anguillicaudatus* when species in the “primitive community” were removed one by one.

The maximum extent of each deletion was 3.2832%. As shown in figure 4.24, there were 44 corresponding competitive fish and 35 reciprocal fish. The maximum niche decrease of *Pseudolaubuca sinensis* happens after the deletion of *Acrossocheilts hemispinus hemispinus*, and the maximum niche increase of *Pseudolaubuca sinensis* occurs after the deletion of *Luciobrama macrocephalus*.

4.2.1.16 *Hemiculter leucisculus* (Basilewsky, 1855)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Hemiculter leucisculus* (Basilewsky, 1855) varied from 0.0166% to 3.8568%. The maximum extent of each deletion was 3.8402%. As shown in figure 4.25, there were 36 corresponding competitive fish and 43 reciprocal fish. The maximum niche decrease of *Hemiculter leucisculus* happens after the deletion of *Puntius semifasciolatus*, and the maximum niche increase of *Hemiculter leucisculus* occurs after the deletion of *Anabas testudineus*.

4.2.1.17 *Squalidus argentatus* (Sauvage and Dabry De Thiersant, 1874)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Squalidus argentatus* (Sauvage and Dabry de Thiersant, 1874) varied from 0.0075% to 2.1739%. The maximum extent of each deletion was 2.1664%. As shown in figure 4.26, there were 47 corresponding competitive fish and 32 reciprocal fish. The maximum niche decrease of *Squalidus argentatus* happens after the deletion of *Rasbora steineri*, and the maximum niche increase of *Squalidus argentatus* occurs after the deletion of *Parasinilabeo assimilis*.

4.2.1.18 *Lcucosoma chinensis* (Osbeck, 1765)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Lcucosoma chinensis* (Osbeck, 1765) varied from 0.6837% to 3.9527%. The maximum extent of each deletion was 3.269%. As shown in figure 4.27, there were 39 corresponding competitive fish and 40 reciprocal fish. The maximum niche decrease of *Lcucosoma chinensis* happens after the deletion of *Balitora kwangsiensis*, and the maximum niche increase of *Lcucosoma chinensis* occurs after the deletion of *Rasborinus lineatus*.

4.2.1.19 *Rhinogobius giurinus* (Rutter, 1897)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Rhinogobius giurinus* (Rutter, 1897) varied from 0.0351% to 3.6267%. The maximum extent of each deletion was 3.5917%. As shown in figure 4.28 there were 48 corresponding competitive fish and 31 reciprocal fish. The maximum niche decrease of *Rhinogobius giurinus* happens after the deletion of *Rasborinus lineatus*, and the maximum niche increase of *Rhinogobius giurinus* occurs after the deletion of *Balitora kwangsiensis*.

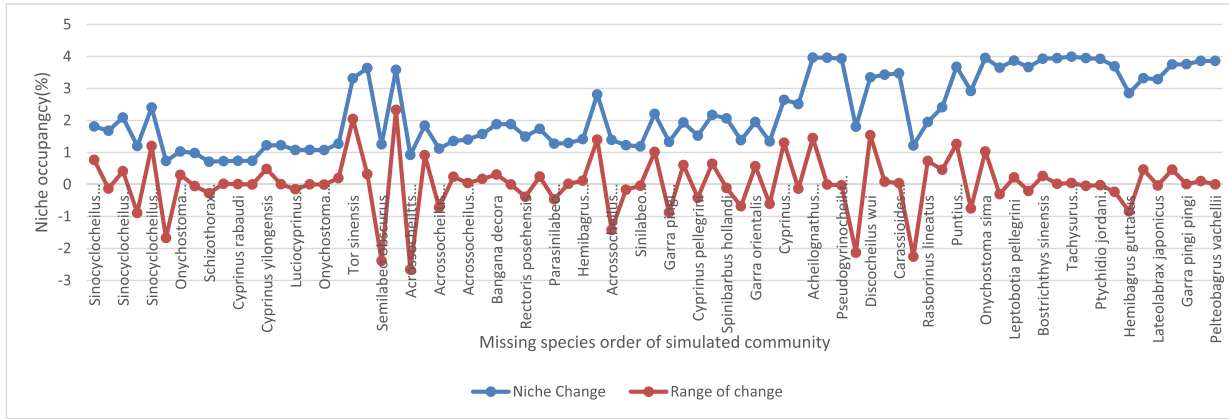


FIG. 4.24 – Amplitude of niche change for *Pseudolaubuca sinensis* when species in the “primitive community” were removed one by one.

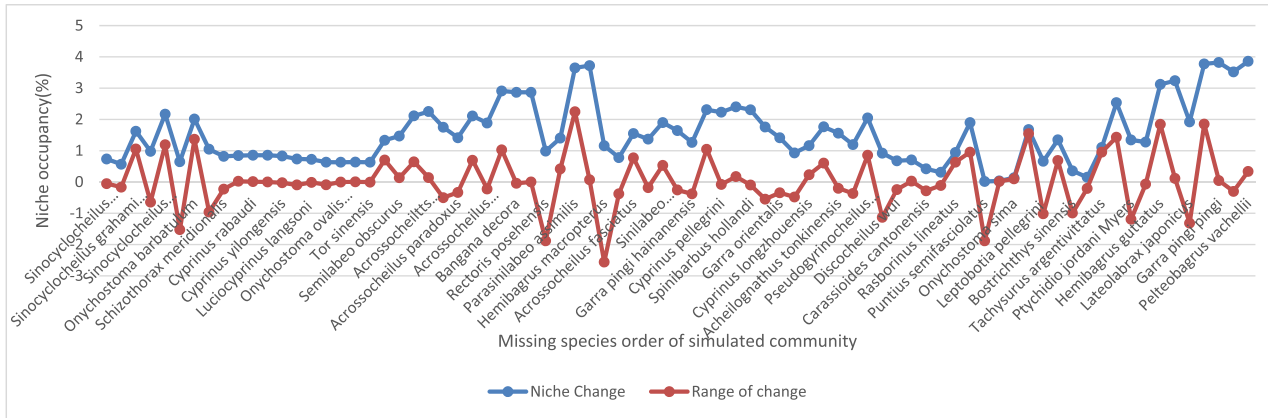


FIG. 4.25 – Amplitude of niche change for *Hemiculter leucisculus* when species in the “primitive community” were removed one by one.

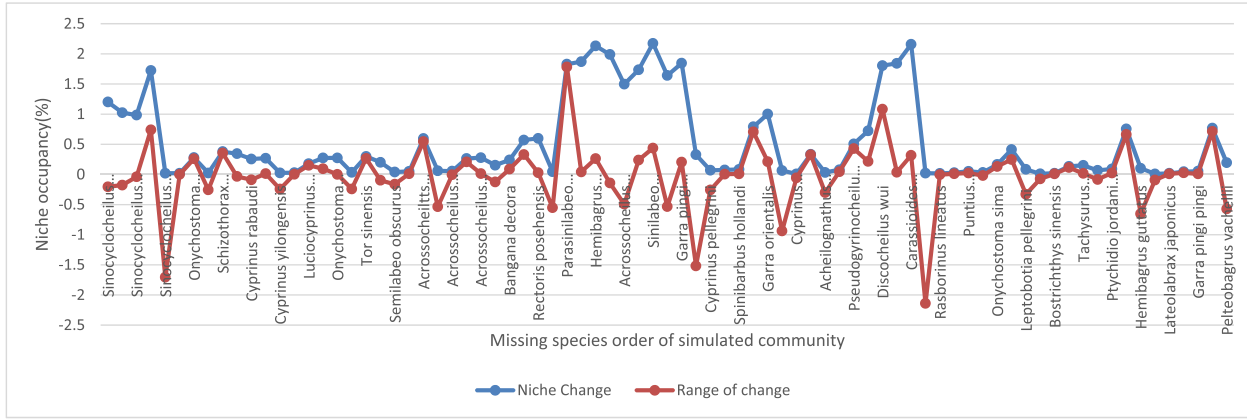


FIG. 4.26 – Amplitude of niche change for *Squalidus argentatus* when species in the “primitive community” were removed one by one.

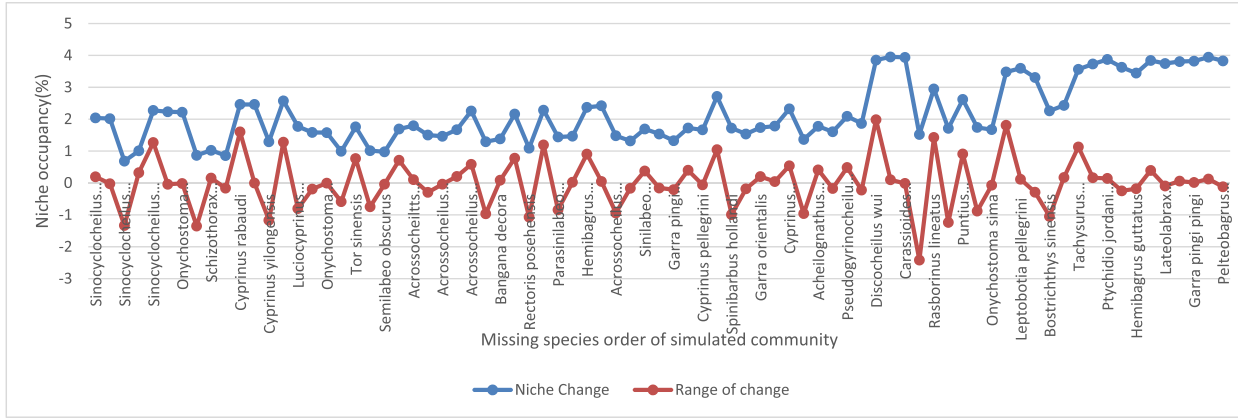


FIG. 4.27 – Amplitude of niche change for *Leucosoma chinensis* when species in the “primitive community” were removed one by one.

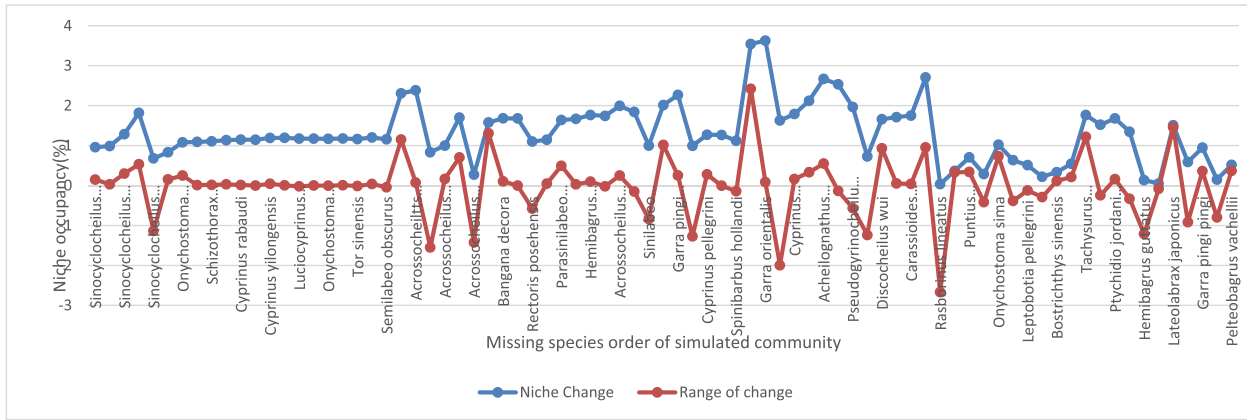


FIG. 4.28 – Amplitude of niche change for *Rhinogobius giurinus* when species in the “primitive community” were removed one by one.

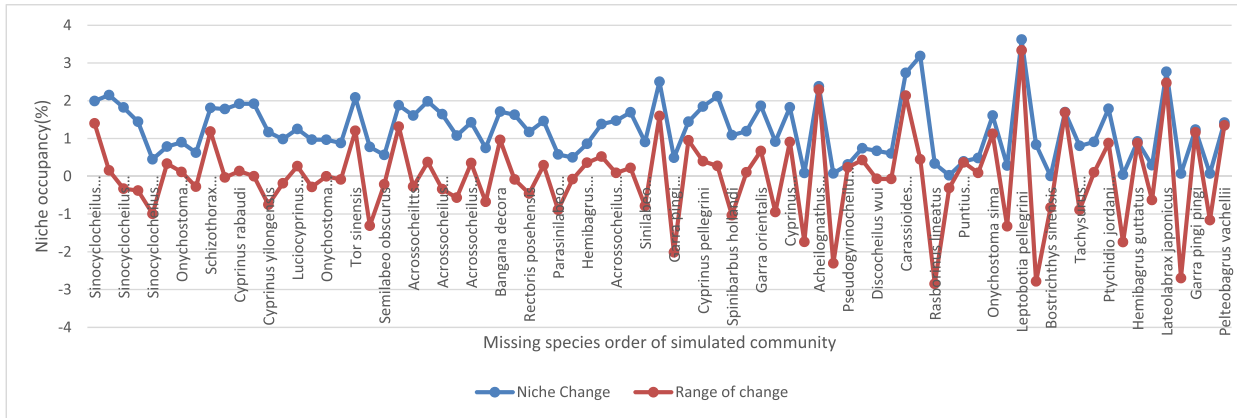


FIG. 4.29 – Amplitude of niche change for *Carassius auratus auratus* when species in the “primitive community” were removed one by one.

4.2.1.20 *Carassius auratus auratus* (Linnaeus, 1758)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Carassius auratus auratus* (Linnaeus, 1758) varied from 0.0068% ~ 3.3347%. The maximum extent of each deletion was 3.6129%. As shown in figure 4.29, there were 41 corresponding competitive fish and 38 reciprocal fish. The maximum niche decrease of *Carassius auratus auratus* happens after the deletion of *Micronemacheilus pulcher*, and the maximum niche increase of *Carassius auratus auratus* occurs after the deletion of *Leptobotia pellegrini*.

4.2.1.21 *Channa maculata* (Lacépède, 1801)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Channa maculata* (Lacépède, 1801) varied from 0.4258% to 3.8833%. The maximum extent of each deletion was 3.4575%. As shown in figure 4.30, there were 49 corresponding competitive fish and 30 reciprocal fish. The maximum niche decrease of *Channa maculata* happens after the deletion of *Rectoris posehensis*, and the maximum niche increase of *Channa maculata* occurs after the deletion of *Onychostoma sima*.

4.2.1.22 *Pelteobagrus fulvidraco* (Richardson, 1846)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Pelteobagrus fulvidraco* (Richardson, 1846) varied from 0.565% to 3.998%. The maximum extent of each deletion was 3.433%. As shown in figure 4.31, there were 43 corresponding competitive fish, 35 reciprocal fish, and one neutral type. The maximum niche decrease of *Pelteobagrus fulvidraco* happens after the deletion of *Acheilognathus tonkinensis*, and the maximum niche increase of *Pelteobagrus fulvidraco* occurs after the deletion of *Sinibotia pulchra*.

4.2.1.23 *Opsariichthys bidens* (Günther, 1873)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Opsariichthys bidens* (Günther, 1873) varied from 0.0052% ~ 2.6641%. The maximum extent of each deletion was 2.6589%. As shown in figure 4.32, there were 49 corresponding competitive fish and 30 reciprocal fish. The maximum niche decrease of *Opsariichthys bidens* happens after the deletion of *Cyprinus longzhouensis*, and the maximum niche increase of *Opsariichthys bidens* occurs after the deletion of *Discogobiolongibarbatas*.

4.2.1.24 *Ancherythroculter lini* (Luo, 1994)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Ancherythroculter lini* (Luo, 1994) varied from 0.007% to 3.752%. The maximum extent of each deletion was 2.6589%. As shown in figure 4.33, there were

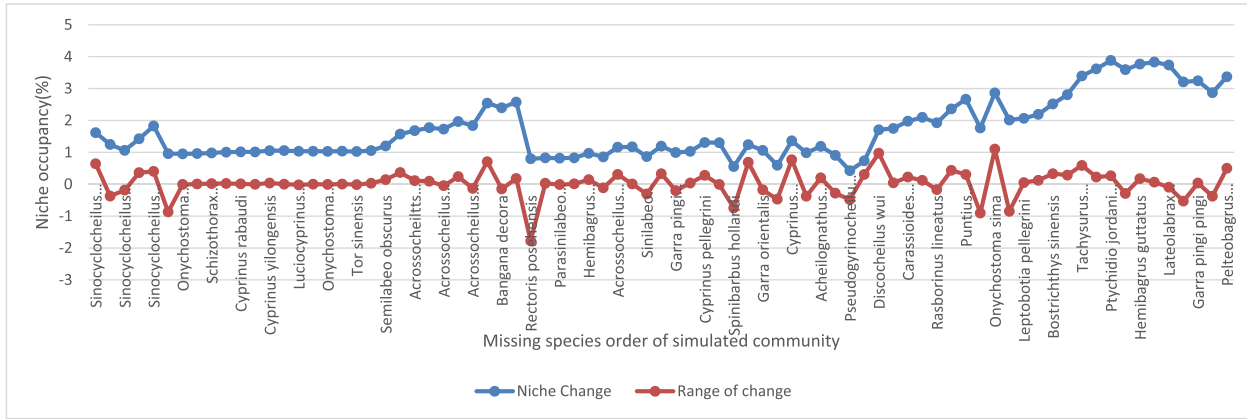


FIG. 4.30 – Amplitude of niche change for *Channa maculata* when species in the “primitive community” were removed one by one.

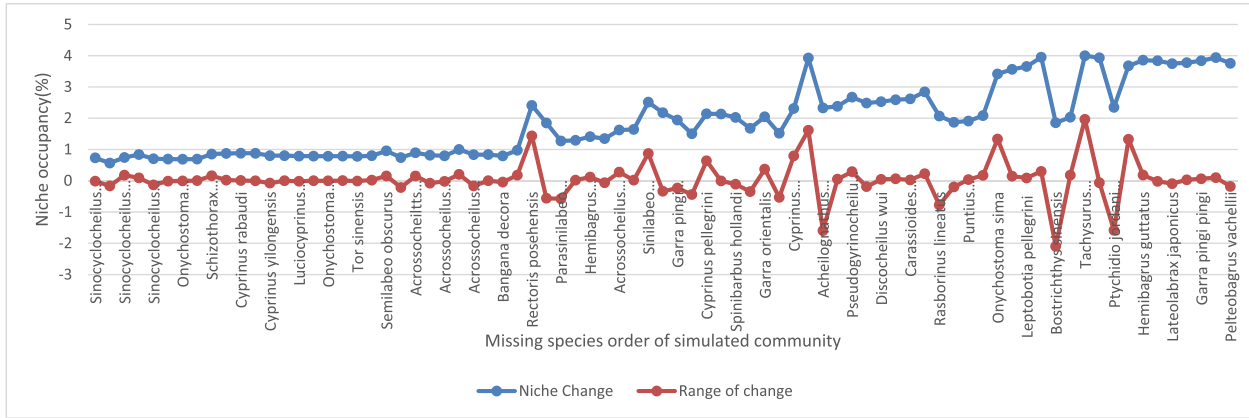


FIG. 4.31 – Amplitude of niche change for *Pelteobagrus fulvidraco* when species in the “primitive community” were removed one by one.

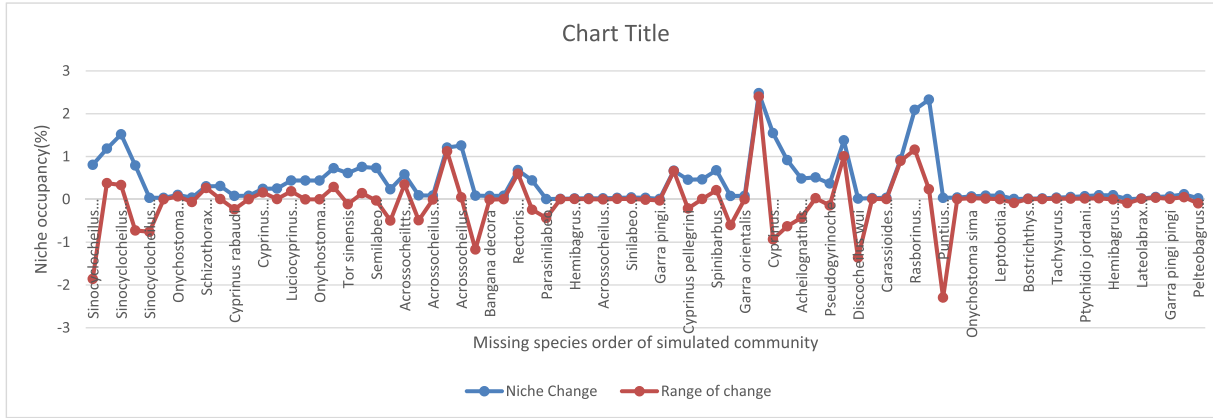


FIG. 4.32 – Amplitude of niche change for *Opsariichthys bidens* when species in the “primitive community” were removed one by one.

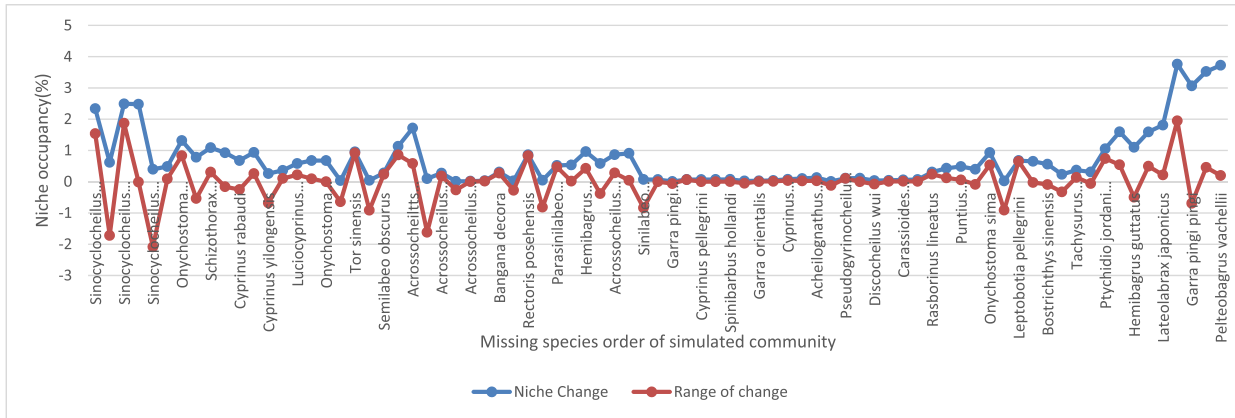


FIG. 4.33 – Amplitude of niche change for *Ancherythroculter lini* when species in the “primitive community” were removed one by one.

49 corresponding competitive fish and 30 reciprocal fish. The maximum niche decrease of *Ancherythroculter lini* happens after the deletion of *Sinocyclocheilus macrocephalus*, and the maximum niche increase of *Ancherythroculter lini* occurs after the deletion of *Sinocyclocheilus grahami tingi*.

4.2.1.25 *Rhodeus ocellatus* (Kner, 1867)

When we deleted 79 species of fish from the “primitive community” one by one, the niche of *Rhodeus ocellatus* (Kner, 1867) varied from 0.006% ~ 3.7226%. The maximum extent of each deletion was 3.7166%. As shown in figure 4.34, there were 46 corresponding competitive fish and 33 reciprocal fish. The maximum niche decrease of *Rhodeus ocellatus* happens after the deletion of *Cyprinus longzhouensis*, and the maximum niche increase of *Rhodeus ocellatus* occurs after the deletion of *Zacco platypus*.

4.2.2 Simulated Community Performance After Species Removal

Taking the initial niche occupation of the 104 species of fish as the reference standard, and assuming that some species disappear one by one, we compared increases and decreases in the niche occupation of each fish species after the removal of each other fish species to evaluate niche correlations between pairs of fish. If the niche occupation value decreased or increased, the removed fish was considered a “mutual benefit type” or “competitive type.” If the niche occupation value remained unchanged or changed very little, it was deemed a “non-competitive type.”

“Competitive,” “mutually beneficial,” and “non-competitive” are represented by “1,” “2,” and “3” in the model. The model analysis results for 74 fish and 25 fish in the “subunit community” were clustered. Figure 4.35 shows that most fish were “competitive” or “mutually beneficial.”

The 79 species of fish were roughly divided into four groups. The first group (the majority) was upstream fish. The second largest group was downstream fish, followed by the fish of the middle reaches, and, finally, highland or cavefish (see table 4.12).

The subcommunity had four clusters of relationships with the removed 79 species: category 1 included two species; category 2 included nine species; category 3 included eight species, and category 4 included six species. In total, 25 fish species exhibited different types of interspecies relationships (see table 4.13).

The classification of 104 fish based on relationships indicated that the model classifies the overall relationships among fish spatial distributions, consistent with the real-world spatial distribution patterns of such fish. The model analysis results also indicated that fish distributed in the upper, middle, and downstream river areas differ with respect to feeding composition, and the described niche relationships comply with the basic law of the food chain. This section describes the niche competition relationships among different types of fish.

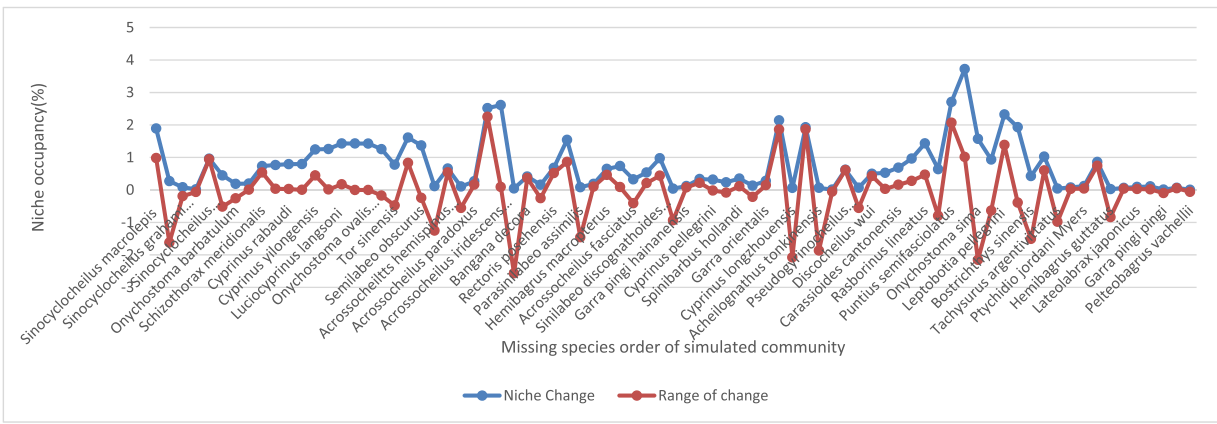


FIG. 4.34 – Amplitude of niche change for *Rhodeus ocellatus* when species in the “primitive community” were removed one by one.



FIG. 4.35 – The response of the “subunit community” to 79 species.

TAB. 4.12 – Cluster characteristics of the 79 fish species removed.

Cluster group	Fish species	Main geographical distribution characteristics
1	<i>Procypris merus</i> , <i>Cyprinus raudii</i> , <i>Cyprinus fuzianensis</i> , <i>Sinocyclocheilus macrocephalus</i> , <i>Sinocyclocheilus anophthalmus</i> , <i>Luciocyprinus langsoni</i> , <i>Folifer brevifilis brevifilis</i> , <i>Tor sinensis</i> , <i>Onychostoma sima</i> , <i>Onychostoma ovalis rhomboides</i> , <i>Onychostoma macrolepis</i> , <i>Sinilabeo rendahli</i> , <i>Parasinilabeo assimilis</i> , <i>Sinilabeo discognathoides</i> , <i>Sinilabeo discognathoides wui</i> , <i>Discocheilus wui</i> , <i>Rectoris posehensis</i> , <i>Discogobio brachyphysallidos</i> , <i>Mystacoleucus marginatus</i> , <i>Pseudocrossocheilus bamaensis</i> , <i>Zacco platypus</i> , <i>Luciobrama macrocephalus</i> , <i>Spinibarbus sinensis</i> , <i>Spinibarbus hollandi</i> , <i>Acrossocheilus iridescens iridescens</i> , <i>Acrossocheilus paradoxus</i> , <i>Acrossocheilus clivosius</i> , <i>Osteochilus salsburyi</i> , <i>Garra pingi hainanensis</i> , <i>Garra orientalis</i> , <i>Balitora kwangsiensis</i> , <i>Sinibotia pulchra</i> , <i>Tachysurus argentivittatus</i> , <i>Carassius Auratus gibelio</i> , <i>Bostrichthys sinensis</i> , <i>Anabas testudineus</i>	Mainly upstream
2	<i>Cyprinus pellegrini</i> , <i>Cyprinus longzhouensis</i> , <i>Semilabeo obscurus</i> , <i>Bangana decora</i> , <i>Discogobiolongibarbatulus</i> , <i>Discogobio tetrabarbatulus</i> , <i>Onychostoma lini</i> , <i>Onychostoma barbatulum</i> , <i>Acrossocheilus beijiangensis</i> , <i>Acrossocheilus fasciatus</i> , <i>Acrossocheilus fasciatus</i> , <i>Acrossocheilts hemispinus hemispinus</i> , <i>Carassioides cantonensis</i> , <i>Rasborinus lineatus</i> , <i>Pseudogyrinocheilus prochilus</i> , <i>Micronemacheilus pulcher</i> , <i>Pelteobagrus intermedius</i> , <i>Spinibarbus denticulatus denticulatus</i> , <i>Ptychidio macrops</i> , <i>Lateolabrax japonicus</i> , <i>Garra pingi pingi</i> , <i>Pelteobagrus vachellii</i> , <i>Hemibagrus macropterus</i> , <i>Ptychidio jordani</i> , <i>Hemibagrus guttatus</i> , <i>Acrossocheilus labiatus</i> , <i>Leptobotia pellegrini</i> , <i>Leiocassis crassilabris</i> , <i>Acheilognathus tonkinensis</i> , <i>Takifugu ocellatus</i>	Midstream
3	<i>Semilabeo notabilis</i> , <i>Cyprinus multitaeniata</i> , <i>Ochetobius elongatus</i> , <i>Pseudolaubuca sinensis</i> , <i>Squalidus argentatus</i> , <i>Hemiculter leucisculus</i> , <i>Puntius semifasciolatus</i> , <i>Pelteobagrus fulvidraco</i> , <i>Siniperca kneri</i> , <i>Xenocypris argentea</i> , <i>Mylopharyngodon piceus</i> , <i>Ctenopharyngodon idella</i> , <i>Hypophthalmichthys molitrix</i> , <i>Hypophthalmichthys nobilis</i> , <i>Elopichthys bambusa</i> , <i>Ancherythroculter lini</i> , <i>Megalobrama terminalis</i> , <i>Parabramis pekinensis</i> , <i>Squaliobarbus curriculus</i> , <i>Misgurnus anguillicaudatus</i> , <i>Cirrhinus molitorella</i> , <i>Rasbora steineri</i> , <i>Opsarichthys bidens</i> , <i>Cyprinus carpio</i> , <i>Carassius auratus auratus</i> , <i>Channa maculata</i> , <i>Rhodeus ocellatus</i> , <i>Lucosoma chinensis</i> , <i>Rhinogobius giurinus</i>	Downstream

TAB. 4.12 – (continued).

Cluster group	Fish species	Main geographical distribution characteristics
4	<i>Garra pingi yiliangensis</i> , <i>Cyprinus chilia</i> , <i>Sinocyclocheilus macrolepis</i> , <i>Sinocyclocheilus yangzongensis</i> , <i>Sinocyclocheilus microphthalmus</i> , <i>Sinocyclocheilus macrolepis</i> , <i>Cyprinus yilongensis</i> , <i>Sinocyclocheilus grahami tingi</i> , <i>Schizothorax meridionalis</i>	Primarily special habitats, such as plateaus or caves

TAB. 4.13 – Cluster characteristics of 25 fish species in the “subunit community.”

Type	Fish species
1	<i>Hypophthalmichthys molitrix</i> , <i>Xenocypris argentea</i>
2	<i>Lucosoma chinensis</i> , <i>Hemiculter leucisculus</i> , <i>Cyprinus carpio</i> , <i>Rhodeus ocellatus</i> , <i>Opsariichthys bidens</i> , <i>Squalidus argentatus</i> , <i>Ancherythroculter lini</i> , <i>Cirrhinus molitorella</i> , <i>Siniperca kneri</i>
3	<i>Squaliobarbus curriculus</i> , <i>Megalobrama terminalis</i> , <i>Carassius auratus auratus</i> , <i>Ochetobius elongatus</i> , <i>Rhinogobius giurinus</i> , <i>Pseudolaubuca sinensis</i> , <i>Channa maculata</i> , <i>Misgurnus anguillicaudatus</i>
4	<i>Elopichthys bambusa</i> , <i>Mylopharyngodon piceus</i> , <i>Ctenopharyngodon idella</i> , <i>Hypophthalmichthys nobilis</i> , <i>Pelteobagrus fulvidraco</i> , <i>Parabramis pekinensis</i>

4.2.3 The Evolutionary Characteristics of the “Subunit Communities”

4.2.3.1 The Changing Patterns of the “Subunit Community”

After 79 rounds of species removal, the niche after species removal (A'_1) minus the niche before species removal (A) was the niche change. Thus, the final niche of each species in the “subunit community” was $\sum (A'_1 - A_i)$, $i = 1, 2, 3, \dots, 79$. This value was used as the final niche change for various fish in the “subunit community” to measure the status of the “subunit community” in the final subunit niche after succession. The analysis uncovers one species with an absolute niche change value $\geq 10\%$, two species with an absolute niche change value $\geq 5\%$ and $< 10\%$, and six species with an absolute niche change value < 0 , corresponding to niche loss (see table 4.14). As the number of species in the community decreased, 80% of the fish species niches in the “subunit community” expanded to varying degrees, and the remaining 20% of the species niches were compressed. The niche gain for the community species during succession was not proportional to species body size. *Mylopharyngodon piceus*, grass carp, silver carp, bighead, and *Elopichthys bambusa* are all large fish. In the “original community” composed of 104 species of fish, the silver carp occupied more than 10% of the niches, and the other four species had similar niches. In comparison to the “original community,” succession in the “subunit community” showed that although the *Elopichthys bambusa* niche increased, this increase was less than the niche increases in *Mylopharyngodon piceus*, grass carp, silver carp, and bighead. In the real-world data, *Elopichthys bambusa* niche increased by more than 5%. These results indicated that the parameters of fish form included niche information and were somewhat consistent with the real-world observations. The “subunit community” gradually evolved into a community dominated by *Mylopharyngodon piceus*, grass carp, silver carp, bighead, and *Elopichthys bambusa*, and the niche advantage of these five species was about 50%, indicating that the species diversity of the community decreased and the niches of the dominant species composed of large fish became prominent. Large fish, including *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, and *Elopichthys bambusa*, also occupied large niches in the “primitive community.” *Hypophthalmichthys molitrix* occupied more than 10% of the niche, and the other four fish had similar niches. When the “primitive community” acted as a “subunit community,” the niche of *Elopichthys bambusa* increased greatly, but this increase was significantly less than niche increases of *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, and *Hypophthalmichthys nobilis*. The final result of the model agreed with the real-world data to some degree. The niche dominance of these five fish species was about 50% when the species diversity of the community declined. The dominant species are large fish that are prominent in the system.

4.2.3.2 Succession Trends

Analysis of the reaction of 25 species of fish to 79 species of fish indicated that these fish had little impact on changes in the four major fish (*Mylopharyngodon piceus*,

TAB. 4.14 – Absolute niche after the simulated community changes into “subunit community.”

Fish	$\sum (A'_i - A_i)$ (%)	Subunit community (%)
<i>Mylopharyngodon piceus</i>	10.745428	12.53558
<i>Ctenopharyngodon idella</i>	7.296465	8.642592
<i>Hypophthalmichthys nobilis</i>	7.131931	10.68294
<i>Hypophthalmichthys molitrix</i>	3.6769	14.32392
<i>Hemiculter leucisculus</i>	3.07087	3.85675
<i>Pelteobagrus fulvidraco</i>	3.011	3.757
<i>Squaliobarbus curriculus</i>	2.99408	3.86841
<i>Ancherythroculter lini</i>	2.923	3.723
<i>Siniperca kneri</i>	2.91154	3.86522
<i>Elopichthys bambusa</i>	2.82953	4.3429
<i>Pseudolaubuca sinensis</i>	2.81752	3.86223
<i>Megalobrama skolkovii</i>	2.811249	3.856686
<i>Misgurnus anguillicaudatus</i>	2.80735	3.86688
<i>Channa maculata</i>	2.3955	3.3709
<i>Cyprinus carpio</i>	2.38339	3.82759
<i>Parabramis pekinensis</i>	2.299093	3.859293
<i>Lucosoma chinensis</i> <i>Lucosoma chinensis</i>	1.98299	3.8274
<i>Ochetobius elongatus</i>	1.2908	1.70556
<i>Carassius auratus auratus</i>	0.82834	1.42218
<i>Xenocypris argentea</i>	-0.031	0.052
<i>Rhinogobius giurinus</i>	-0.29253	0.51655
<i>Cirrhinus molitorella</i>	-0.3175	0.0096
<i>Rhodeus ocellatus</i>	-0.9047	0.006
<i>Squalidus argentatus</i>	-1.21162	0.19488
<i>Opsariichthys bidens</i>	-2.6397	0.0244

Ctenopharyngodon idella, *Hypophthalmichthys nobilis*, and *Hypophthalmichthys molitrix*). This indicated that the fish community patterns in the Pearl River have formed over evolutionary time, and it is usually difficult for changes in other species to affect this pattern. According to Lu (1990), the fishing output of the middle and lower reaches of the Pearl River reached 40%–50% in the 1980s, and in this output, *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys nobilis*, and *Hypophthalmichthys molitrix* were primarily the dominant species. In recent decades, human activities have greatly changed the river ecosystem. These habitat changes have affected fish diversity and distribution patterns, especially those of dominant and rare species. Some of these fish have become endangered, while others have disappeared or gone extinct. It is meaningful to study changes in fish distribution patterns in order to understand species' roles and the impact of each fish on other fish. Such studies will help us to maintain and restore the structure and function of the ecosystem species community.

4.2.3.2.1 Negatively Correlated Variants

For the 25 fish species in the “subunit community,” changes in the community niche were negatively correlated with increases in “primitive community” species, such as *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, and *Elopichthys bambusa*. As the community species decreased, the ecological niche increased continuously; the initial niche of *Mylopharyngodon piceus* was 1.79%, and the niche increased to a maximum value of 12.83%, which corresponded to a niche elevation of 11.04% (see figure 4.36). In the figure, the dotted trend line indicates that the niche of *Mylopharyngodon piceus* was negatively correlated with species density.

The niche succession analysis suggested that the niches of species in the disturbed community were in a “disordered” state. The species in the “primitive community” were lost, and the niche change of 80% of the fish species in the “subunit community” did not exceed 5%. Moreover, the ecological niches of various fish were in a state of “alternating mutual complement” during community species change, maintaining the stable niche expansion of the dominant species in the system.

4.2.3.2.2 Positively Correlated Variants

The community niche changes of the “subunit community” species were positively associated with reductions in the “primitive community” species. The niche of *Opsariichthys bidens* decreased from 2.6641% to 0.0052% as the species were removed (see figure 4.37). Species of this type included *Rhodeus ocellatus*, *Rhinogobius giurinus*, *Opsariichthys bidens*, *Squalidus argentatus*, and *Carassius auratus auratus*.

Rhodeus ocellatus is a small fish in the simulated community that has a small niche in the original community. Overall, the niche of *Rhodeus ocellatus* diminished as species decreased. However, the disappearance of some fish species led to up to 4% increases in the niche of *Rhodeus ocellatus*, suggesting that these fish exist in a mutually beneficial relationship with *Rhodeus ocellatus*. Alternatively, another mechanism may adjust the balance of the community’s ecological niche (see figure 4.38).

The analysis of the interspecific relationships within the simulated communities showed that the niches of some fish increased with the loss of the niches of other fish, and the niches of some fish decreased with the loss of the niches of other fish. These fish comprised the majority of the simulated communities.

4.2.3.2.3 Zigzag Patterns

The initial niche of *Megalobrama skolkovii* was 1.0454%, and the maximum niche value was 3.9841%, an increase of 2.9387% (see figure 4.39). *Megalobrama skolkovii* was a medium-sized fish in the simulated community that had a small niche in the original community. As the community species decreased, the niche of *Megalobrama skolkovii* fluctuated, with some fish removals decreasing the niche of *Megalobrama skolkovii* and some increasing this niche (up to a maximum of 4%). This alternation between competition and mutual benefit showed that *Megalobrama skolkovii* cannot adapt to environmental changes. Alternatively, some other mechanism may adjust the balance of community niches. Similar species included *Parabramis pekinensis*,

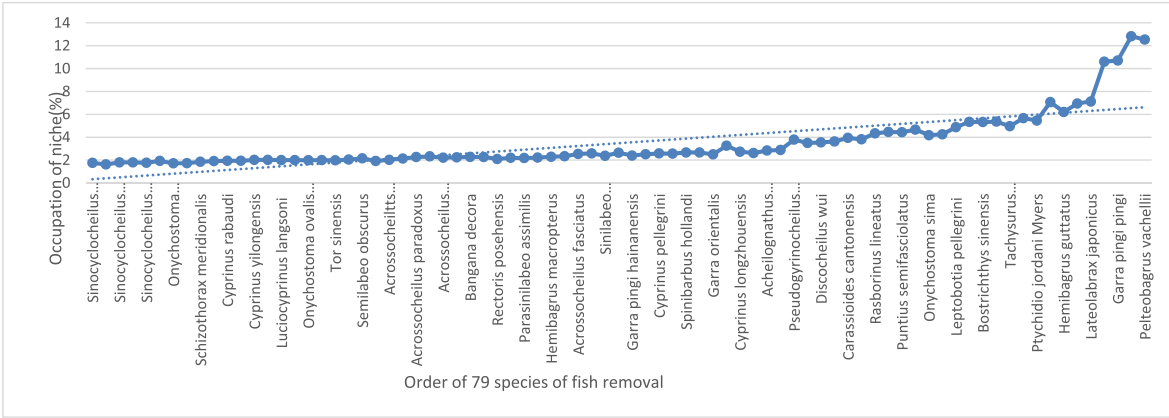


FIG. 4.36 – The niche of *Mylopharyngodon piceus* increased as 79 fish species were removed from the community.

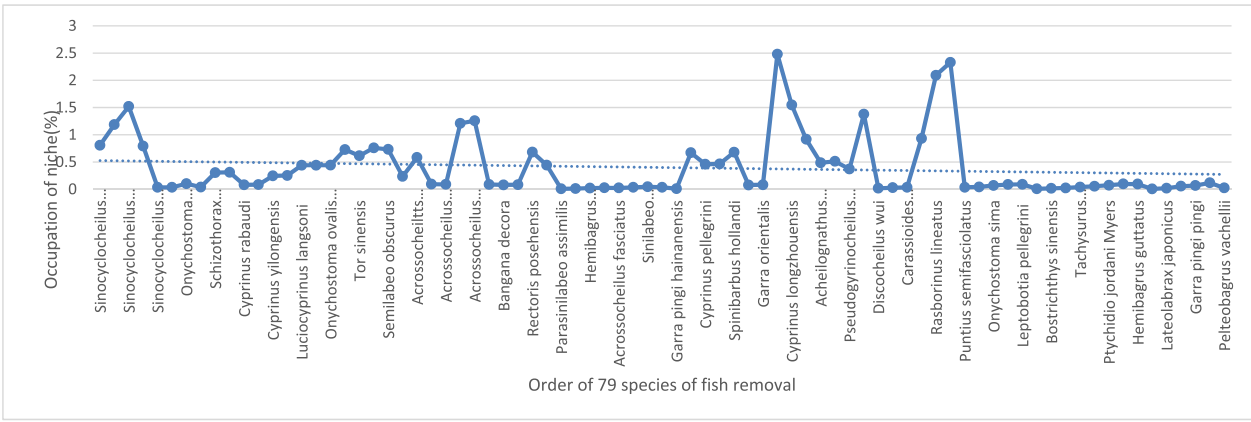


FIG. 4.37 – The niche of *Opsariichthys bidens* declined as 79 species were removed from the community.

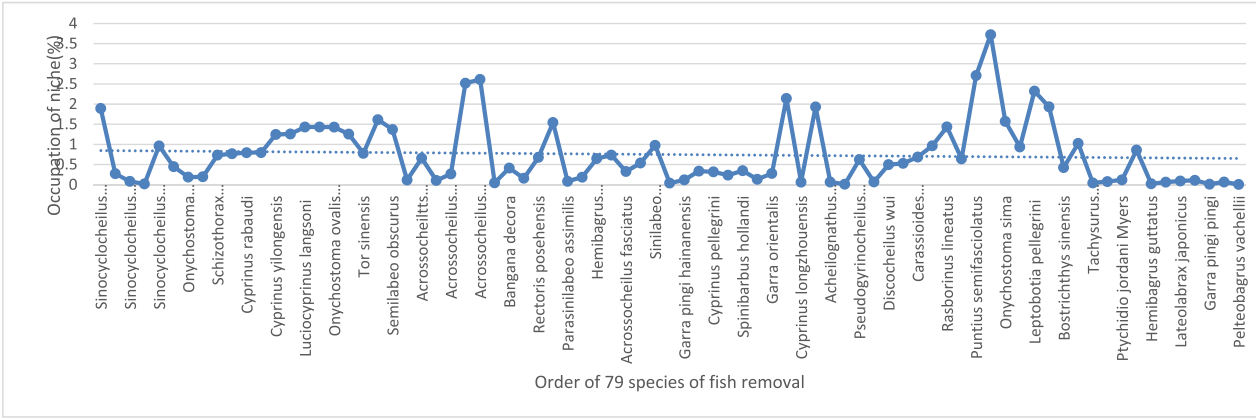


FIG. 4.38 – The niche of *Rhodeus ocellatus* decreased as 79 fish species were removed from the community.

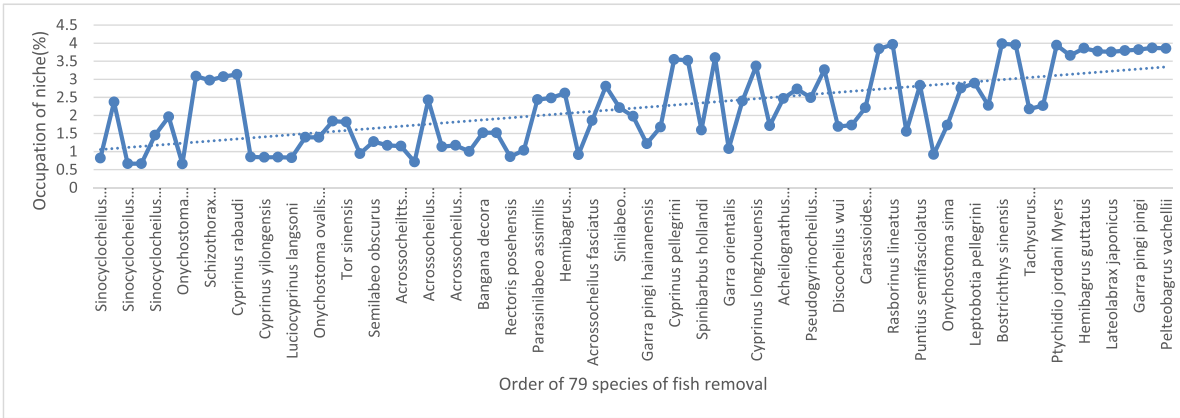


FIG. 4.39 – The niche of *Megalobrama skolkovii* changed in a zig-zag pattern as 79 fish species were removed from the community.

Megalobrama skolkovii Dybowski, *Squaliobarbus curriculus*, *Misgurnus anguillicaudatus*, *Pelteobagrus fulvidraco*, *Pseudolaubuca sinensis*, *Lucosoma chinensis*, *Ochetobius elongatus*, *Siniperca kneri*, *Cirrhinus molitorella*, *Hemiculter leucisculus*, *Channa maculata*, *Cyprinus carpio*, *Ancherythroculter lini*, and *Xenocypris argentea*.

Chapter 5

Fish Community Reconstruction and Case Study

The basic characteristics of a biological community include species diversity, community growth form and structure (*i.e.*, spatial structure, temporal composition, and species structure), dominant species (*i.e.*, species whose size, number, or activity have decisive effects on community characteristics), relative abundance (*i.e.*, the relative proportions of different species in a community), and nutrition structure. From small organisms to large animals and plants, a community of species is not a collection of many separate species; it is instead a complex ecosystem formed due to interaction mechanisms that include resource competition, nutritional symbiosis, quorum sensing, and gene transfer (Chen *et al.*, 2021; Cao and He, 2015; Cai *et al.*, 2002). The community is in a dynamic succession pattern that is constantly changing, and ecological processes can be understood from changes in species distribution, abundance, and spatial patterns (Tanner *et al.*, 1995).

In natural ecosystems, species are interdependent and are characterized by many weak interactions in addition to a few strong interactions (*e.g.*, the likelihood that one species will be consumed by another). In the food web system, the types of interaction between weak and strong communities are important for the permanence and stability of community formation (McCann *et al.*, 1998). Interactions are the bonds of community stability (Stachowicz, 2001). Fish species co-exist by adjusting their abundance, body size, and age at sexual maturity (Jennings *et al.*, 1998). Community composition and distribution are adaptive characteristics, formed over species evolution, that reflect the state of resource utilization (Wang *et al.*, 2006; Hanskii, 1999). Because of the unprecedented loss of biodiversity due to human activities, current community research focuses on species succession and changes in dominant species, as well as the functional role and maintenance of species in the ecosystem food chain, in order to reveal the relationship between ecological function and community niche (de Mazancourt, 2001).

5.1 River Ecosystems

Aquatic ecosystems can be divided into abiotic environments and biotic communities. The abiotic environment includes water bodies and their carriers, as well as sunlight, atmosphere, water, inorganic matter, and organic matter (*e.g.*, proteins, carbohydrates, lipids, and humus). The abiotic environment provides energy, nutrients, and space for living organisms. Every water body, including ponds, lakes, reservoirs, and rivers, is a water ecosystem.

Communities are classified according to their ecological functions into producers (*e.g.*, phytoplankton and aquatic vascular plants), consumers (*e.g.*, zooplankton, benthic animals, and fish), and decomposers (*e.g.*, bacteria and fungi); some bacteria also act as producers. Producers use the energy and nutrients of the abiotic environment to survive, while consumers and decomposers use producers in a food chain. According to the law of energy transfer, an ecological niche equilibrium can be realized among different biological levels. The distribution patterns of various organisms are the result of long-term adaptation and natural selection.

The number of water resources and terrestrial nutrients determines the state of river productivity. Environmental diversity affects the structure of basic organisms, while the structure of the food chain determines the function of the river ecosystem, the structure of the fish community, the distribution patterns of species diversity, and the construction mechanisms of the fish community. These factors contain many elements of the ecosystem as well as regional environmental characteristics. Understanding the mechanism of biological community construction from the perspective of the structural characteristics of species themselves is a new method in community ecology research. This study will provide different perspectives that may help to clarify ecosystem function, biodiversity maintenance, and functional community composition. The diverse patterns of species coexistence and the functional complementary mechanisms of niche overlap in fish communities are signs of mature communities and a stable systematic environment. Species attributes and environmental conditions together determine the niche allocation of the community (Mason *et al.*, 2008). In developing community systems, organismal dispersal capacity may determine the spatial extent of species distribution (Heino *et al.*, 2015), and ecosystems that are unstable are constantly disturbed at different frequencies and intensities. Therefore, natural communities are usually in a state of dynamic equilibrium. Community structure in river systems is determined by the spatiotemporal processes of species competition and niche allocation (Vanschoenwinkel *et al.*, 2010). An understanding of interspecific niche relationships is necessary in order to maintain ecosystem function and to ensure that ecosystem functions meet the needs of human development.

In addition to natural disasters, the factors that cause drastic changes in the river ecosystem include the obstruction of river connectivity, the smoothing of channels, the hardening of riverbeds, the crowding of the river space, water reductions, changes in hydrological characters, and water pollution. Changes in these factors are most likely to affect higher-order aquatic species, such as aquatic animals, as these organisms need spawning grounds for reproduction and aquatic plants.

Fish are particularly affected by these changes. As shown in the food analysis section, fish use primary phytoplankton, as well as high-grade plants, animals, humus, and almost all organic matter in aquatic ecosystems. Fish are the main biological group affecting the river ecosystem. The study of the functional mechanisms of fish community niche formation is of great significance to the construction and maintenance of fish communities. This work helps to ensure the optimal composition of river ecosystem functional groups and to achieve the goal of the functional guarantee of river ecosystems based on human needs.

5.1.1 River Structure

The total water reserves of the earth's hydrosphere are 1.386 billion cubic kilometers, most of which are held in oceans, rivers, lakes, reservoirs, marshes, and soils. Some of this water is in solid form in vast polar ice sheets, glaciers, snow packs, and frozen soils; a small amount of water exists as a vapor, primarily in the atmosphere. In total, 1.338 billion cubic kilometers of water are in the ocean, accounting for 96.5% of the total reserves, and only 2.53% of the total water reserves are freshwater. The water distributed on the continent is about half surface water and half groundwater. Surface water bodies are produced by rainfall, melting glaciers, and melting snow. During our planet's evolution, water flows have fluctuated from high to low, carving long, narrow concave channels in the earth's surface and forming a connected network. At the lower end of each channel is the mouth of a river. The water that eventually flows into the ocean is called the outflow river. Rivers that eventually flow into inland lakes or disappear into deserts are known as internal flow rivers.

The river is a complete continuum, consisting of upper and lower reaches as well as left and right banks. Most of the places of origin of rivers are springs, streams, glaciers, lakes, or marshes. In the middle reaches of a river, the slope is gradual and gentle, the river channel becomes wider, and there are often floodplains on both sides of the river. Changes in erosion and deposition are not obvious in the middle reaches, and the riverbed is relatively stable. The lower reaches of river channels are generally in plain areas. In this part of the river estuaries, the channel is broad, the riverbed slope and flow velocity are low, deposition is obvious, and shoals are abundant. Estuaries tend to form deltas due to the rapid deceleration of river flow and the associated sediment deposition. River width describes the transverse distance across a river and its adjacent vegetated area. The factors affecting river width include edge conditions, community composition, environmental gradients, and disturbances (including anthropogenic activities); these can affect adjacent ecosystems. Connectivity and width are important structural characteristics of the river ecosystem.

The structure of the river also includes the ecosystem. River morphological structure includes the biological species, the population sizes, spatial population patterns, temporal changes in the population, and the vertical and horizontal structure of the community. River nutritional structure, a functional unit that tightly links biotic and abiotic factors associated with nutrition, is composed of three

major functional groups: producers, consumers, and decomposers. There is a closed cycle of matter and energy flow between these functional groups and the environment.

5.1.2 River Ecological Function

The longitudinal zonation, water carrier, and flow of a river determine its ability to carry materials, such as nutrients and water, into an ecosystem. This transport function is part of the earth's material and energy cycle.

The continuity of the upper and lower reaches of the river, the complete spatial system between the left and right banks, and the moist range of the water flow from the inner habitat and the marginal habitat, which together determine the habitat function of the river ecosystem. Habitat is an area where plants and animals (including humans) can live, grow, feed, reproduce, and perform other important parts of the life cycle. The inner habitat is the direct environment for aquatic life. Water is an essential element for living things, in conjunction with elements such as carbon (C), hydrogen (H), oxygen (O), nitrogen (N), phosphorus (P), sulfur (S), potassium (K), magnesium (Mg), calcium (Ca), silicon (Si), iron (Fe), manganese (Mn), zinc (Zn), copper (Cu), Boron (B), Molybdenum (Mo), chlorine (Cl), sodium (Na), and nickel (Ni). The aquatic communities interact with the aquatic environment to form a dynamic equilibrium with a certain structure and function *via* the circulation of materials and the flow of energy.

Aquatic organisms primarily include bacteria, fungi, phytoplankton, zooplankton, benthic organisms, aquatic plants, and fish. The basic components of aquatic life are carbon, hydrogen, oxygen, nitrogen, phosphorus, sulfur, iron, zinc, and other elements. Carbon, hydrogen, and oxygen together account for more than 90% of the dry weight of plants and are the main components of plants. These elements exist in the form of various carbohydrates, such as cellulose, hemicellulose, and pectin, which are components of the cell wall. Plant organisms use photosynthesis to synthesize sugars from water nutrients and solar energy during growth, as well as to form active substances, such as certain celluloses and plant hormones. Carbon, hydrogen, and oxygen are also components of sugars, fats, and acids. Plants are primary producers. In the food chain, plants provide energy and growth elements for animals and in this way participate in the material and energy cycle of the ecosystem.

Organisms form the structure of the food chain, participate in the energy and material cycle, and maintain the balance of aquatic ecosystems. Phytoplankton, as producers, is at the first trophic level of the food chain, while phytoplankton-consuming zooplankton is at the second trophic level. Zooplankton consumers, such as small fish and shrimp, are at the third trophic level, and large fish are at the next trophic level. Humans that consume fish are participating in the material and energy cycle of the river ecosystem. The size of the catch depends largely on plankton production. Fish biomass can control the food chain of the aquatic ecosystem, regulate the structure of the food chain under the energy system, and help balance the ecosystem.

The food chain and the food web are complex nutrient relationships among species. A trophic level is the sum of all the species at a particular link in the food chain. For example, producers in the Chloroplastida and all autotrophs form the first trophic level at the beginning of the food chain. All of the animals that feed on producers, primarily chloroplastids, fall into the second trophic level, which can also be considered the herbivore trophic level. The third trophic level includes all carnivores that feed on herbivores. Logic dictates that a further fourth and fifth trophic level may exist (*e.g.*, second and third carnivore trophic levels). In an ecosystem, only about 10% of the energy at each trophic level is transferred to the next trophic level. As a result of the food relationship, living things are mutually restricted as the energy and nutrients produced by photosynthesis are transferred along the food chain. Only about 10% of the material and energy is transferred to the next trophic level. The remaining 90% is dissipated into the environment as heat. This is known as Lindemann's 10th law, which states that the rate of energy conversion between organisms is influenced by environmental conditions.

The productivity of water bodies and the transformation of bait organisms also differ among regions. Table 5.1 shows the P/B coefficients of various bait organisms in lakes and reservoirs in different regions of China (in a given water area, P = annual bait production, and B = annual average biomass). These data indicate the impact of the environment on water productivity and the utilization of organisms (see table 5.2).

However, the relationships among food chains within communities or ecosystems are complex, and most organisms, with the exception of producers and herbivores, tend to belong to more than one trophic level. The trophic level of many organisms varies with age and conditions.

TAB. 5.1 – P/B coefficients of different bait organisms across regional lakes and reservoirs.*

Region	P/B ratio				
	Phytoplankton	Zooplankton	Zoobenthos	Periphyton	Small fish and shrimp
North China	40–90	15–30	2–6	40–80	1.5–2.0
Central and eastern regions of China	100–1150	25–40	3–6	80–120	2.2–2.5
China plateau area	40–120	20–35	2–5	40–100	1.5–2.5
Southern China	150–200	30–40	4–8	100–120	2.0–2.5

*See SC/T 1149-2020.

TAB. 5.2 – Maximum utilization rates and bait coefficient for different ecological nutrient types.**

Bait type	Maximum utilization rate allowed (%)	Bait coefficient
Organic detritus	50	200
Phytoplankton	40	80
Zooplankton	30	10
Aquatic vascular plant	25	100
Zoobenthos	25	6
Periphyton	20	100
Small bait fish	20	4

**See SC/T 1149–2020.

The trophic level reflects species composition, functional level, the mode of material and energy flow and transfer in the ecosystem, and the environmental state. Community niche is closely related to trophic level. To form a functional community, organisms first must determine their niche based on a trophic cascade effect. Community species support ecosystem functions. There are many species at the same trophic level, which is of great significance for the construction of functional ecosystem communities.

5.1.3 Composition of the Fish Diet

Fish species, as consumers in aquatic ecosystems, are differentiated into herbivores, carnivores, and omnivores. This demonstrates the ecological dominance of fish. The composition of the fish community is closely related to the bait organisms in the aquatic ecosystem, and the fish community must be constructed based on the characteristics of the food chain in the environment. There are many bait organisms in rivers, but it is difficult to identify bait species from the digestive tract because bait organisms are digested quickly and do not leave distinguishable remains. Figure 5.1 shows the dietary composition of 10 species of fish in the 1980s. The fish species included are *Tenualosa reevesii* (Richardson, 1846), *Clupanodon thrissa* (Linnaeus, 1758), *Coilia grayii* (Richardson, 1845), *Coilia mystus* (Linnaeus, 1758),

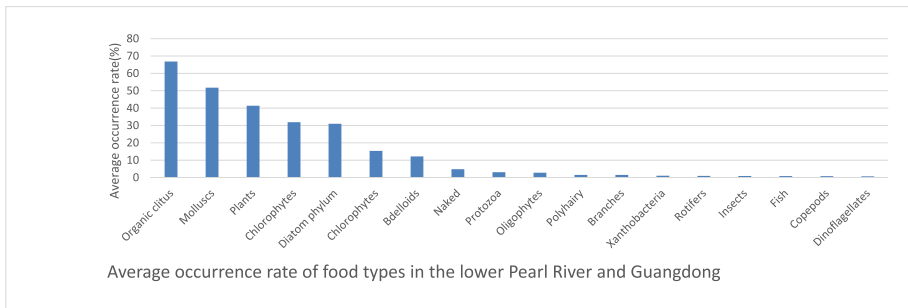


FIG. 5.1 – Components of the diets of 10 fish species in the Pearl River.

Mylopharyngodon piceus, *Ctenopharyngodon idella*, *Squaliobarbus curriculus*, *Erythroculter pseudobrevicauda* (Nichols, 1927), *Megalobrama terminalis* (Richardson, 1846), and *Parabramis pekinensis*. The diets of these fish include many species of zooplankton, benthic animals, aquatic plants, and other fish.

An analysis of gut contents of *Megalobrama terminalis* from the lower reaches of the Pearl River and the associated river network was performed. Although the identification of consumed food items was complicated by incomplete digestive residues and morphological ambiguity, diatoms, green algae, Cyanophyta, Gymnophyta, dinoflagellates, Xanthophyta, polychaetes, oligochaetes, protozoans, rotifers, Cladocera, Copepoda, insects, leeches, aquatic plants, and fish were detected in the gut samples. Figure 5.2 demonstrates that fish consume a wide range of foods, that fish are picky eaters, that the dietary composition of the same fish is different in different waters and that fish feed on different items based on environmental conditions. The results showed that the main foods of *Megalobrama terminalis* were organic detritus (about 60%–70%), mollusks (about 35%–70%), plants (about 14%–57%), and zooplankton (about 13%–55%) (Yuguo Xia *et al.*, 2017).

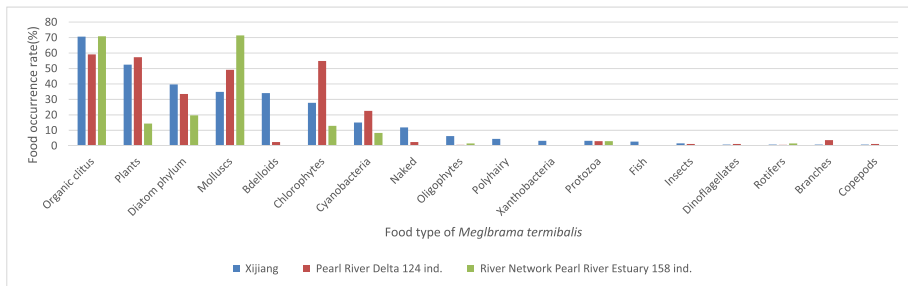


FIG. 5.2 – Frequency of food items detected in the intestines of *Megalobrama terminalis*.

Individual *Megalobrama terminalis*, including small juveniles, large juveniles, sub-adults, and adults, were collected from the middle and lower reaches of Pearl River for analysis of intestinal food samples. The weight percentage of residue (*i.e.*, a recognizable substance in the digestive tract) in the intestine with respect to average biomass was more than 84%. Many types of juvenile prey were consumed. In addition to the residues, the most commonly consumed taxa were the Chaetophorales (5.5% W, W = the weight percentage of residue), followed by the Cirsidiales (0.2% W) and crustaceans (0.2% W). Table 5.3 shows that there were no significant differences in the proportions of intestinal contents among groups and that the percentage of detritus consumed as the main food item changed little among groups. However, the percentage of prey items varied greatly among groups (see table 5.3; Yuguo Xia *et al.*, 2020).

The small juveniles fed mainly on prey in the Myrosiales, Chaetophorales, Chlorophyta, and Sarcoidoidea; these groups accounted for 87.1% of the total intestinal contents. Large juveniles fed mainly on residues, mussels, Chaetophorales,

TAB. 5.3 – Identifiable black amur bream prey in each size group sorted by taxa. Items with a percentage by weight over 0.1% and a percentage by number over 1% are listed. Percentage by number excludes detritus.

Food item	Small juvenile (N = 9)	Large juvenile (N = 6)	Sub-adult (N = 15)	Adult (N = 7)
Percent by weight % (%W, mean \pm SD)				
Detritus	94.1 \pm 16.4	84.5 \pm 33.3	96.8 \pm 7.0	92.7 \pm 14.8
Chaetophorales	5.5 \pm 16.4	1.4 \pm 3.0	0	0
Mytiloida	0	13.8 \pm 33.9	3.1 \pm 7.1	7.2 \pm 14.9
Coscinodiscales	0.2 \pm 0.2	0.02 \pm 0.04	0.08 \pm 0.15	0.05 \pm 0.08
Araphidiales	0.2 \pm 0.5	0.05 \pm 0.12	0.04 \pm 0.08	0.01 \pm 0.01
Ulvaes	0	0.1 \pm 0.3	0	0
Percent by number % (%N, mean \pm SD)				
Coscinodiscales	53.6 \pm 39.3	9.4 \pm 22.6	45.0 \pm 32.6	50.2 \pm 36.7
Chaetophorales	12.5 \pm 35.3	43.4 \pm 49.1	0	0
Chlorococcales	11.7 \pm 26.0	0.1 \pm 0.3	16.0 \pm 29.7	14.4 \pm 21.1
Mytiloida	0	16.7 \pm 40.8	0.02 \pm 0.03	0.02 \pm 0.04
Araphidiales	5.9 \pm 12.6	2.4 \pm 5.8	11.7 \pm 11.2	6.7 \pm 7.9
Chroococcales	0.1 \pm 0.3	0	0	26.2 \pm 38.7
Osillatoriales	1.0 \pm 2.9	0	9.9 \pm 17.5	0
Ulvaes	0	6.1 \pm 13.5	0	0
Biraphidinales	4.4 \pm 6.5	21.8 \pm 36.0	15.5 \pm 30.3	1.1 \pm 1.7
Aulonorphidinales	9.3 \pm 20.5	0	0.5 \pm 1.5	0.4 \pm 0.6

*(Yuguo Xia *et al.*, 2020).

and *Ulva*. The weight of intestinal residues and mussels in the adult population was 92.7% and 7.2%, respectively. In the large juveniles, the main items consumed were Chaetophorales, accounting for 43.4% of the total, while the subadults preyed mainly on Cirsidiales, accounting for 45.0% of the total, in addition to small amounts of Pycinidae and mussels. In the adult population, the main feeding targets were Coscinodiscales (50.2%). Also found were a few shell sutures, double shell sutures, tube shell sutures, and mussels.

In a water body, fish food sources are diverse. Analysis of the feeding habits of *Megalobrama terminalis* at different growth stages showed that the dietary composition of fish differs among growth stages. The results showed that fish trophic levels vary across developmental stages and productivity structures. This complexity must be carefully considered when constructing fish systems during ecosystem restoration. Fish use a certain range of food resources, and species communities with more fish can utilize diverse food sources efficiently.

The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ contents in potential food source samples ranged from 31.00‰ to 13.07‰ and from 2.87‰ to 17.35‰, respectively. C4 plants on the riverbank had the highest $\delta^{13}\text{C}$ levels and the lowest $\delta^{15}\text{N}$ levels, with an average of 13.29 ± 0.21 ‰ and 4.06 ± 1.44 ‰, respectively. The lowest levels of $\delta^{13}\text{C}$ and the highest levels of $\delta^{15}\text{N}$ were found in *Corbicula fluminea* and *Macrobrachium nipponense*, respectively. The $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ levels in other potential food groups

varied greatly, with some degree of overlap. The means and standard deviations of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ for each group and a potential food source are shown in table 5.4. Among the potential food sources, c/n (% by weight, c = total carbon in food, n = total nitrogen in food) ranged from 3.17% to 77.32%, with the value of *Macrobrachium nipponense* being the lowest and the value of river bank C4 plants being the largest (0.02%–0.15%; see table 5.5). The contribution of 12 potential food species to the small-sized larvae ranged from 0.05% to 0.10%, the highest of which was *Corbicula fluminea*, and bank C4 plants were the lowest. Other contributions to juveniles were similar. In all groups, the contribution of the riparian C4 plants was low, while the contributions of zooplankton, *Macrobrachium nipponense*, and *Corbicula fluminea* were high. These taxa are very important to the *Megalobrama terminalis* population. The contributions of *Macrobrachium nipponense* and *Corbicula fluminea* to the growth of *Megalobrama terminalis* also increased between juveniles and adults; this was the result of dietary changes during fish growth (see table 5.4).

The total breadth of the nutrient niche was assessed using total niche width (TNW) to determine enteral nutrients and using corrected standard elliptic area (SEAC) to identify stable isotopes. Higher values corresponded to a greater niche width in a given group. The results of both analyses were similar and showed that the width of the nutrient niche increased with body length. The TNW and SEAC analyses found the highest values in the adult group, followed by the sub-adults, large juveniles, and small juveniles (see table 5.6).

During growth, juvenile fish have been observed to prefer zooplankton, while adult fish devour more animal prey. Many previous studies in the Pearl River Delta have shown that *Megalobrama terminalis* mainly feeds on detritus. Xia *et al.* (2017) used 18S rDNA sequencing to analyze the contents of the gut and found that juvenile *Megalobrama terminalis* had abundant plant ingredients, while adults had abundant benthic animals. Stable isotopes can be used to determine carbon sources and nutrients (Carreonmartinez and Heath, 2010), but it is preferable to detect specific nutrient interactions because the isotope values of potential prey often overlap (Hardy *et al.*, 2010). Stable isotope analysis showed that *Megalobrama terminalis* can use almost all food types (table 5.6); however, the different populations at different growth stages differ in specific bait utilization, and the results were consistent with the intestinal inclusion analysis. Isotopic analysis of gut inclusions identified far fewer species compared to microscopy and molecular identification (Xia *et al.*, 2017). Isotope approaches can support tracking the circulation of three foods *in vivo* (Peterson *et al.*, 1986), but they cannot yet address more than three foods (Post, 2002; Peterson and Howarth, 1985; Peterson *et al.*, 1985; tables 5.5 and 5.6).

Analysis of the dietary composition of fish species in Zhaoqing River showed that silver carp in this river section mainly feed on phytoplankton, and four groups of algae [*Melosira granulata* (Ehrenberg) Ralfs, 1861; *Melosira granulata* Her; *Melosira varians* (von Stosch, 1951); *Synedra ulna*] accounted for about 79% of the diet; detritus was not included. In the Zhaoqing river section in Guangdong, *Megalobrama terminalis* and *Cirrhinus molitorella* mainly fed on diatoms and benthic animals, while *Hypophthalmichthys nobilis* primarily consumed algae and benthic animals.

TAB. 5.4 – Summary statistics (mean \pm SE) of $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and C/N in the different size groups of *Megalobrama terminalis* and potential prey sources in the sampling site. Values are mean \pm SD.

Group/taxon	Code	N	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	C/N
Small juvenile	Small juvenile	11	-24.67 ± 1.75	11.60 ± 2.75	–
Large juvenile	Large juvenile	12	-25.53 ± 1.43	12.54 ± 2.44	–
Sub-adult	Sub-adult	15	-26.10 ± 1.52	13.53 ± 2.69	–
Adult	Adult	8	-25.58 ± 1.33	14.57 ± 3.09	–
Zooplankton	Zooplankton	2	-30.25 ± 1.07	9.95 ± 2.08	6.11 ± 0.66
Phytoplankton	Zooplankton	2	-27.94 ± 0.24	8.21 ± 1.13	8.40 ± 1.86
Riparian C ₄ plants	C ₄ _P	4	-13.29 ± 0.21	4.06 ± 1.44	63.89 ± 10.35
<i>Potamogeton</i> sp.	Psp	6	-25.29 ± 3.26	7.45 ± 2.43	10.46 ± 1.26
<i>Macrobrachium nipponense</i>	Mni	9	-26.97 ± 0.66	15.95 ± 1.32	3.30 ± 0.07
<i>Anodonta woodiana</i>	Awo	2	-24.72 ± 0.65	6.46 ± 0.04	3.84 ± 0.06
<i>Limnoperna fortunei</i>	Lfo	1	-26.53	4.96	4.65
<i>Semisulcospira cancellata</i>	Sca	1	-24.16	9.51	3.91
<i>Corbicula fluminea</i>	Cfl	4	-30.27 ± 0.26	11.99 ± 0.31	4.65 ± 0.43
<i>Bellamyia</i> sp.	Bsp	4	-22.28 ± 0.34	4.42 ± 1.02	4.14 ± 0.27
Benthic detritus	Bde	2	-26.11 ± 1.20	7.08 ± 1.50	12.57 ± 4.50
Sediment	Sediment	3	-25.23 ± 0.39	5.85 ± 0.23	10.47 ± 1.23

*(Yuguo Xia *et al.*, 2020).

TAB. 5.5 – Contributions of different potential prey taxa to different size groups of *Megalobrama terminalis*, according to stable isotope Bayesian mixed models. CI95%, lower – higher confidence intervals.

Prey	Small juvenile		Large juvenile		Sub-adult		Adult	
	Mean	CI95%	Mean	CI95%	Mean	CI95%	Mean	CI95%
Zooplankton	0.09	0–0.18	0.11	0–0.21	0.13	0–0.25	0.11	0–0.21
Phytoplankton	0.08	0–0.17	0.09	0–0.19	0.09	0–0.20	0.09	0–0.19
C ₄ _P	0.05	0–0.11	0.03	0–0.07	0.02	0–0.06	0.03	0–0.08
Psp	0.09	0–0.17	0.08	0.01–0.18	0.07	0–0.18	0.08	0–0.19
Mni	0.09	0–0.17	0.11	0–0.20	0.14	0.03–0.25	0.15	0.02–0.27
Awo	0.08	0–0.17	0.07	0–0.17	0.05	0–0.14	0.06	0–0.15
Lfo	0.09	0–0.17	0.08	0–0.17	0.06	0–0.15	0.06	0–0.16
Sca	0.08	0–0.17	0.07	0–0.17	0.06	0–0.15	0.07	0–0.16
Cfl	0.10	0–0.18	0.13	0.01–0.23	0.15	0.01–0.29	0.12	0.01–0.23
Bsp	0.08	0–0.16	0.06	0–0.14	0.04	0–0.10	0.05	0–0.13
Bde	0.08	0–0.17	0.08	0–0.18	0.09	0–0.20	0.09	0–0.19
Sediment	0.08	0–0.17	0.08	0–0.18	0.09	0–0.20	0.09	0–0.19

*(Yuguo Xia *et al.*, 2020).

TAB. 5.6 – Metrics quantifying trophic niche and individual specialization in *Megalobrama terminalis*. Same superscript letters indicate no significant differences (Kruskal–Wallis test, $p > 0.05$).

	Small juvenile	Large juvenile	Sub-adult	Adult
Taxonomic richness per gut (mean \pm SD)	5.5 \pm 3.0 ^a	2.8 \pm 1.5 ^a	4.8 \pm 2.2 ^a	4.8 \pm 1.6 ^a
TNW	0.058	0.699	0.862	0.938
WIC/TNW	0.250	0.951	0.637	0.610
Diet similarity	0.339	0.185	0.375	0.386
NR (‰)	9.49	7.77	10.07	9.56
CR (‰)	5.54	4.80	6.13	3.48
CD (‰)	2.61	2.40	2.47	2.55
MNND \pm SD (‰)	1.04 \pm 1.07	1.37 \pm 0.62	1.15 \pm 1.10	1.69 \pm 2.05
TA	19.22	21.74	30.87	17.52
SEA (‰)	9.446	10.041	11.240	12.957
SEA _c (‰)	10.496	11.045	12.104	15.116

*(Yuguo Xia *et al.*, 2020).

The energy stored in organic matter in an ecosystem is carried through the food chain layer by layer. The food chain can be subdivided into the predator food chain, scavenger food chain, and parasitic food chain based on the relationships among organisms. The structure of the food web is related to species diversity, and the loss or extinction of key species greatly affects the structure of the food web (Dunne *et al.*, 2002). Intensive exploitation of fish communities often results in a significant reduction in the abundance of targeted species, affecting the structure and stability of the entire ecosystem. Intensive exploitation of fish communities often results in a significant reduction in the abundance of key species, affecting the structure and stability of the entire ecosystem. This reduction of key species is also reflected in changes in the average nutrient levels of fish communities. Declines in average nutrient levels are due to decreases in the number of large fish-eating species and increases in the number of smaller pelagic species foraging at lower nutrient levels. The decline in spawning stock biomass of traditional target species is associated with intensive fishing and long-term climate variability. In general, a decrease in the average trophic level of the entire fish community may enable the system to maintain a high fishery yield (Pinnegar *et al.*, 2002).

5.1.4 Fish Trophic Levels

The earth's biosphere is a large ecosystem. The species, abundance, biomass, life history, and spatial distributions of organisms in the system must follow the laws of energy flow and material circulation. Biotic communities are not only adapting to the environment in the ecosystem but also changing the appearance of the surrounding environment in the ecosystem. Various growth factor materials closely link biological

communities together. There are more than 15 000 species of freshwater fish worldwide (Fishbase, 2021; <https://www.fishbase.de/2021,0810>). The composition of the fish community differs among river ecosystems. The trophic level of the fish community reflects the state of the river ecosystem. In the food chain of the ecosystem, fish species with common functional attributes at the same level can be classified into a single niche type, such as predatory niche-type fish. This classification can simplify the complicated relationships among the species in the food web and help to guide studies of the mechanisms of fish community construction and the reconstruction of fish communities in the aquatic ecosystem. The food web is also implicated in a number of other processes related to community structure, including the regulation of habitat heterogeneity, changes in community function over time with respect to the environment, productivity gradients, the direct and indirect cascade effects of predation, intra-row predation, indirect reciprocity, species competition, ecosystem stability, and nutrient dynamics. Food web analyses cannot be reduced to a few linear “trophic levels” and thus must also consider trophic cascades, as well as top-down/bottom-up mechanisms of community regulation (Polis *et al.*, 1996a; Winemiller and Polis, 1996).

The mass of existing organic matter in a community is its biomass, and the biomass and biomass accumulation rates of different types of communities differ noticeably. The biological composition of the community includes the food chain, which progresses from plants to herbivores to carnivores. Due to various types of energy consumption, the productive forces gradually decrease. Primary productivity only accounts for 0.1%–1% of solar energy, while secondary productivity represented by animals only accounts for 10% of the former.

Each kind of living creature obtains the energy needed for survival, growth, and reproduction in a unique way. There are complex food relationships among the organisms in an ecosystem; fish can be divided into herbivores and carnivores according to their feeding habits. There may be two to five trophic levels. In one system, with the exception of fish at high trophic levels (more than 4.5), the trophic levels of other species were not obviously regular. This may be partially due to variability in the fish diet; that is, the fish diet may change independently according to environmental conditions to most efficiently obtain energy. Alternatively, it may be that the fish themselves have the ability to access different food sources, and the artificial trophic level classification results in a lack of comprehensive understanding of the ways in which fish obtain energy in the ecosystem.

Two species may be competitive or symbiotic, depending on their mutual interests. Relationships may be parasitic, partially symbiotic, or mutually beneficial. If two species use the same resource (niche overlap), they will compete and one species will be excluded. However, if the resource demands of one species change (niche differentiation), the two species may coexist. The longer a community has evolved, the more favorable the environment is and the more stable the species are. As more species are included in the community, the trend of community development is toward niche differentiation and species increase. The diversity of community species structures indicates that the complexity of community structure increases with the resources used in the ecosystem. Complex communities have more ecological niches, the competition among the various organisms in the community is

relatively weak, and the community structure is relatively stable. Therefore, the species structure of the community reflects the environmental characteristics of the ecosystem, and the organisms in a community are in constant interaction.

When the total primary productivity of a community is greater than the total community respiration, and the net primary productivity is greater than ingestion by animals and decomposition by microorganisms, organic matter accumulates. The community then grows until it reaches a mature stage, where accumulation ceases and production and respiratory consumption are balanced. This process is called succession, and the final stage of succession is called a climax community. The productivity of the climax community is not equivalent to maximum productivity, but the biomass is maximized and the net ecosystem production is very low or even zero. In a climax community, species diversity may decrease, but the community is maximally complex and stable. Communities do not have the same processes of genetic regulation and humoral integration as individuals, and the path of succession is entirely determined by interspecies interactions and the balance of material and energy flow. Therefore, the characteristics of climax communities depend on both the environmental conditions and the species.

The trophic level depends on species composition, functional level, mode of material and energy flow and transfer in the ecosystem, and environmental state. Community niche is closely related to trophic level. To form a functional community, the organism niche must first be determined based on the trophic cascade effect. As shown in table 5.7 (Fishbase, 2021; <https://www.fishbase.de/2021,0810>), freshwater fish generally have a trophic level of approximately 2–4.7. The data from some regions suggest that the average trophic level of fish communities in freshwater ecosystems may be 3.0–3.8. The average trophic level can reflect the trends in changing group compositions and trophic levels in the ecosystem.

TAB. 5.7 – Average trophic level of fish in some inland waters (Fishbase, 2021; <https://www.fishbase.de/2021,0810>).

	Average nutritional level
Azores	3.7991
New Zealand	3.2985
Australia	3.2714
France	3.263
USA (North America)	3.1875
Philippines	3.1838
South Africa	3.1827
Cambodia	3.1669
Zimbabwe	3.1633
Thailand	3.1524
Malaysia	3.139
Vietnam	3.1061
Angola	3.0611
Hawaii (USA)	3.0537
China	3.0374

For example, the fish are shown in table 5.8 (Fishbase, 2021; <https://www.fishbase.de/2021,0810>) were classified in increments of 0.5. Analysis of the proportion of freshwater fish at each trophic level in the community showed that the functions of species in different regions were plastic. The trophic level composition provides insight into the function of species diversity within the ecosystem.

TAB. 5.8 – The nutritional grade composition (%) of some fish in inland waters.

%	4.51–	4.0–4.5	3.5–3.99	3.0–3.49	2.5–2.99	2–2.49
China		2.3	7.0	49.9	27.6	13.3
Australia		7.1	18.8	46.8	22.1	5.0
Cambodia		4.7	25.2	37.3	17.3	15.6
France		13.3	9.2	49.0	21.4	7.1
Angola		3.7	10.0	46.8	24.4	15.1
Azores	57.1	14.3	28.6			
Hawaii (USA)	1.7	5.0	21.7	25.0	20.0	26.7
Malaysia		3.8	18.6	44.2	19.7	13.8
New Zealand		12.9	9.7	64.5	6.5	6.5
Philippines		6.0	22.2	39.5	19.8	12.6
South Africa	0.6	3.4	22.7	42.0	20.5	10.8
Thailand		4.1	18.5	46.3	18.6	12.5
Vietnam		2.6	17.1	43.9	22.0	14.3
Zimbabwe		6.6	19.2	38.4	22.5	13.2
USA (North America)	0.1	5.2	8.5	56.6	24.8	4.8

Of the 1581 freshwater fish recorded in China, about 1225 species (accounting for 78% of the total) have trophic levels ranging from 2.5 to 3.49 (see table 5.9), with omnivorous fish being the most common.

TAB. 5.9 – Nutritional grade distributions of major fish species in China.

Nutritional level range	Species	%
4.0–4.5	36	2
3.5–3.99	110	7
3.0–3.49	789	50
2.5–2.99	436	28
2–2.49	210	13

Xiong *et al.* (2015) found that the trophic levels of fish in Yangtze Plain floodplain lakes were 2.0–4.2, with a mean trophic level of about 3. Nearly 40% of the fish in the Yangtze Plain assemblage were omnivorous fish, which is common in aquatic food webs, especially in fish communities (Zhang *et al.*, 2013; Vanni *et al.*, 2005; Drenner *et al.*, 1996; Diehl, 1992). The results of GAM analysis showed that

only fish at high and low trophic levels had relatively stable morphological characteristics; at medium trophic levels, there was not sufficient evidence of speciation. Ba *et al.* (2015) found that the trophic levels of the main biological resources in the middle reaches of the Yangtze River were between 2.42 and 4.88, but that the trophic levels of most resources were 2.83–3.61. The average trophic level of fish was 3.28, and 80.85% of all species had trophic levels greater than 2.83. This indicated that the fish community structure differed in different environments within the same river.

Ye *et al.* (2021) analyzed the trophic level of Yangcheng Lake and found that the fish community's level was 2.12–3.75 in the summer. The fish with the lowest trophic level was *Xenocypris microlepis*, while the fish with the highest trophic level was *Taenioides cirratus* (Blyth, 1860). The trophic level range was 2.56–3.71 in the autumn, with *Hypophthalmichthys nobilis* and *Culter dabryi* having the lowest and highest trophic levels, respectively. The range of fish trophic levels in the spring was 2.03–3.98, with *Ctenopharyngodon idella* and *Cultrichthys erythropterus* having the lowest and highest trophic levels, respectively. These results suggest that trophic levels also differ among water bodies. Xu *et al.* (2020) analyzed the nutritional structure of the aquatic animal food network in the Yangtze estuary and found that trophic levels ranged from 2.0 to 4.0; the trophic levels of the 12 dominant fish were lower than was previously calculated in the 1990s, with decreases from 3.80 to 2.87. In addition, the number of species and the abundance of fish at high trophic levels decreased. For example, the trophic levels of *Lateolabrax japonicus* decreased by 1.28. Finally, the proportion of low-grade carnivorous fish decreased from 20.0% to 18.6%. The results indicated that the trophic levels of a single species might change depending on the food source during environmental change.

The trophic levels of Pearl River fish ranged from 2 to 4.5, with an average trophic level of 3.40 (see table 5.10). The average trophic level of estuarine and migratory species was 3.54, and that of exotic species was 2.94. The trophic level of estuarine fish was the highest, followed by the average trophic level of native freshwater fish. The trophic level of exotic species was relatively low. This suggested that the niche of fish at low trophic levels in Chinese rivers may be vacant and that there is an opportunity for alien species to fill this niche.

Historical records indicate that the 16 fish species with trophic levels between 2 and 2.5 in the Pearl River have not changed. Table 5.10 showed that there were 10 species at the same trophic level that were invasive alien species in the community. We speculate that there are two explanations for this finding: First, the biomass of indigenous fish on the same trophic level may be insufficient, resulting in a loss of niche space and allowing the alien species to enter the niche. Thus, the invasive species coexist with the indigenous fish in the same region. Alternatively, fish in the same niche in local areas of linear rivers may have been lost, similarly giving alien species the opportunity to fill the niche. The trophic level of *Ctenopharyngodon idella* and *Hypophthalmichthys molitrix* was 2.00, and the catch output of the “four big fish” in the middle and lower reaches of the Pearl River was more than 40%

greater than historical catches, mainly due to catches of *Ctenopharyngodon idella* and *Hypophthalmichthys molitrix*. In aquatic ecosystems, productivity and nutrients cannot be exported through the food chain, and the system requires fish at lower trophic levels.

Some scholars argue that conservation efforts can be strongly complementary to all types of diversity and that systematic conservation planning is an important means of allocating limited resources (Strecker *et al.*, 2011). It has been suggested that in restoring or reconstructing ecosystem species communities, it is possible to optimize the energy output for species collocation.

Material and energy in river ecosystems are transmitted step by step from autotrophic groups to heterotrophs and eventually to fish; the quality of nutrients brought in by the water cycle determines the basic biomass of the system. Over evolutionary time, organisms adapt to the environment and form fixed types of communities; community types can be identified based on the species composition of the biological system. Currently, the average daily output of fish in the Guangdong section of the Pearl River system is 189.2 tons, including 30 species (classes) of fish, with an average nutritional level of about 3.1 (see table 5.11). Fish with biomass $\geq 1\%$ include *Pseudolaubuca sinensis*, *Hemiculter leucisculus*, *Megalobrama terminalis*, *Tilapia* spp., *Cyprinus carpio*, *Hypophthalmichthys molitrix*, *Cirrhinus molitorella*, *Ctenopharyngodon idella*, *Hypophthalmichthys nobilis*, *Squaliobarbus curriculus*, *Pelteobagrus fulvidraco*, and *Culter alburnus*. About 66.3% of all fish species have relative biomass values between 1% and 20%, with an average nutritional level of 2.73. Analysis of the nutrient levels of exported fish products indicates that the proportion of the low-nutrient-level fish in the waters where the aquatic products were produced was relatively small, and it was inferred that the utilization of the primary productivity of the water body was insufficient. The annual runoff volume of the Pearl River is about 330 billion cubic meters, and the annual runoff volume of 330 billion cubic meters can export about 165 000 tons of fish products per year; the water body (runoff) was calculated to export 0.5 g/cubic meter. From 2016 to 2018, only about 40 000–60 000 tons of fish were caught from the river system every year, suggesting the lack of fish in river ecosystems. The food chain system should be restored and ecosystem function should be repaired by recovery starting with the fish with low trophic levels.

The average daily output of fish over three years in Guangxi was 222.0 tons and included 29 species (classes) of fish, with an average nutritional level of about 3.1. The fish with biomass values $\geq 1\%$ included *Tilapia* spp., *Cyprinus carpio*, *Hemiculter leucisculus*, *Hypophthalmichthys nobilis*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, *Squaliobarbus curriculus*, *Pelteobagrus fulvidraco*, *Lucosoma chinensis*, *Carassius auratus auratus*, and *Cirrhinus molitorella*. Species comprising about 79.2% of the total fish biomass contributed 1.3% and 22% to the total biomass and had an average nutritional level of 2.55 (see table 5.12).

TAB. 5.10 – Trophic level distributions of fish in the Pearl River.

Nutritional level range	$2 < x \leq 2.5$	$2.5 < x \leq 3$	$3 < x \leq 3.5$	$3.5 < x \leq 4$	$4 < x \leq 4.5$	Mean trophic level/species
Species number	33	20	176	90	35	3.40/354
No. alien species	23	20	173	86	33	3.42/335
Native species	16	6	83	20	6	3.24/131
Estuary and migration type	7	13	90	66	27	3.54/204
Exotic species	10		3	4	2	2.94/19

TAB. 5.11 – Sample river fish varieties captured using ships in Guangdong Province from 2016 to 2018.

Species	Trophic level	Total output over three years	Proportion of total fish %
<i>Siniperca kneri</i>	4.5	393.9	0.190121097
<i>Silurus asotus</i> (Linnaeus, 1758)	4.4	754	0.36
<i>Clarias gariepinus</i> (Burchell, 1822)	4.4	1	0.00
<i>Ictalurus punctatus</i> (Rafinesque, 1818)	4.4	17	0.01
<i>Siniperca scherzeri</i> (Steindachner, 1892)	3.9	7	0.00
<i>Hemibagrus guttatus</i>	3.7	62.84	0.03
<i>Monopterus albus</i> (Zuiew, 1793)	3.6	22	0.01
<i>Cranoglanis boudierius</i> (Richardson, 1846)	3.5	154.6	0.07
<i>Pelteobagrus fulvidraco</i>	3.5	2742.75	1.32
<i>Elopichthys bambusa</i>	3.5	41.8	0.02
<i>Culter alburnus</i>	3.4	2259.4	1.09
<i>Culter recurviceps</i>	3.3	13.95	0.01
<i>Megalobrama terminalis</i>	3.3	12618.45	6.09
<i>Lcucosoma chinensis</i>	3.2	1439	0.69
<i>Pseudolaubuca sinensis</i>	3.2	43361.5	20.93
<i>Mylopharyngodon piceus</i>	3.2	214.05	0.10
<i>Pseudohemiculter dispar</i>	3.2	563.5	0.27
<i>Cyprinus carpio</i>	3.1	8776.2	4.24

TAB. 5.11 – (continued).

<i>Channa maculata</i>	3	85.5	0.04
<i>Hypophthalmichthys nobilis</i>	2.8	5991.5	2.89
<i>Mastacembelus armatus</i> (Lacepede, 1800)	2.8	43.55	0.02
<i>Hemiculter leucisculus</i>	2.8	21767	10.51
<i>Squaliobarbus curriculus</i>	2.7	3700.05	1.79
<i>Tilapia</i>	2	11396.5	5.50
<i>Cirrhinus molitorella</i>	2	8385.5	4.05
<i>Hypophthalmichthys molitrix</i>	2	8732.45	4.21
<i>Carassius auratus auratus</i>	2	651.7	0.31
<i>Ctenopharyngodon idella</i>	2	7581.1	3.66
<i>Parabramis pekinensis</i>	2	1226	0.59
Trash fish	3.1	64179.95	30.98
			100.00

TAB. 5.12 – Sample river fish varieties captured using ships in Guangxi from 2016 to 2018.

Species	Trophic level	Average of species (%)
<i>Siniperca kneri</i>	4.5	0.8
<i>Silurus asotus</i> (Linnaeus, 1758)	4.4	0.6
<i>Clarias gariepinus</i> (Burchell, 1822)	4.4	0
<i>Plecoglossus altivelis</i> (Temminck & Schlegel, 1846)	4.2	0
<i>Siniperca scherzeri</i> (Steindachner, 1892)	3.9	0.1
<i>Hemibagrus guttatus</i>	3.7	0.7
<i>Pelteobagrus fulvidraco</i>	3.5	3.5
<i>Elopichthys bambusa</i>	3.5	0
<i>Culter alburnus</i>	3.4	0.2
<i>Megalobrama terminalis</i>	3.3	0.4
<i>Culter recurviceps</i>	3.3	0
<i>Lcucosoma chinensis</i>	3.2	2.5
<i>Mylopharyngodon piceus</i>	3.2	0.9
<i>Cyprinus carpio</i>	3.1	13
<i>Hemiculter leucisculus</i>	2.8	10.2
<i>Hypophthalmichthys nobilis</i>	2.8	7.3
<i>Mastacembelus armatus</i>	2.8	0.6
<i>Squaliobarbus curriculus</i>	2.7	5.2
<i>Xenocypris argentea</i>	2.6	0
<i>Cirrhinus mrigala</i> (Hamilton, 1822)	2.2	0.7
<i>Labeo rohita</i> (Hamilton, 1822)	2.2	0.3
<i>Tilapia</i>	2	21.7
<i>Ctenopharyngodon idella</i>	2	7
<i>Hypophthalmichthys molitrix</i>	2	6.1
<i>Carassius auratus auratus</i>	2	1.4
<i>Cirrhinus molitorella</i>	2	1.3
Trash fish	3.057	15.2

5.2 Molecular Mechanisms of Energy Utilization and Ecological Niche

Niche is a reflection of the interspecific relationships within communities and the food chain established by certain basic organisms, such as fish and invertebrates in aquatic ecosystems; niche also indicates habitat characteristics (Green, 1971). Living things and the environment comprise the river ecosystem. Freshwater ecosystems comprise only about 1% of the earth's surface but provide rich biodiversity and irreplaceable ecosystem services, such as drinking water and aquatic products. However, due to population growth, the rapid development of the social economy, and the unreasonable exploitation and utilization of biological resources, the ecological environment has been seriously damaged. The degradation of ecosystem function is manifested by changes in biodiversity and damage to species

communities. Major causes of the rapid decline in freshwater ecosystem function include dam constructions that block fish movement, loss of fish spawning grounds due to reservoir formation, overfishing, deterioration of water quality, eutrophication, and the invasion of alien species (Shuai *et al.*, 2017a, 2017b).

The molecular mechanisms of energy utilization are those processes of community niche differentiation, species development, evolution, and functional community construction that aim to obtain energy. Solar radiation is the basic energy source on the surface of the earth and is the main energy source influencing changes in ecosystems and biological communities. Biological organisms convert solar energy into biological energy, which can be transferred to other organisms through the food chain, supporting growth. Algae and plants produce energy from the sun using photosynthesis, while bacteria convert chemical energy into biological energy. Biological energy is fixed at about 1.55×10^{11} tons of carbon per year (Zhou Liangjun, 1986), or about 4.5×10^{21} j. Species aim to obtain energy. Since the emergence of unicellular organisms hundreds of millions of years ago, energy competition has been a factor in interspecies relationships. For example, plant organisms differentiated into trees, shrubs, and herbs to better compete for and utilize energy in “three-dimensions.”

The energy that supports the activities of animal life comes from the food chain, which is based on the energy of plant origin. Animals differentiate into species with different feeding habits and form biological communities containing diverse species, rich food chain levels, and perfect energy circulation. Each species establishes its niche with the goal of maximizing energy. Thus, the mechanisms used in the establishment of community relationships function along the energy axis. The law of energy transfer through the food chain and food web means that community species must use different methods of energy acquisition, which corresponds to dietary differentiation in the fish community. River fish can be divided into carnivorous, herbivorous, filter-feeding, and omnivorous species according to their feeding habits. Starch is the basis of energy in the food chain, and amylase is involved in the hydrolysis of starch and glycogen. Amylase is widely found in animals, plants, and microorganisms. Salivary amylase and pancreatic amylase are predominant in mammals (Takahiro *et al.*, 1986). As an important digestive enzyme, pancreatic amylase plays an important role in obtaining energy for fish.

5.2.1 Fish Amylase

Amylase (AMY/AMS) generally acts on enzymes that hydrolyze α -1,4-glycosidic bonds, such as soluble starch, amylose, and glycogen. Enzyme hydrolysates can be classified into alpha-amylase (EC3.2.1.1.) and β -amylase (EC3.2.1.2.) based on the isomerization type of the hydrolysates. Alpha-amylases are found in animals (*e.g.*, in saliva and the pancreas), plants (*e.g.*, malt and arugula), and microorganisms. Alpha-amylases break down internal A-1-4-glycosidic bonds (M.J. Darias *et al.*, 2006). From archaea to mammals, alpha-amylase plays a role in providing energy to living organisms (A. Pandey *et al.*, 2000; M. Machius *et al.*, 1995). Most organisms initially store energy as carbohydrates, including starch and glucose

polymers. More than 50% of the body's energy comes from the breakdown of carbohydrates, which begins with the enzyme amylase in saliva in the mouth and travels through the digestive system to the small intestine (W.F. Caspary, 1992). Amylase has various tissue-specific characteristics, including composition, copy number, and expression pattern. Salivary amylase is encoded by the *amy1* gene and pancreatic amylase is encoded by the genes *amy2a* and *amy2b*. The copy number of the salivary amylase gene varies greatly among species and even within the same population; *amy1* is more widely distributed than *amy2a* and *amy2b*. Changes in the *amy1* copy number are closely related to the levels of amylase in the saliva and serum. The changes in the *amy1* copy number in humans are related to the starch content of the diet. Unlike salivary amylase, pancreatic amylase is mainly derived from the pancreas and parotid gland. The mRNA expression levels of amylases in the AR42J cells of the pancreas are related to the activity of the amylases. The glucose/carbohydrate contents in the AR42J cells of the pancreas regulate the expression of amylases at the transcription level (C.D. Logsdon *et al.*, 1987). Glucocorticoid regulates the expression of pancreatic amylase and binds to the glucocorticoid receptor binding site on the amylase promoter, mediating hormonal responses (E.P. Slater *et al.*, 1993). The glucocorticoid response element (GR) in fish stimulates amylase gene expression using cortisol (glucocorticoid) (P. Ma *et al.*, 2004a; P. Ma *et al.*, 2004b). Transcription factor NF-Y plays an important role in glucose metabolism (Goel *et al.*, 2003) and can be positively regulated (Kawata *et al.*, 2003). ARNTL/BMAL1 is a transcriptional activator that controls various physiological processes by regulating the circadian rhythm of gene expression for about 24 h, thus affecting the metabolism and behavioral rhythms.

As an important digestive enzyme, amylase plays a critical role in the energy gain of fish. Amylase is found in all fish. The organs that secrete amylase differ among fish. In some fish, amylase is mainly secreted by the pancreas, while, in other fish, the intestine is an important organ of amylase secretion. Fish amylase is associated with feeding habits, and amylase activity levels are higher in herbivorous fish than in omnivorous fish (Douglas *et al.*, 2000; Akira *et al.*, 1987). Agrawal *et al.* (1975) compared amylase activity levels among carnivorous, omnivorous, and herbivorous fish and found that the amylase activity of herbivorous fish was greater than that of carnivorous fish. Hidalgo *et al.* (1999) showed that the amylase activity of *Anguilla japonica* was higher than that of *Oncorhynchus mykiss* and that the amylase activity levels of herbivorous and omnivorous fish were higher than those of carnivorous fish. It is necessary to characterize the mechanisms of fish community construction in the river ecosystem in order to understand the relationships between fish dietary differentiation and energy pathways.

The alpha-amylase gene in fish is similar to the *amy2a* gene in humans. The cDNA sequences of alpha-amylase genes have been published for many fish, such as *Siniperca kneri*, *Latescal cadfer*, *Brachydani orerio*, *Salmo salar*, *Tetraodon nigroviridis*, *Anguilla japonica*, *Myxocyprinus asiaticus*, *Pleuronectes americanus*, and *Epinephelus coioides*. The cDNA sequences of alpha-amylase genes are highly similar across fish species. For example, the saddle tail grouper amylase gene was 91.8% similar to the amylase genes of various other fish (Hu and Gallo, 2010, Chen *et al.*, 2009). The homology of the coding region of the amylase gene was 79.7% between

zebrafish and *Siniperca kneri*. The coding region of the amylase gene is highly conserved across fish taxa. Chen (2007) cloned the cDNA sequence of *Myxocyprinus asiaticus* alpha-amylase and studied the expression patterns of alpha-amylase in different tissues of *Myxocyprinus asiaticus*. Qin Bangyong *et al.* (2013) cloned the alpha-amylase gene of *Cynoglossus semilaevis* and studied the effect of feed additives on alpha-amylase gene expression.

5.2.2 Gene Sequence and Niche

The regulatory region at the 5'-terminal of the pancreatic alpha-amylase gene in various fish with different diets contains different gene sequences. The 5'-terminal region has many important regulatory factors, such as promoter and transcription factor binding sites, regulating the alpha-amylase gene expression. Alpha-amylase mRNA expression levels are correlated with enzyme activity, and alpha-amylase genes are transcriptionally regulated (Moal *et al.*, 2000). Multiple regulatory elements were found in the 5'-terminal regulatory region of the alpha-amylase gene of *Siniperca kneri* (Chen *et al.*, 2009). Ma *et al.* (2004a, 2004b) found that the transcription factor GR regulated amylase gene expression in *Latescal cadfer*. The tissue-specific regulation of the pancreatic amylase gene may be related to the presence of transcription factor 1 (PTF1) in the pancreas (S.L. Weinrich *et al.*, 1991; M. Cockell *et al.*, 1989). Gene transcription sites related to salinity and food quantity, including GATA-1, AP-1, and SP1, were identified in the structural gene of the amylase by Huang *et al.* (2016), suggesting a close relationship between amylase gene expression and food source.

The nucleotide sequences at the 5'-terminals differ, but functionally regulated sequences are more evolutionarily conserved (Ludwig M Z *et al.*, 2000). The evolution of gene transcriptional regulation is driven by changing gene expression, rather than by changing the amino acid sequence (German *et al.*, 2016; Shapiro M D *et al.*, 2004). Fish amylase genes were highly similar within species lineages from the start codon to 167 bp upstream; beyond 167 bp upstream, many fragments were missing.

We performed a phylogenetic analysis of the amylase genes from 32 fish belonging to 12 orders and 19 families. In the resulting tree, some species clustered into recognized classes and families such as the Perciformes and Cyprinidae. Omnivorous fish in the same lineage were not clustered into a specific category, and a variety of omnivorous fish was identified in a single lineage, suggesting that the fish-amylase gene promoter sequence was conserved at the order level (Zhu *et al.*, 2020). More closely related families and species did not appear in the same lineage, suggesting the existence of a community niche differentiation mechanism with the energy system as the axis. The functional system of the animal energy axis includes the amylase system, which is related to hydrolyzed carbohydrates (feeding); the decomposing system, which includes lipases and proteases (carnivorous); and the mixed hydrolyzed carbohydrate and lipoprotein enzyme system (omnivorous). There are three types of functional enzyme systems in animal bodies. However, the transcriptional regulation of the gene region that encodes the functional enzymes of the energy system varies according to community functional differentiation.

5.2.3 Amylase Gene Transcription and Animal Feeding Habits

Plants differentiate into high- and low-order community species in order to compete for solar energy. Animals can be divided into herbivores, omnivores, and carnivores. In order to compete for energy, different animals have different feeding habits. Although all three groups of animals have characteristic amylase genes, the expression levels of the amylase genes differ among animals. Feeding preference may influence the expression of the amylase gene, and differences in the regulatory sequence of the amylase gene (tf, transcription factor) may correlate with niche differentiation in animal communities. In addition to the single transcriptional mechanisms regulating pancreatic amylase gene expression, there are also multi-transcriptional co-mediated regulation mechanisms. Upregulation of the pancreatic amylase gene is associated with the binding of hepatocyte nuclear factor 3(HNF3) β or 3 γ and Ptf1 to the promoter, and these cytokines act synergistically to strongly express the pancreatic amylase gene (M. Cockell *et al.*, 1995). The co-regulation of the pancreatic amylase gene *via* multiple transcriptions increased gene specificity and flexibility for the control of development, differentiation, and growth (Z. Tan *et al.*, 2018; Z. Wang *et al.*, 2018; P.D. Radler *et al.*, 2017; D.T. Odom *et al.*, 2006; M.I. Arnone and Davidson, 1997). Transcriptional functional recognition methods (Z. Hu *et al.*, 2007, 2010) were used to analyze and evaluate the transcription factors that might be involved in regulating the pancreatic amylase 2A gene in herbivores, omnivores, and carnivores. It was found that GR, NFAT, and PR were the common hubs of the tf-tf interaction network across the different feeding groups. GR was the only tf regulatory network in herbivores and SPZ1 was the only tf regulating carnivore genes. Sequence analysis showed that tfs were highly similar within animal populations.

Amylase gene expression in omnivorous animals is also regulated by a variety of transcription factors (tfs). The interactions among transcription factors that regulate the pancreatic amylase gene in three different feeding habit groups were analyzed. The selected subjects included 77 herbivores, 25 omnivores, and 118 carnivores. Computer simulations of tf regulatory networks indicate that known pancreatic-specific tfs (such as GR, NFAT, and PR) may have non-pancreatic-specific tf mechanisms in the tf-tf interaction network. This may provide flexibility for the control of pancreatic amylase gene expression in different feeding habit groups. The results of this study suggested that combinatorial transcriptional regulation may be a key component in controlling the expression of the pancreatic amylase gene. The results suggested that the differential expression of energy-producing genes, including amylase, may be the key to the formation of herbivores, omnivores, and carnivores. Although amylase genes are present in all animal communities, their expression patterns in each community are controlled by different mechanisms, and tf regulation is probably related to food differentiation (Xinhui Li *et al.*, 2020). Wang *et al.* (2015) found that, during herbivore transformation, the expression pattern of circadian rhythm-related genes in the gut was reset, and that *Ctenopharyngodon idella* might obtain enough available nutrients to sustain its rapid growth through continuous high-intensity food intake.

Transcription factor Pax-2 performs the transcriptional activation of glucagon gene expression (Hoffmeister *et al.*, 2002). Glucocorticoid plays a major role in carbohydrate and protein metabolism. Any process which changes cellular state or activity (including movement, secretion, enzyme production, and gene expression) can be induced by glucocorticoid stimulation. MyoD is involved in the cellular response to glucocorticoids and regulates cellular metabolic processes.

We analyzed the upstream sequences of 32 pancreatic alpha-amylase genes from fish, including the promoters (Zhu Shuli *et al.*, 2020), to identify the major transcription factors that influence differences in fish feeding habits. Potential transcriptional factors that contributed more than 3.5% to differences in fish feeding habits were identified. It was found that E47, C/EBPalpha, NF-Y, and Pax-2 were the main transcription factors to identify fish as herbivorous or carnivorous, while deltaEF1, MyoD, NF-Y, AREB6, and Pax-2 were the main transcription factors to identify fish as herbivorous or omnivorous. The transcription factors that differ between omnivores and carnivores were GATA-1, SRY, MyoD, HFH-8, AREB6, Pax-2, STAT5A, and AP-1 (Li Xinhui *et al.*, 2020). This suggested that the differences in the transcription factors associated with the 5'-terminal region of the pancreatic alpha-amylase gene were related to dietary differentiation in fish. Transcription factors E47 and C/EBPalpha contribute 3.57% to the difference between herbivorous and carnivorous fish, and these are potential transcription factors that distinguish herbivorous and carnivorous fish. E47 plays a more important role in the expression of the alpha-amylase genes in herbivorous fish. PAX-2 also differed significantly between herbivorous-omnivorous and carnivorous fish; the difference contribution of NF-Y was 3.77% and that of PAX-2 was 4.67%. The MyoD and alpha-amylase genes differed significantly between omnivorous-herbivorous and carnivorous fish, and these genes were less likely to occur in carnivorous fish (Zhu *et al.*, 2020). When food resources are restricted, the differentiation among transcription factors led to niche differentiation in species communities. This mechanism of community construction focused on energy has theoretical significance for species management, conservation, and restoration, as well as the preservation of ecosystem communities.

5.3 Species Selection for Community Construction

River aquatic ecosystems are facing pressure due to global climate change, environmental changes caused by economic development, and the over-exploitation of water resources. These factors affect the sustainable development of the social economy. Fish biomass is an important factor for water system stability and water quality security. As a result of human activities and natural changes, river ecology has changed substantially. For example, the food chain system cannot fully utilize nutrients from the water body. In addition, deteriorated water quality cannot be used by people, which restricts the sustainable development of society. Against a background of the increasing pressure of environmental pollution, functional guarantees of the river ecosystem, which are demanded by human beings, have become a

target of social concern. Fish play the role of “scavenger” in the functional guarantee of water quality in the river ecosystem, and growing fish continuously transfer materials out of the water body and purify the water. The ecological management of rivers is developing toward the goal of guaranteed water quality, and the biological reconstruction of river fish is developing to meet the needs of energy circulation in the river ecosystem. It is important to control fish communities and biomass according to the needs of the river water-quality guarantee. The nutrients in the river enter the food chain through the absorption and growth of autotrophs, and these nutrients are transported from primary productivity to secondary productivity to the fish output. If fish are lacking, the transport chain will become blocked, surplus organisms will decay, and the quality of the ecosystem will deteriorate. This will become a vicious cycle.

The environmental characteristics of the river basin are formed during natural evolution and the biomass of the river ecosystem is subject to the number of mineral nutrients entering the system. At present, many aspects of the fish living environment are under stress, especially with respect to insufficient biomass. The food chain is responsible for material transport in the aquatic ecosystem. In addition to community types, biomass is also required to meet the needs of the ecosystem. Observation of the nutrients in river water, the productivity of the water ecosystem, fish food availability, and product availability will help evaluate the functional quality of river ecosystems, help evaluate the management objectives, and help establish a river ecosystem management scheme with fish biomass as the target. The increase of fish resources in the water space can greatly enhance the nutrient output of the aquatic ecosystem and reduce the pressure of nitrogen and other nutrients on water quality in the river ecosystem. Wang *et al.* (2020) suggested that the reciprocal relationships between species have a significant top-down influence on predators and consumers and that these reciprocal relationships have a greater influence on predators than on consumers. Low trophic-level biomass has a significant bottom-up effect on consumers and predators, and this effect decreases with the increase in nutrient level. High-trophic-level organisms are influenced from bottom to top by low-trophic-level biodiversity, and carnivores are mainly influenced by mutually beneficial organisms from top to bottom. Food webs with reciprocal relationships are highly complex, and the interactions between trophic levels are significant. A food web can be functionally assessed by measuring the “total input–output interaction intensity” (Xu *et al.*, 2020).

The functional connections between biological communities and ecosystems have received substantial research attention. However, it is now common practice to equate biodiversity with species diversity (Díaz and Cabido, 2001), ignoring the role of interspecies relationships in ecosystem processes. Ecosystem function depends not only on the number of species but also on the functional traits of the species (Lepš *et al.*, 2001). Two communities with the same number of species are likely to exhibit significant differences in functional diversity due to the differences in species traits and characteristics between the communities (Leps *et al.*, 2006). Therefore, it has been increasingly commonly proposed that community studies should consider the diversity of functional traits instead of species diversity (Jiang and Zhang, 2010). The functional diversity represented by the traits is closely related to the function of the

ecosystem, which more clearly reflects the interactions among species in the community (Hooper *et al.*, 2002; Díaz and Cabido 2001). That is, functional diversity is defined as the overall differences in or diversity of functional characteristics among species within a community (Petchey and Gaston, 2006). In comparison to ecosystems with low functional diversity, ecosystems with high functional diversity tend to have more differentiated niches, to maximally use resources, to be more stable, to have higher productivity (Tilman *et al.*, 1997), to be more resilient (Nyström and Folke, 2001), and to have a higher resistance to invasion (Dukes, 2001; Prieur-Richard and Lavorel, 2000). Community functional ecology has become an important present-day approach to the treatment of ecological problems (Cameron, 2002; Loreau *et al.*, 2001).

The spatial differences among fish communities are closely related to the environment (Shuai *et al.*, 2017b, 2020; Zhang *et al.*, 2020). In each specific environment, fish communities show common functional characteristics, such as the long narrow bodies favorable for fast swimming in fish adapted to rapids' habitats. Thus, fish develop special functional structures to adapt to the niche (Shuai *et al.*, 2016, 2017b, 2018a, 2018b). In a study of the niches and functions of fish in ecosystems, Mason *et al.* (2008) proposed three independent functional diversity indices: functional richness (FRic), functional evenness (FEve), and functional divergence (FDvar). The niche space occupied by a species, the distribution of species characters in the space occupied, and the degree of niche complementarity among species in a community were characterized. The functional divergence (FDvar) index was proposed to quantify the distribution and multiplicity of species in the functional space (Sébastien Villéger *et al.*, 2008). Fish community ecological process should be based on indices of functional specialization (FSpe), functional dispersion (FDis), and functional entropy (FEnt). These are the factors to be considered in the construction of fish groups. Hoenighaus *et al.* (2007) collated species abundance data from 157 streams in Texas, USA, and classified fish into functional groups based on trophic level and life history features. Taxonomic analysis showed that the fish communities were determined by the size of the river and the classification of the biogeography patterns. Functional group analysis showed that the characteristics of the fish community were not related to river scale and geographical region but were instead related to habitat type. It was concluded that the structures of the fish assemblages in local rivers were ultimately determined by factors at multiple scales, and the relative importance of each scale depended on the biological unit (species or functional group) used (Hoenighaus *et al.*, 2007). The river ecosystem is linear, with a large environmental span, and high heterogeneity. Thus, differences among fish communities are correspondingly great, and fish community construction studies need to consider the indigenous biology priority program. Fish community construction must be guided by community niche theory. Although the same species of fish occur in different communities, the occupied niche may differ. Thus, it is necessary to consider various river environments when building different fish communities.

Some species have a wider geographical range in nature than others, and such widespread species are well adapted to the environment, independent of their size (Lester *et al.*, 2007). These species are of concern for the formation of diverse

communities. Biological interactions shape the spatial distributions of species at local spatial scales, but the role of these interactions beyond the local scale (10 km to global) is generally considered unimportant. Contemporary and palaeoecological analyses show that biological interactions shape species distributions, including individual species ranges, functional groups, and species richness patterns, beyond local scales. This suggests that biological interactions leave clear marks on species distributions and influence species assemblages across all spatial scales (Wiszniewski *et al.*, 2013). One long-standing concept of community research is that closely related species compete more fiercely than do distant relatives. Ecologists invoke this “restricted similarity hypothesis” to explain patterns in community structure and function and to provide information for conservation, restoration, and the management of invasive species. However, Fritschie *et al.* (2014) found evidence that interspecies interactions are independent of phylogenetic distance. Thus, interactions between any pair of species (including alien species) may be random. The potential free association of species may be supported by the food differentiation theories discussed above.

River fish communities have unique patterns and are affected by non-random processes (Ostrand and Wilde, 2002; Jackson *et al.*, 2001). Environmental factors (*e.g.*, water flow velocity, dissolved oxygen concentration, water temperature, and dissolved organic matter) affect the fish composition and distribution (Sharma *et al.*, 2011; Mason *et al.*, 2007; Mouillot *et al.*, 2007; Peres-Neto *et al.*, 2006; Grenouillet *et al.*, 2004) and affect strategies for fish resource use (Poff NL and Allan, 1995). Ecosystems have changed dramatically since humans have come to dominate the environment, and conservation action is urgently needed (Banse, 2007). During the process of ecological protection, the reconstruction of biological communities must be guided by the theory of continuous development. Importantly, the key to stable community reconstruction is a detailed understanding of the relationships between organisms and habitats and of the interactions among species. The processes of ecosystem evolution and community succession should be considered during ecosystem reconstruction. The mechanisms of energy output by primitive communities and the evolution of species from low trophic levels to higher trophic levels may act as a reference. That is, the efficiency of energy output should be considered first in the process of reconstruction, and low trophic-level organisms should be given priority in community construction.

The reconstruction of a biological community should be performed with the goal of ecosystem function. The reconstruction of biological communities in the river ecosystem can be divided into two aspects according to functional needs. The first aspect is biodiversity reconstruction, which includes the maintenance of harmonious relationships among target species and community reconstruction (aquatic) species; the problem of regional biodiversity collapse must be solved through community reconstruction. The second aspect is ecosystem restoration, which must serve human needs and consider special functions, such as the functional impairment of energy delivery systems due to the loss of species or biomass in river ecosystems disturbed by human activities. Ecosystem communities that need rebuilding are usually located in eutrophic waters and waters with high primary productivity, and

functional communities need to be constructed using fish communities that can utilize primary productivity and the available biomass.

Niches are linked in a community by the food chain, and species niches are often manipulated by superior predators from above (Grange and Duncan, 2006). Urban (2004) postulated that community species diversity, abundance, and nutrient structure were determined at the time of ecosystem formation and by local environmental changes. Heterogeneous perturbations determine community structural properties, barriers restrict many taxa to localization, species cannot adapt to the disturbance, and a normal biological interaction relationship could not be established. Blocking may also affect local dynamics through mass effects. Many species have “abandoned their adaptation to dam disturbance” and “abandoned their adaptation to biological interactions” due to the formation of “artificial freshwater lakes” in rivers due to cascade dams. Changes in the fish community lead to deficiencies in the food chain system, which is reflected in changes in the composition of fish trophic levels. The relationship between phylogenetic distance and ecological similarity is the key to an understanding of community mechanisms. Phylogenetic information can be used to infer the mechanisms of community construction, niche conservation (habitat filtration), and species interaction (competition or promotion). These factors affect community composition, and phylogenetic relationships affect species interaction intensity (Jean H. Burns and Strauss, 2011). Alien species can alter resource availability and trophic level structure at the ecosystem level, thus affecting ecosystem function and even ecosystem stability (Cucherousset *et al.*, 2012; Britton *et al.*, 2010).

Space, time, and trophic niche are three typical dimensions considered in the construction of a species niche, as these variables independently describe the ecological location and animal resource use. When multiple species coexist in the same community, they inevitably interact within various dimensions of the niche (Sæbø, 2016). Spatial, temporal, and trophic niches provide three variable coordinate dimensions for predators, and animals can adjust their ecological breadth and range in each dimension through adaptation or behavior changes in interspecies competition to maximize returns (Bruno *et al.*, 2003; Schoener, 1974). Spatial, temporal, and trophic niches also reduce the intensity of competition among species, including interference competition (direct encounter and killing) and exploitation competition (Kronfeld-Schor and Dayan, 2003). The plasticity and adaptation of predators in space, time, and trophic niches is an evolutionary consequence that retards competition and promotes the coexistence of different species (Sæbø, 2016). The spatial niche is fundamental for an understanding of the regional coexistence and interactions of carnivores. Species can potentially interact in niche dimensions such as time and nutrition only if they co-exist within certain spatial scales (Farris *et al.*, 2020). Differences in spatial use can effectively promote species coexistence, but species with similar ecological characteristics do not enforce complete spatial competitive exclusion on a global scale; instead, species chose spaces with similar resources to promote coexistence (Davis *et al.*, 2018).

The trophic niche, which is an important attribute of the wildlife niche, can affect the functions of animals in the ecosystem and the food web. Comparison of the food habits of sympatric species can reveal the extent of trophic niche overlap among

different species and serve as a measure of potential competition among species (de Satgé *et al.*, 2017). The trophic niche overlap among carnivores tended to decrease in a non-linear manner as body weight difference increased; at the minimum body weight difference (about 0.01 kg), the predicted trophic niche overlap was 62%. The predicted trophic niche overlap was 12% at the maximum weight difference (160 kg) (Lanszki *et al.*, 2019). Large and small carnivores distributed in the same domain generally do not have a high degree of food overlap (Gómez-Ortiz *et al.*, 2015), and similar-sized predators are more likely to engage in intense interspecies competition and even slaughter one another as they hunt similar prey (Donadio and Buskirk, 2006). Coordinated and complementary spatial, temporal, and trophic niches promote the regional coexistence of species, and it is important to consider differences in habitat preference among species.

Li *et al.* (2021) devised an indexing system to evaluate the functional state of the river ecosystem and to provide a reference for the biomass demand of the river fish community. The eutrophication of the river water ecosystem must be reversed by eliminating nutrient input and establishing a biological output system. It has become commonly understood in society that it is necessary to reduce the point and non-point flow of nutrients into water bodies, but methods to establish biological community systems and to enhance the export of water substances still remain to be developed. By determining the biomass demand of fish, the fish community morphological model can be used to analyze the relationships between fish and the community and may provide an analytical method for niche construction for community species.

5.3.1 *Multispecies Model*

It is difficult to measure the impact of biodiversity on ecosystem function because it is difficult to design and analyze the impact of a species loss on ecosystems in nature. Changes in the consumption patterns of large vertebrates, such as humans, are highly disruptive to ecosystems, and the main effects of biodiversity changes on ecosystem functioning appear to be in the form of nutrient-mediated processes (Duffy, 2003), particularly energy processes. In ecosystems disturbed by anthropogenic activities, the special ecological functions of human beings should be considered during the reconstruction of the biological community. The effects of humans on the energy transmission functions of the ecosystem are greater than those of high-level carnivores. Usually, the energy output (removal) of the system depends on the realization of high-level carnivore function, and the reduction in carnivore numbers will thus affect the material transfer functions of the system. Because human beings are unusual animals in the biological community due to their broad-spectrum diet, wide feeding abilities, and creativity, humans do not replace the carnivores in the original system. Instead, humans act as the terminal energy output of various natural ecosystems, changing the structure and function of the ecosystem. Therefore, with human effort, the ecosystem's energy output can finally become balanced.

During adaptation, large and small organisms form a pattern of coexistence, and the relationships among species niches in the community are not exclusive. These mechanisms should be related to the maximum utilization of energy in the ecosystem. Aarssen *et al.* (2006) indicated that the ecological niche of large, tall trees was dominant. However, the gaps among the trees provided space for the survival of small species and promoted the coexistence of large and small species. In the animal system, energy circulation requires the predator-prey relationship in the food chain. If the predator has the exclusive behavior in the niche, there is no basis for its existence. Therefore, the “linking” of the niche is the rational distribution of energy in the system, which provides the basic condition for the coexistence of large and small species. It is not only when competition is weak that positive intraspecific interactions, such as cooperative defense against predation or enhanced foraging success, occur. Thus, interactions can be positive only if the benefits outweigh the costs, but competition is not necessarily weak and interactions can have a net positive outcome. Strong benefits, such as improved survival, can outpace the negative competitive effects of growth, and net positive interactions are costly for co-existing species. The overall biota may also be more resilient to physiological stress than isolated individuals. In stressful environments, bioclusters can cushion the environment. On coasts with higher plant and animal densities, shade from some species compensates for extreme variability of air temperatures, reducing evaporation.

The evolutionary relationships and phylogenetic positions of coexisting species can be studied in a community. In a balanced system, biological heterogeneity (*e.g.*, due to alien species) can lead to imbalance. The coexistence of species with spatial differences in resource supply must balance at a regional scale along the environmental gradient. Environmental heterogeneity may lead to the exclusion of the dominant competitors of a given species within certain ranges (Shurin *et al.*, 2004). There is no single explanation for the patterns of species richness found in coexistence studies of plant species. A more general interpretation of species coexistence makes use of the idealized concept of an empty community, defined as an undisturbed community within a stable band of vegetation that includes an entire pool of potential species. The number of potential species will then depend largely on evolutionary factors (*e.g.*, speciation). For historical reasons, most “real” communities have small species pools, and migration has not yet taken place. In this local pool of species, asymmetrical interspecific competition is a major force in reducing species richness. The competition takes place at the ecological level (within ecological time scales), but the outcome is governed by the characteristics determined by individual evolution. Ecological factors affect the number of coexisting species in two cases: First, when habitat conditions change and one species pool replaces another; second when certain external factors (such as grazing or mowing) that change the character of an individual also change the outcome of the competition (Zobel, 1992). Methods of systematic species construction, reconstruction, community niche modeling, and development need to be devised (Webb *et al.*, 2002).

The structure of the food web is related to species diversity, and the loss and extinction of key species greatly affect the structure of the food web (Dunne J A *et al.*, 2002). Interspecific and intraspecific interactions have a significant impact on

the structure and function of ecosystems, as does the avoidance of physical stress. The idea that changes in the habitat of one species might benefit another species was submitted by early plant ecologists decades ago as an important mechanism for community succession. Understanding how biological communities form has been a central issue in ecological research. The traditional concept, with set rules, reflects the idea that species do not co-occur at random, but are restricted by interspecies competition, and that the co-occurrence of species is the product of opportunity. Thus, historical patterns of speciation and migration, diffusion, abiotic environmental factors, and biological interactions are not mutually exclusive (Gtzenberger *et al.*, 2012).

Trade-offs between mutually beneficial actors (Wang *et al.*, 2008), namely the prevention of competition (conflict) between the recipient and the cooperating actor in a cooperative system, remain one of the biggest problems in evolutionary biology. Self-restraint, fragmentation, or spatial restraint can prevent direct competition for local resources or any other common resource, thus maintaining stable cooperative interactions, but these restrictions do not sufficiently prevent the symbiote from using more local resources at the expense of the recipient. The conflict may disrupt cooperative interactions, which occur when local resources are saturated by symbiotes. Increased repression of symbionts, which in turn inhibits the use of local resources during the conflict, is essential for sustaining and developing cooperation.

There are significant differences among types of predator-prey interactions; these interactions may be stable, competitive, and fluctuating. The probability of the stability of the predator-prey network is reduced by the presence of a large number of weak interactions in the food network structure. As long as the predator-prey pair is tightly coupled, a stable predator-prey network can be arbitrarily complex (Allesina and Tang, 2012).

Food chain systems are associated with species coexistence, and increasing ecosystem size can also promote coexistence, increasing the length of the food chain by changing the predator or increasing the number of prey (Takimoto *et al.*, 2012). There are many alternatives to consider when rebuilding biological communities in a given environment, but maintaining multiple species at multiple trophic levels is challenging, and in measuring the stability of multispecies coexistence, differences in the mean fitness and stability of species are key (Chesson, 2018). Community research needs to focus on mutualism and coevolution, as well as on quantifying species richness in communities. An improved understanding of these forces will improve our understanding of mutualism (coevolution), including the identification of coevolutionary units and coevolutionary interactions (Hall *et al.*, 2020).

5.3.2 The Food Chain is Fully Functional

The community of the aquatic ecosystem is influenced by environmental factors, such as global warming. Due to global warming, the tropical community has expanded toward the subtropical and temperate zone; such an expansion has been observed globally in tilapia. Human introduction hastened the expansion of tilapia. Human-induced cascade development accelerated the loss of habitat for indigenous

low-trophic fish, giving tilapia the opportunity to expand and occupy the vacant niche. In the future, it remains possible to further develop the river ecosystem from the current environment. The main body of the fish community in the aquatic ecosystem must be low trophic-level or omnivorous fish, as these provide the environmental conditions necessary for low trophic-level fish like tilapia to enter the food chain system of the river ecosystem in China. As an invasive species, tilapia (*Oreochromis niloticus*) has attracted much international attention in studies of community structure and ecosystem function (Shuai *et al.*, 2018a, 2018b, 2019; Yuguo Xia *et al.*, 2019; Tan *et al.*, 2012), as well as in analyses of biological invasion using geo-material information for niche modeling, of invasion pressure using niche conservation models (showing stagnation and maintaining niche parameters over time), and of niche evolution (adapting to changes in niche parameters) (Dudei and Stigall, 2010). To model biological modification, environmental regulation, and species distribution (Linder *et al.*, 2012), the biological modification environment can be expressed as additional variables in the SDMS. Exotic species have altered the body sizes of freshwater fish worldwide (Blanchet *et al.*, 2010) to varying degrees, thereby affecting the functional diversity of freshwater fish (Matsuzaki *et al.*, 2013). The impacts of alien species can be analyzed and predicted by establishing models based on population expansion data (Shuai *et al.*, 2015). However, it is difficult to predict the distribution trends of invasive species by using species distribution models when data are limited. At present, the differences in a niche location, size, and structure between local and large scales are quantified using multivariate similarity data and climate prediction factors (annual minimum and maximum temperatures). The niche change of the target species was realized by repeating the niche study using simulated species (Larson and Olden, 2012). Tilapia, which grows in natural waters, is not easily utilized by humans and is, at present, the lowest trophic-level fish that is gradually expanding in the southern Chinese rivers. From the point of view of ecosystem function, tilapia replenish the niche space of low trophic-level fish, and this species has a strong ability to adapt to the environment.

We analyzed the community structure of native fish. Data from resource supplements and the literature indicated that the entire Chinese river ecosystem was short of fish, especially low trophic-level fish such as *Ctenopharyngodon idella* and *Hypophthalmichthys molitrix*. We suggest that tilapia may be environmentally suitable for entrance into the food chain system in Chinese rivers, as tilapia, which are low trophic-level fish, fill the ecological gap left by native fish. It is necessary to comprehensively study the impacts of exotic species on the functions of the river ecosystem in China. Niche models can be used to study the mechanisms of invasion and niche expansion of a species in a community (Ebeling *et al.*, 2008). The analysis of fossil data can reveal the processes and forms of niche evolution (Malizia and Stigall, 2011). In general, the functional differences in the resource requirements of alien aquatic organisms have a significant impact on aquatic ecosystem processes (Azzurro *et al.*, 2014; Zhao *et al.*, 2014).

5.3.3 Species Compatibility

Community construction must solve the problems of species coexistence and species diversity maintenance, but the mechanisms of local community construction remain controversial. Maron *et al.* (2004) argued that phenotypic plasticity is the main mechanism by which alien plants adapt to rapidly evolving environments.

Phenotypic change processes reflect the niche change process of the species. Species with different traits (especially those that determine their impact on the environment) coexist differently depending on evolutionary history or the interactions among resources, predation, and prey. In near-equilibrium local communities, species coexistence is compatible, not random, with strong interaction mechanisms (Leibold, 1998). Chai and Yue (2016) holds that it is possible for theoretical ecologists to study the mechanisms of community construction based on functional traits and community genealogical structure; in addition, spatiotemporal scale, environmental factors and vegetation type, species and numbers of selective traits, intraspecific trait variation, and human disturbance should also be considered. Meng *et al.* (2020) studied the construction of microbial communities and concluded that microbial survival, growth, and reproductive traits can reflect microbial responses to environmental changes. Subsequently, microbial species distribution patterns, community construction mechanisms, and corresponding ecosystem functions are affected. In community construction research, it is also important to study the relationship between characters and species distribution patterns, biodiversity, ecosystem function, and response to environmental change. During community restoration and reconstruction, species, functional, and phylogenetic diversity can recover rapidly given small- and large-scale surrounding natural vegetation (Sun *et al.*, 2018). This edge effect requires that the physical environment of the system not be fragmented.

When transforming nature, it is important to consider the law of material metabolism. That is, we should only exploit biological resources at a reasonable level in the process of production, and we should not exhaust fishing resources. Biological evolution is a product of the interactions between living things and the environment. Living things continuously import and export materials from the environment, and the material environment that is changed by living things in turn influences or selects living things. This process is commonly known as normal natural succession. The environmental characteristics of the watershed are formed over the course of natural evolution, and the biomass of the system is related to the mineral nutrition of the water body. Similarly, the biological capacity of a river ecosystem is limited by the number of nutrients entering the system. Currently, many aspects of the fish habitat are under stress, especially with respect to insufficient biomass, although the food chain performs material transport in the aquatic ecosystem. River ecosystems need more reasonable community diversity, and biomass is also required to meet the needs of the ecosystem. The increase in fish resources in the water space can greatly enhance the nutrient output of the aquatic ecosystem and reduce the pressure of nitrogen and other nutrients on water quality in the river ecosystem (Li *et al.*, 2021a, 2021b).

In the study of species relationships within a community, the species relationship model can be used as a dynamic ecological model with which to study the

coexistence and competitive exclusion of species in the community. Community construction is mainly driven by deterministic and stochastic processes, and one of the key issues in community construction is the quantification of the relative contributions of deterministic and stochastic processes to community construction (Luo *et al.*, 2021). Deterministic processes are controlled by the inherent properties of the species in the community, and the stochastic processes reflect the results produced by the environment. Xu Y. *et al.* (2019) suggested that the decomposition and quantification of environmental impact factors should be considered in community construction, in addition to the impact of spatiotemporal dynamic changes. An improved understanding of the shared and unique traits of construction mechanisms among communities will emphasize integration with other ecological processes and the mechanisms of community construction. The regional coexistence of terrestrial animals is a relatively stable state that has been formed over evolutionary time; animals coexist through niche separation, and co-adaptation plays an important role in their regional coexistence (Li Zhilin *et al.*, 2021). In community construction, the degree of contribution of each species is usually determined by niche relationship or population number. Yang *et al.* (2020) analyzed the construction mechanisms of fish communities at different spatial scales in the middle reaches of the Yangtze River and found that the competitive effects of small-scale environmental filtration become those of large-scale environmental filtration. Lou *et al.* (2020) used phylogenetic community structure analysis to study the characteristics of fish communities in rivers filtered by the environment. These studies indicated that the important roles of competition and coexistence in maintaining the stability of the fish community should be fully considered in the process of community reconstruction. Zhou *et al.* (2021) used feeding habits and trophic levels to rank the 30 most common fish species based on the dominance of resources, biomass, and catch rate in the Wanshan area of the Pearl River estuary. Based on the trophic level of the food chain, seven key functional groups of fishery resources in marine ranches, which were composed of the top economic species, were identified, and the proportion of each trophic level in the food chain was determined. The functional groups included the Silver Pomfret Functional Group, the Sea Bass Functional Group, the Spiny Pomfret Functional Group, the Grey Mullet Functional Group, the *Penaeus monodon* Functional Group, and the Green Crab Functional Group. In aquatic ecosystems, microorganisms and plants convert inorganic and chemical energy into biological energy and enter the food chain. As the end group of energy output in the food chain of the aquatic ecosystem, fish species are rich in diversity and have different feeding habits, including filter-feeding on phytoplankton and zooplankton, omnivory, saprophagy, and carnivory. These groups play different roles in the chemical and energy cycles within the system. Lin *et al.* (2020) postulated that predation is the most important interspecific interaction in the Pearl River Delta fish community because predation is the most direct means by which the whole community and ecosystem can achieve energy flow and express ecological functions. Predation is also a direct factor that fish community construction efforts need to consider. Fish are the most important carriers of material circulation and energy flow in river ecosystems. The structure and stability of the fish community determine the

function of the river ecosystem. Interspecific relationships are particularly important in fish community restructuring.

In the river ecosystem, fish depend on the environment and have different reproduction requirements. Therefore, it is necessary to consider whether there is a suitable condition for fish reproduction and to build fish communities that enable reproduction. The distribution patterns of fish species differ in different areas due to the differences in climate and environment between the north and south. Fish species require different water temperatures for breeding. For example, the breeding water temperature of *Lota lota* (Linnaeus, 1758) in Heilongjiang is about 0 °C; the breeding water temperature of *Cyprinus carpio* and *Carassius auratus* is about 15 °C; the breeding water temperature of *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, and *Hypophthalmichthys nobilis* is above 18 °C; and the optimum water temperature for tilapia reproduction is 25–28 °C. The spawning behaviors of broodstock are mainly determined by water temperature, and spawning behaviors are inhibited if the water temperature is too high or too low (Wang and Li, 2010; Mu Yunlei, 1999; Yu Zhitang *et al.*, 1985). The characteristics of the fish community are controlled by temperature, and the suitability of the species for the temperature environment should be considered in the construction of fish communities. Some fish require suitable conditions for the development of floating eggs. After the eggs are laid, they swell to form a large periovale, which is a little lighter than water. The fertilized eggs are suspended in the water column in running water and sink to the bottom in still water. Fish that produce floating eggs include *Ctenopharyngodon idella*, *Mylopharyngodon piceus*, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, *Squaliobarbus curriculus*, *Sinibotia pulchra*, *Luciobrama macrocephalus*, *Ochetobius elongatus*, and *Elopichthys bambusa*. Some fish spawn and sink to the bottom or adhere to the pebbles, gravel, or developing reefs. The viscous spawning fish, such as *Acipenser sinensis* (Gray, 1834), *Megalobrama terminalis*, *Zacco platypus*, *Oreonectes furcocaudalis*, *Spinibarbus denticulatus denticulatus*, *Spinibarbus denticulatus denticulatus*, *Onychostoma sima*, *Onychostoma meridionale*, *Onychostoma meridionale*, *Onychostoma gerlachi*, *Discogobio tetrabarbatus*, *Glyptothorax fukiensis*, and species in the Tetraodontidae, have eggs that sink to the bottom or adhere to pebbles, gravel, or reefs. Some eggs are lighter than water and float. There are a few species of river fish that produce floating eggs, including *Coilia* spp., *Monopterus albus*, *Channa argus*, *Siniperca kneri*, *Maropodus* spp., and *Tenualosa reevesii*. The eggs are generally small and contain oil globules. Fish eggs are typically colorless, transparent, and free-floating in the upper water layers. The presence, color, number, size, and distribution of the oil globules in the egg are important classification characteristics for fish eggs. Some fish eggs have only a single oil globule, such as those of *Clupanodon punctatus*. Some fish eggs, such as those of *Tenualosa reevesii*, contain several different sizes of oil globules, and they are known as multi-oil-globule eggs. During embryonic development, the oil globules of single-oil-globule eggs are located at the plant pole of the ovum, while the oil globules of multiple-oil-globule eggs are scattered around the yolk. The oil globules collect before and after hatching, forming an oil lump at one end of the yolk sac that is eventually absorbed or removed. Some fish produce eggs after adhering to the stems and leaves of aquatic plants (these fish include *Cyprinus carpio* and *Carassius*

auratus). Other fish, such as *Acheilognathus macropterus*, *Acheilognathus barbatulus* (Günther, 1873), and *Acheilognathus tonkinensis* (Vaillant, 1892), lay their eggs in mussels. During the construction of fish communities, fish breeding habits should be considered to ensure that species can reproduce in the system environment.

Habitat restoration projects, especially the estimation of the habitat requirements of a spawning ground, should consider the niche of the fish community. Li xinhui *et al.* (2020a, 2020b, 2020c, 2020d, 2020e, 2021a, 2021b) established a theoretical system of ovipositing field function, which involves several functional concepts including unit, hydrodynamics, and flux.

5.4 Examples of Model Application

The river environment changes dramatically under the influence of non-natural factors. Thus, species whose life history processes depend on a certain habitat and cannot adapt to environmental change will go extinct. The surviving species are fixed in the community relationship formed by ecosystem evolution and cannot adapt to the niche vacancy caused by species loss in a short time. The food chain breaks, the energy cycle is disrupted, and the ecosystem falls into disarray. The function of the river ecosystem deviates from human needs, and the river ecosystem needs to be rebuilt. For this effort, species compatibility, including the compatibility among species within the community and between species and environmental factors, is important. An understanding of the biological characteristics of the species, combined with a model to predict the structure of the proposed community, will help to clarify the need for ecosystem reconstruction.

5.4.1 Relationship Between Model “Potential” and Community Abundance

Data from field observations are affected and restricted by many factors. Particularly due to data mutations and discontinuity, it is not easy to obtain satisfactory quantitative results. Analysis of the species richness of the community formed by the early fish resources in the Pearl River showed that seasonal spawning times, spawning rate, and spawning frequency differed among the 19 species of fish. However, for certain species, it is only possible to use traditional methods of studying fish biomass; due to larval size gaps and data discontinuity at different times (some fish may be present as larvae for only 10–20 days per year), a valid quantitative model cannot be obtained. During the establishment of the niche model of community species based on morphological parameters, the spatial relationships among community species were determined within the framework of species morphological parameters. Then, the niche “site-abundance” relationship was calculated based on the measured biomass indices (*i.e.*, the abundance of supplementary resources). Specifically, the number of species (species richness) was added to the morphological parameter matrix of the species, and the corresponding analysis was performed in the model to obtain the two-dimensional graph of community

relationships. In this graph, the ecological niches of all fish were shown along the two principal axes (*i.e.*, the x - and y -coordinates).

Because there is not a simple linear relationship between the biomass abundance of a given species and its morphology-derived community niche, screening and comparison must be performed to establish a regression model using powers of 1–2 or 1–3. Thus, the relationship between the abundance of each fish and its biomass can be calculated as follows:

Ci (The abundance of a certain fish in a community) = F(X_{A1} (x coordinates of the first fish),... X_{A19} (x coordinates of the 19th fish), Y_{A1} (Y coordinates of the first fish),... Y_{A19} (y coordinates of the 19th fish). The x , x^2 , x^3 and y , y^2 , y^3 in the functional equation are the different powers of the x and y coordinate values of fish, respectively. For example, in the equation of *Mylopharyngodon piceus*, CA1 indicates the abundance value (%) in the community, CA1 function:

$$C_{A1} = 12.74 + 247.15 \times X_{A1} - 2150.9 \times X_{A1}^2 - 27023.41 \times X_{A1}^3 + 401.68 \times X_{A2} + 740.17 \times X_{A2}^2 - 21797.43 \times X_{A2}^3 - 41.25 \times X_{A3} - 6629.48 \times X_{A3}^2 - 42345.35 \times X_{A3}^3 + 740.52 \times X_{A4} + 2389.03 \times X_{A4}^2 - 7406.19 \times X_{A4}^3 + 245.65 \times X_{A5} + 96.31 \times X_{A5}^2 + 45.63 \times X_{A5}^3 + 197.55 \times X_{A6} - 3905.47 \times X_{A6}^2 - 69146.59 \times X_{A6}^3 + 247.17 \times X_{A7} + 44.18 \times X_{A7}^2 + 228.1 \times X_{A7}^3 + 249.05 \times X_{A8} - 10.79 \times X_{A8}^2 + 92.41 \times X_{A8}^3 + 202.98 \times X_{A9} + 389.08 \times X_{A9}^2 - 2285.08 \times X_{A9}^3 + 226.82 \times X_{A10} - 2615.18 \times X_{A10}^2 + 76161.11 \times X_{A10}^3 + 4342.93 \times X_{A11} + 97153.81 \times X_{A11}^2 + 751580.81 \times X_{A11}^3 - 1302.99 \times X_{A12} - 56317.97 \times X_{A12}^2 - 769106.03 \times X_{A12}^3 - 3810.92 \times X_{A13} - 89458.46 \times X_{A13}^2 - 640143.54 \times X_{A13}^3 + 313.24 \times X_{A14} - 290.36 \times X_{A14}^2 - 5112.97 \times X_{A14}^3 + 245.59 \times X_{A15} + 218.4 \times X_{A15}^2 - 375.99 \times X_{A15}^3 + 216.01 \times X_{A16} - 578.24 \times X_{A16}^2 + 3150.88 \times X_{A16}^3 + 257.13 \times X_{A17} + 350.13 \times X_{A17}^2 - 2579.95 \times X_{A17}^3 + 130.23 \times X_{A18} - 460.38 \times X_{A18}^2 + 928.24 \times X_{A18}^3 + 184.09 \times X_{A19} + 71.55 \times X_{A19}^2 + 50.98 \times X_{A19}^3 - 65.08 \times Y_{A1} - 7691.46 \times Y_{A1}^2 + 46708.18 \times Y_{A1}^3 + 180.01 \times Y_{A2} + 5441.13 \times Y_{A2}^2 + 103436.12 \times Y_{A2}^3 + 514.6 \times Y_{A3} + 2628.08 \times Y_{A3}^2 - 3297.11 \times Y_{A3}^3 + 214.22 \times Y_{A4} - 1990.7 \times Y_{A4}^2 + 37700.16 \times Y_{A4}^3 + 346.43 \times Y_{A5} - 172.25 \times Y_{A5}^2 + 485.56 \times Y_{A6} - 284.79 \times Y_{A6}^2 - 32269.7 \times Y_{A6}^3 + 350.65 \times Y_{A7} - 121.46 \times Y_{A7}^2 + 200.29 \times Y_{A7}^3 + 312.95 \times Y_{A8} - 122.38 \times Y_{A8}^2 + 282.14 \times Y_{A9} + 124.56 \times Y_{A9}^2 + 213.07 \times Y_{A10} + 2051.29 \times Y_{A10}^2 + 478.6 \times Y_{A11} + 994.49 \times Y_{A11}^2 + 302.75 \times Y_{A12} - 1675.17 \times Y_{A12}^2 + 524.59 \times Y_{A13} + 3987.59 \times Y_{A13}^2 + 493.71 \times Y_{A14} + 127.88 \times Y_{A14}^2 + 327.07 \times Y_{A15} - 335.11 \times Y_{A15}^2 + 250.3 \times Y_{A16} + 98.87 \times Y_{A16}^2 + 250.95 \times Y_{A17} + 2653.54 \times Y_{A17}^2 + 255.89 \times Y_{A18} - 517.99 \times Y_{A18}^2 + 254.84 \times Y_{A19} - 216.44 \times Y_{A19}^2$$

Comment: A1 (*Mylopharyngodon piceus*), A2 (*Ctenopharyngodon idella*), A3 (*Hypophthalmichthys molitrix*), A4 (*Hypophthalmichthys nobilis*), A5 (*Megalobrama terminalis*), A6 (*Parabramis pekinensis*), A7 (*Xenocypris argentea*), A8 (*Squaliobarbus curriculus*), A9 (*Cirrhinus molitorella*), A10 (*Cyprinus carpio*), A11 (*Elopichthys bambusa*), A12 (*Ochetobius elongatus*), A13 (*Siniperca kneri*), A14 (*Sinibotia pulchra*), A15 (*Hemiculter leucisculus*), A16 (*Squalidus argentatus*), A17 (*Pseudolaubuca sinensis*), A18 (*Lucosoma chinensis*), A19 (*Rhinogobius giurinus*)

The equation for *Mylopharyngodon piceus* biomass abundance has a mean relative error of 0.50% and an average relative error of 2.91% across the 103 samples (table 5.13).

TAB. 5.13 – Factors and statistical errors of biomass abundance and the spatial coordinates of community species with the model of fish morphological parameters.

Fish	The power of x - and y -coordinates (the independent variables) in the fitting function	The rejudgment error of fitted samples (%)	The relative error of assessment sample (%)
<i>Mylopharyngodon piceus</i>	All six species of fish XY contained one or two or three power, such as A1, A2, A3, A4, A7, A6. Thirteen species of fish X contain two or two powers, such as A5, A8, A9, A10, A11, A12, A13, A14, A16, A17, A15, A18, A19 <i>Rhinogobius giurinus.</i>	0.50	2.91
<i>Ctenopharyngodon idella</i>	Like <i>Mylopharyngodon piceus</i>	1.06	2.21
<i>Hypophthalmichthys molitrix</i>	Like <i>Mylopharyngodon piceus</i>	0.18	6.03
<i>Hypophthalmichthys nobilis</i>	Like <i>Mylopharyngodon piceus</i>	0.25	2.12
<i>Megalobrama terminalis</i>	Like <i>Mylopharyngodon piceus</i>	0.06	5.04
<i>Parabramis pekinensis</i>	Like <i>Cirrhinus molitorella</i>	0.48	9.64
<i>Xenocypris argentea</i>	Like <i>Mylopharyngodon piceus</i>	2.3	7.63
<i>Squaliobarbus curriculus</i>	XY all contains one, two, and three powers such as A8, A16, A19. X contains one, two powers, and Y contains one, two, and cubic powers such as A1, A2, A3, A4, A5, A6, A7, A9, A10, A11, A12, A13, A14, A15. XY all contains one, two, and three powers such as A17, A18.	8.4	5.49

TAB. 5.13 – (continued).

Fish	The power of x - and y -coordinates (the independent variables) in the fitting function	The rejudgment error of fitted samples (%)	The relative error of assessment sample (%)
<i>Cirrhinus molitorella</i>	XY all contains one, two, and three powers such as A5, A7, A8, A9, A18, A19 . Thirteen species of fish X contain one or three powers, The y contains a quadratic power such as A1, A2, A3, A4, A6, A10, A11, A12, A13, A14, A16, A17, A15 .	0.27	9.21
	Mean relative error	1.23	6.40

The average return error across the 19 fish models was $\leq 23\%$ (*i.e.*, the accuracy was 98.7%; table 5.13). The regression model for all fish was 2.3%, with the exception of *Squaliobarbus curriculus* and *Squalidus argentatus*, which were $>4\%$; *Xenocypris argentea* was $>2\%$. Assessment with nine real-time samples from different years returned an average relative error of 6.4% across the 19 fish species, which better illustrated the validity of the model. This analysis demonstrated that ecological niches could be used to characterize the relationships among fish biomass abundance. Notably, the main reason for the larger error with the predictive sample is that relative error increased with smaller sample sizes. Alternatively, the percentage of outlier data may have increased. The error for several fish, such as *Parabramis pekinensis*, *Cirrhinus molitorella*, *Squalidus argentatus*, *Pseudolaubuca sinensis*, *Lucosoma chinensis*, and *Rhinogobius giurinus*, was 9%–11%. However, this error is within acceptable limits for an ecological study.

In this section, a method to determine the biomass niche based on spatial relationships was established by using multivariate statistical correspondence analysis to convert between biomass abundance and model data. Thus, the biomass abundance of community species at any spatial location can be deduced to evaluate the niche assignments of the model.

5.4.2 Environmental Impact Analysis Reference Frame

Based on the data for the early fish resources in the Zhaoqing section of the Pearl River, the number of various types of replenishments of the early fish resources is constantly changing due to environmental fluctuations. According to monitoring and analysis data, 19 species (taxa) were found among the early drifting resources in the Zhaoqing section of the Pearl River, accounting for about 70% of the biomass of the fishing resources in the study area (Li Xinhui *et al.*, 2021b). It is thus important to study the niche relationships among fish communities in the regional ecological units containing these species. The impacts of annual environmental changes on a fish niche can be characterized by using biomass to reflect abundance or niche occupation. These results are shown in table 5.14. The model values provide a common frame of reference for the impact of annual environmental changes.

Because of the rapid changes in the aquatic ecological environment, fish living conditions and competition parameters have changed greatly. The overall trajectory of community change (the state of deviation from the center of the coordinates; see figure 5.3) can be understood by using the coordinate values of the “integrated impact factors” (*i.e.*, the integrated factors reflecting the changes in various abundance values; see table 5.15). Table 5.15 shows that variations in the abundance of *Squaliobarbus curriculus* are increasing, which is consistent with the trend of the “comprehensive impact factor” away from the origin of the coordinates. This trend may reflect a tendency toward niche expansion in this dominant species. In contrast, the niches of *Mylopharyngodon piceus*, *Ctenopharyngodon idella*, *Hypophthalmichthys molitrix*, *Hypophthalmichthys nobilis*, and other fish were decreasing. Thus, the community niche was in an unbalanced state.

TAB. 5.14 – Abundance of the early floating resources of the main species (classes) in Xi River in the Pearl River system*.

Fish/Time	Model value	2006	2007	2008	2009	2010	2011	2012	2013
<i>Mylopharyngodon piceus</i>	8.07	0.072	0.591	0.205	0.54	0.419	0.071	0.1	0.22
<i>Ctenopharyngodon idella</i>	9.1	2.148	0.29	1.138	1.275	1.194	1.02	1.1	1.3
<i>Hypophthalmichthys molitrix</i>	12.31	2.86	1.078	4.512	2.971	2.205	1.755	2.7	2.9
<i>Hypophthalmichthys nobilis</i>	11.39	1.136	0.124	0.513	0.778	0.572	0.296	1.12	0.99
<i>Megalobrama terminalis</i>	5.715	29.592	29.951	12.132	15.104	11.914	20.337	21.8	4.1
<i>Parabramis pekinensis</i>	4.04	1.208	1.389	1.569	0.573	0.735	0.673	0.55	0.43
<i>Xenocypris argentea</i>	7.019	22.024	19.318	10.44	8.913	26.534	5.878	9.11	5.9
<i>Squaliobarbus curriculus</i>	7.666	26.113	24.842	45.226	46.208	32.506	43.969	49.35	54
<i>Cirrhinus molitorella</i>	7.159	4.584	14.064	8.789	11.549	10.434	10.704	4.49	7.8
<i>Cyprinus carpio</i>	1.312	0.145	0.021	0.103	0.022	0.01	0.01	0.002	0.008
<i>Elopichthys bambusa</i>	4.75	0.32	0.249	0.615	0.4	0.419	0.847	0.66	0.32
<i>Ochetobius elongatus</i>	2.07	0.052	0.052	0.072	0.324	0.316	0.306	0.3	0.123
<i>Siniperca kneri</i>	3.782	0.341	0.093	0.369	0.227	0.204	0.092	0.15	0.15
<i>Sinibotia pulchra</i>	2.79	0.227	0.404	4.451	3.771	2.246	1.184	1.36	2.1
<i>Hemiculter leucisculus</i>	1.834	4.605	5.161	4	2.895	4.033	8.041	3.36	1.1
<i>Squalidus argentatus</i>	6.792	1.105	0.363	2.718	2.409	5.431	3.245	0.9	18
<i>Pseudolaubuca sinensis</i>	1.904	1.642	1.005	1.979	0.951	0.47	0.571	0.39	0.35
<i>Lucosoma chinensis</i>	0.94	1.198	0.591	0.656	0.14	0.092	0.378	0.13	0.077
<i>Rhinogobius giurinus</i>	1.35	0.63	0.415	0.513	0.951	0.265	0.622	0.19	0.134

*of total annual output (%).

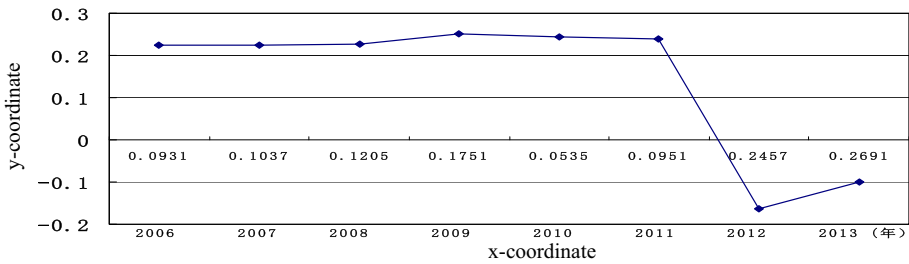


FIG. 5.3 – The X and Y coordinates of the “abundance factor” of 19 fish species of Xijiang River changed from 2006 to 2013.

TAB. 5.15 – The x - and y -coordinates of the “composition factors” of 19 fish species in the Xijiang River from 2006 to 2013.

Time	2006	2007	2008	2009	2010	2011	2012	2013
x	0.0931	0.1037	0.1205	0.1751	0.0535	0.0951	0.2457	0.2691
y	0.2252	0.2244	0.2268	0.251	0.2439	0.2399	-0.164	-0.1

5.4.3 Analysis of the Niche of the Artificial Community

Changes in the niche of a specific species will lead to various niche changes across the community. In reality, river biodiversity and resource recovery conservation efforts usually target specific species, such as the silver carp and bighead carp, in lakes with high primary productivity. However, due to the serious declines in river fishery resources, it is necessary to evaluate the niche structure and environmental capacity of certain species in the community by investing in black carp, grass carp, silver carp, and bighead carp. There are also random releases of species, which require systematic evaluation. In an ecosystem, an increase in the biomass of one species will affect the niches of other species in the community. If a given increase is not suitable for the food chain, new imbalances in the ecosystem may arise.

The species community niche structure is different from the concept of the environmental capacity of a given species. The species community niche structure is based on the composition of the food chain system and the niche value of each species, and this concept emphasizes the biological match between the niche and the energy cycle. The environmental capacity of a species refers to the environmental matching between the ecological niche and the energy cycle based on the total energy requirement.

Using the model, we can analyze and calculate the preset target amounts of biodiversity conservation and resource restoration for one species (or several species) while observing the niche changes of other species (species matching and biomass matching). This provides an analytical tool for community management. Using different fish as targets, the resulting niche composition of the remaining fish in the community was calculated to reflect the function of the model, as shown in table 5.16.

TAB. 5.16 – Simulated community niches targeting different fish (%).

Fish/sample name	Model 1	Model 2	Model 3	Model 4	Model 5	Model 6	Model 7	Model 8	Model 9
<i>Mylopharyngodon piceus</i>	1.19	7.03	8.8	14.6	8.89	15.15	13	13.5	13.9
<i>Ctenopharyngodon idella</i>	1.4	10.03	12	11	12.5	17.9	16	16.5	15.28
<i>Hypophthalmichthys molitrix</i>	12.95	9.18	8.5	10.3	16	11.5	10	11.5	16.76
<i>Hypophthalmichthys nobilis</i>	11.87	10.88	9.3	11.3	12	9.2	15	15.5	16.66
Total target ecological niche of <i>Mylopharyngodon piceus</i> , <i>Ctenopharyngodon idella</i> , <i>Hypophthalmichthys molitrix</i> , <i>Hypophthalmichthys nobilis</i>	27.41	37.12	38.6	47.2	49.39	53.75	54	57	62.6
<i>Megalobrama terminalis</i>	6.47	5.14	5.14	3.6	4.27	3.9	4.99	3.99	3.39
<i>Parabramis pekinensis</i>	4.64	1.03	2.03	1.4	3	3.2	1.02	1.02	1.7
<i>Xenocypris argentea</i>	9.94	8.45	7	7.6	5.5	3.7	5.5	4.5	2.14
<i>Squaliobarbus curriculus</i>	9.93	9.25	8	4.2	4	2.8	5.99	4.99	3.71
<i>Cirrhinus molitorella</i>	9.6	2.85	3.63	3.1	4.72	2.75	4.7	3.7	5.52
<i>Cyprinus carpio</i>	1.62	0.71	1.5	0.58	0.28	0.28	1.1	1.1	1.38
<i>Elopichthys bambusa</i>	4.02	4.44	3.8	4.91	1.5	6.4	2.6	2.6	1.08
<i>Ochetobius elongatus</i>	0.54	4.57	3.7	2.6	3	4.3	1.18	1.68	2.65
<i>Siniperca kneri</i>	5.69	4.57	5	4.4	2	5.5	2.04	2.04	2.42
<i>Sinibotia pulchra</i>	4.62	2.4	3.2	3.6	1.9	4.3	1.39	1.89	1.36
<i>Hemiculter leucisculus</i>	2.16	2.68	1.6	3.7	5.2	2.2	4.64	4.14	1.91
<i>Squalidus argentatus</i>	8.3	7.65	6.2	4.6	5.2	3.05	4.63	4.13	6.32
<i>Pseudolaubuca sinensis</i>	2.16	3.85	4.1	5.4	4	1.85	3.53	3.53	1.7
<i>Lucosoma chinensis</i>	1.65	3.63	4	0.77	3.04	1.45	1.63	2.13	1.59
<i>Rhinogobius giurinus</i>	1.29	1.66	2.5	2.34	3	0.21	1.06	1.56	0.53

References

- Aarssen L. W., Schamp B. S., Pither J. (2006) Why are there so many small plants? Implications for species coexistence, *J. Ecol.* **94**, 569.
- Abellán P., Bilton D., Millán A., *et al.* (2006) Can taxonomic distinctness assess anthropogenic impacts in inland waters? A case study from a Mediterranean river basin, *Freshw. Biol.* **51**, 1744.
- Acevedo P., Jiménez-Valverde A., Lobo J. M., Real R. (2012) Delimiting the geographical background in species distribution modelling [J], *J. Biogeogr.* **39** (8), 1383.
- Agrawal V. P., Sastry K. V., Kaushab S. K. (1975) Digestive enzymes of three teleost fishes [J], *Acta Physiol. Acad. Sci. Hungaricae* **46** (2), 93.
- Akira H., Mitsuru E., Naohiro T., *et al.* (1987) Primary structure of human pancreatic α -amylase gene: Its comparison with human salivary α -amylase gene [J], *Gene* **60** (1), 57.
- Allesina S., Tang S. (2012) Stability criteria for complex ecosystems (Article), *Nature* **483** (7388), 205. <https://doi.org/10.1038/nature10832>.
- Anderson R. P., Raza A. (2010) The effect of the extent of the study region on GIS models of species geographic distributions and estimates of niche evolution: Preliminary tests with montane rodents (genus *Nephelomys*) in Venezuela, *J. Biogeogr.* **37** (7), 1378.
- Arnone M. I., Davidson E. H. (1997) The hardwiring of development: Organization and function of genomic regulatory systems, *Development* **124** (10), 1851.
- Austin M. (2007) Species distribution models and ecological theory: A critical assessment and some possible new approaches [J], *Ecol. Model.* **200** (1–2), 1.
- Austin M. P. (2002) Spatial prediction of species distribution: An interface between ecological theory and statistical modeling, *Ecol. Model.* **157** (2/3), 101.
- Azzurro E., Tuset V. M., Lombarte A., Maynou F., Simberloff D., Rodríguez-Pérez A., Solé R. V. (2014) External morphology explains the success of biological invasions, *Ecol. Lett.* **17** (11), 1455.
- Ba J., Deng H., Duan X., Liu S., Li Y., Chen D. (2015) Stability isotope (^{13}C , ^{15}N) technology was applied to study the nutritional level of major fish in the middle reaches of the Yangtze River [J], *J. Zool.* **4**, 537 (巴家文, 邓华堂, 段辛斌, 刘绍平, 李云, 陈大庆. 应用稳定性同位素($\delta^{13}\text{C}$ 、 $\delta^{15}\text{N}$)技术研究长江中游干流主要鱼类的营养级 [J], *动物学杂志*, 2015, **4**, 537).
- Banse K. (2007) Do we live in a largely top-down regulated world? *J. Biosci.* **32** (4), 791.
- Barbet-Massin M., Jiguet F., Albert C. H., Thuiller W. (2012) Selecting pseudo-absences for species distribution models: How, where and how many? [J], *Methods Ecol. Evol.* **3** (2), 327.

- Batchelder H. P., Edwards C. A., Powell T. M. (2002) Individual-based models of copepod populations in coastal upwelling region: Implications of physiologically and environmentally influenced diel vertical migration on demographic success and nearshore retention [J], *Prog. Oceanogr.* **53**, 307.
- Bellwood D. R., Wainwright P. C., Fulton C. J., *et al.* (2002) Assembly rules and functional groups at global biogeographical scales, *Funct. Ecol.* **16**, 557.
- Bertness M. D., Leonard G. H. (1997) The role of positive interactions in communities: Lessons from intertidal habitats [J], *Ecology* **78**, 1976.
- Blackburn T. M., Gaston K. J., Quinn R. M., *et al.* (1997) Of mice and wrens: The relation between abundance and geographic range size in British mammals and birds, *Phil. Trans. R. Soc. B* **352**, 419.
- Blanchet S., Grenouillet G., Beauchard O., Tedesco P. A., Leprieur F., Durr H. H., Busson F., Oberdorff T., Brosse S. (2010) Nonnative species disrupt the worldwide patterns of freshwater fish body size: Implications for Bergmann's rule, *Ecol. Lett.* **13** (4), 421.
- Bowman R. E. (1986) Effect of regurgitation on stomach content data of marine fisheries [J], *Environ. Biol. Fish* **16**, 171.
- Britton J. R., Davies G. D., Harrod C. (2010) Trophic interactions and consequent impacts of the invasive fish *Pseudorasbora parva* in an ativeaquatc food web: A field investigation in the UK, *Biol. Invasions* **12** (6), 1533.
- Bronstein J. L. (1994) Our current understanding of mutualism, *Q. Rev. Biol.* **69** (1), 31.
- Brown J. H., Stevens G. C., Kaufman D. M. (1996) The geographic range: Size, shape, boundaries, and internal structure, *Annu. Rev. Ecol. Syst.* **27**, 597.
- Bruno J. F., Stachowicz J. J., Bertness M. D. (2003) Inclusion of facilitation into ecological theory, *Trends Ecol. Evol.* **18** (3), 119.
- Burns J. H., Strauss S. Y. (2011) More closely related species are more ecologically similar in an experimental test [J], *Proc. Nat. Acad. Sci. U.S.A.* **108** (13), 5302.
- Cai L., Niu J., Liu C., Zou M., Xie P., Adakbek, Liu J., Li H. (2017) Species diversity and dominant species of fish in different reaches of Yili River, Xinjiang, *Aquat. Biol. J.* **4**, 819 (蔡林钢, 牛建功, 刘春池, 邹明, 谢鹏, 阿达克·白克, 刘建, 李红, 新疆伊犁河不同河段鱼类的物种多样性和优势种, 水生生物学报, 2017, 4, 819).
- Cai W. (2013) *Fish map in the Yellow River Basin*. Northwest A & F University Press (蔡文仙, 黄河流域鱼类图志, 西北农林科技大学出版社, 2013年).
- Cai Y., Liao Z. (2002) Progress of soil microbial ecology, *J. Ecol. Environ.* **11**, 167 (蔡燕飞, 廖宗文. 土壤微生物生态学研究方法进展. 生态环境学报, 2002, 11, 167).
- Cameron T. (2002) The year of the 'diversity-ecosystem function' debate, *Trends Ecol. Evol.* **17** (11), 495.
- Cao P., He I. (2015) Theoretical framework of microbial ecology, *J. Ecol.* **35**, 6 (曹鹏, 贺纪正. 微生物生态学理论框架. 生态学报, 2015, 35, 6).
- Carreonmartinez L. B., Heath D. D. (2010) Revolution in food web analysis and trophic ecology: Diet analysis by DNA and stable isotope analysis, *Mol. Ecol.* **19**, 25.
- Casparly W. F. (1992) Physiology and pathophysiology of intestinal absorption, *Am. J. Clin. Nutr.* **55** (1 Suppl), 299S.
- Castiglione S., Mondanaro A., Melchionna M., *et al.* (2017) Diversification rates and the evolution of species range size frequency distribution, *Front. Ecol. Evol.* **5**. <https://doi.org/10.3389/fevo.2017.00147>.

- Cavender-Bares J., Keen A., Miles B. (2006) Phylogenetic structure of floridian plant communities depends on taxonomic and spatial scale [J], *Ecology* **87** (7), 109.
- Chai Y., Yue M. (2016) Progress in the mechanism of plant community construction [J], *J. Ecol.* **15**, 4557 (柴永福, 岳明. 植物群落构建机制研究进展 [J], *生态学报*, 2016, **15**, 4557).
- Chave J. (2004) REVIEW Neutral theory and community ecology, *Ecol. Lett.* **7**, 241.
- Chen C. (2007) *Study on cDNA cloning and tissue expression of mulmine-amylase* [D]. Southwest University, Chongqing (陈春娜. 胭脂鱼 α -淀粉酶的 cDNA 克隆与组织表达研究[D]. 重庆: 西南大学, 2007).
- Chen L., Liang X., Wang L., et al. (2009) Cloning and sequence analysis of fish trypsinase, large-eye mandarin *Siniperca kneri* Garman, 1912, and pepsinogen genes [J], *Chin. J. Biochem. Mol. Biol.* **25** (12), 1115 (陈亮, 梁旭方, 王琳, 等. 大眼鳊 *Siniperca kneri* Garman, 1912 鱼胰蛋白酶和淀粉酶与胃蛋白酶原基因的克隆与序列分析 [J], *中国生物化学与分子生物学报*, 2009, **25** (12), 1115).
- Chen M., Wei Z., Tian L., et al. (2021) Construction and application of synthetic microbial communities, *Sci. Bull.* **66**, 273 (陈沫先, 韦中, 田亮, 等. 合成微生物群落的构建与应用. *科学通报*, 2021, **66**, 273).
- Chesson P. (2018) Updates on mechanisms of maintenance of species diversity, *J. Ecol.* **106** (5), 1773.
- Choudoir M. J., Barberán A., Menninger H. L., et al. (2018) Variation in range size and dispersal capabilities of microbial taxa, *Ecology* **99**, 322.
- Chu X., Chen Y., et al. (1989) *On the fish records in Yunnan*. Science Press, Beijing (褚新洛, 陈银瑞等, 云南鱼类志 上, 科学出版社, 北京, 1989).
- Chu X., Chen Y., et al. (1990) *The annals of Yunnan fish*. Science Press, Beijing (褚新洛, 陈银瑞等, 云南鱼类志 下, 科学出版社, 北京, 1990).
- Clarke K., Warwick R. (1999) The taxonomic distinctness measure of biodiversity: Weighting of step lengths between hierarchical levels, *Mar. Ecol. Prog. Ser.* **184**, 21.
- Cockell M., Stevenson B. J., Strubin M., Hagenbuchle O., Wellauer P. K. (1989) Identification of a cell-specific DNA-binding activity that interacts with a transcriptional activator of genes expressed in the acinar pancreas, *Mol. Cell. Biol.* **9** (6), 2464.
- Cockell M., Stolarczyk D., Frutiger S., Hughes G. J., Hagenbuchle O., Wellauer P. K. (1995) Binding sites for hepatocyte nuclear factor 3 beta or 3 gamma and pancreas transcription factor 1 are required for efficient expression of the gene encoding pancreatic alpha-amylase, *Mol. Cell. Biol.* **15** (4), 1933.
- Connell J. H. (1978) Diversity in tropical rain forests and coral reefs: High diversity of trees and corals is maintained only in a nonequilibrium state [J], *Science* **199**, 1302.
- Convertino M. (2011) Neutral metacommunity clustering and SAR: River basin vs. 2-D landscape biodiversity patterns, *Ecol. Model.* **222** (11), 1863.
- Cucherousset J., Blanchet S., Olden J. D. (2012) Non-nativespecies promote triphic dispersion of food webs, *Front. Ecol. Environ.* **10** (8), 406.
- Darias J., Murray H. M., Gallant J. W., Astola A., Douglas S. E., Yufera M., Martinez-Rodriguez G. (2006) Characterization of a partial alpha-amylase clone from red porgy (*Pagruspagrus*): Expression during larval development, *Compar. Biochem. Physiol. B* **143** (2), 209.
- Davis C. L., Rich L. N., Farris Z. J., Kelly M. J., Di Bitetti M. S., Blanco Y. D., Albanesi S., Farhadinia M. S., Gholikhani N., Hamel S., Harmsen B. J., Wultsch C., Kane M. D., Martins Q., Murphy A. J., Steenweg R., Sunarto S., Taktehrani A., Thapa K., Tucker J. M., Whittington J., Widodo F. A., Yoccoz N. G., Miller D. A. W. (2018) Ecological correlates of the spatial co-occurrence of sympatric mammalian carnivores worldwide, *Ecol. Lett.* **21**, 1401.

- de Mazancourt C. (2001) The unified neutral theory of biodiversity and biogeography, *Science* **293** (5536), 1772.
- de Satgé J., Teichman K., Cristescu B. (2017) Competition and coexistence in a small carnivore guild, *Oecologia* **184**, 873.
- Devictor V., Mouillot D., Meynard C., Jiguet F., Thuiller W., Mouquet N. (2010) Spatial mismatch and congruence between taxonomic, phylogenetic and functional diversity: The need for integrative conservation strategies in a changing world, *Ecol. Lett.* **13** (8), 1030.
- Díaz S., Cabido M. (2001) Vival difference: Plant functional diversity matters to ecosystem processes, *Trends Ecol. Evol.* **16** (11), 646.
- Diehl S. (1992) Fish predation and benthic community structure: The role of omnivory and habitat complexity, *Ecology* **73** (5), 1646.
- Donadio E., Buskirk S. W. (2006) Diet, morphology, and interspecific killing in Carnivora, *Am. Nat.* **167**, 524.
- Dong C., Xia Z., Jiang Z., Zhao C., Tian F., Jiang W., Qu X., Wang J. (1996a) Fish composition characteristics of the Mohe River section upstream of Heilongjiang, *Heilongjiang Aquat. Prod.* **4**, 19 (董崇智, 夏重志, 姜作发, 赵春刚, 田富河, 蒋文革, 曲孝忠, 王军, 黑龙江上游漠河江段的鱼类组成特征, 黑龙江水产, 1996, 4, 19).
- Dong C., Zhao C., Jin Z., Xie F., Wang W. (1996b) Preliminary study of Suifenhe Fish Zone, *Chin. Sci. Fish.* **4**, 125 (董崇智, 赵春刚, 金贞礼, 解福君, 王维坤, 绥芬河鱼类区系初步研究, 中国水产科学, 1996, 4, 125).
- Douglas S. E., Mandla S., Gallant J. W. (2000) Molecular analysis of the amylase gene and its expression during development in the winter flounder, *Pleuronectes americanus* [J], *Aquaculture* **190** (3–4), 247.
- Drake J. M., Randin C., Guisan A. (2006) Modelling ecological niches with support vector machines [J], *J. Appl. Ecol.* **43** (3), 424.
- Drenner R. W., Smith J. D., Threlkeld S. T. (1996) Lake trophic state and the limnological effects of omnivorous fish, *Hydrobiologia* **319** (3), 213.
- Dudei N. L., Stigall A. L. (2010) Using ecological niche modeling to assess biogeographic and niche response of brachiopod species to the Richmondian Invasion (Late Ordovician) in the Cincinnati Arch [J], *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **296** (1–2), 28.
- Duffy J. E. (2003) Biodiversity loss, trophic skew and ecosystem functioning, *Ecol. Lett.* **6** (8), 680.
- Dukes J. S. (2001) Biodiversity and invasibility in grassland microcosm, *Ecologia* **126** (4), 563.
- Dunne J. A., Williams R. J., Martinez N. D. (2002) Network structure and biodiversity loss in food webs: Robustness increases with connectance [J], *Ecol. Lett.* **5** (4), 558.
- Ebeling S. K., Welk E., Auge H., Bruelheide H. (2008) Predicting the spread of an invasive plant: Combining experiments and ecological niche model, *Ecography* **31**, 709.
- Editorial Committee of Fishery Resources Survey of Pearl River System, Fishery Resources Survey and Research Report of Pearl River System, Volume 3, 1985 (珠江水系渔业资源调查编委会, 珠江水系渔业资源调查研究报告, 第三分册, 1985).
- Editorial Committee of Report on the investigation of the natural resources of the Guangxi Zhuang Autonomous Region inland waters fishery, Guangxi Fisheries Research Institute, 1985 (广西水产研究所, 广西壮族自治区内陆水域渔业自然资源调查研究报告, 1985).
- Elith J., Graham C. H., Anderson R. P., Dudik M., Ferrier S., Guisan A., Hijmans R. J., Huettmann F., Leathwick J. R., Lehmann A., Li J., Lohmann L. G., Loiselle B. A., Manion G., Moritz C., Nakamura M., Nakazawa Y., Overton J. M., Peterson A. T., Phillips S. J.,

- Richardson K. S., Scachetti-Pereira R., Schapire R. E., Soberón J., Williams S., Wisz M. S., Zimmermann N. E. (2006) Novel methods improve prediction of species' distributions from occurrence data, *Ecography* **29** (2), 129.
- Eliot J., Leathwick J. R. (2009) Species distribution models: Ecological explanation and prediction across space and time, *Annu. Rev. Ecol. Evol. Syst.* **40**, 677.
- Enquist B. J., Haskell J. P., Tiffney B. H. (2002) General patterns of taxonomic and biomass partitioning in extant and fossil plant communities [J], *Nature* **419**, 610.
- Erős T., Sály P., Takács P., *et al.* (2012) Temporal variability in the spatial and environmental determinants of functional metacommunity organization-stream fish in a human-modified landscape, *Freshw. Biol.* **57**, 1914.
- Farris Z. J., Gerber B. D., Karpanty S., Murphy A., Wampole E., Ratelolahy F., Kelly M. J. (2020) Exploring and interpreting spatiotemporal interactions between native and invasive carnivores across a gradient of rainforest degradation, *Biol. Invasions* **22**, 2033.
- Fausch K. D., Torgersen C. E., Baxter C. V., *et al.* (2002) Landscapes to riverscapes: Bridging the gap between research and conservation of stream fishes a continuous view of the river is needed to understand how processes interacting among scales set the context for stream fishes and their habitat, *BioScience* **52** (6), 483.
- Fisheries Research Institute, Guangxi Zhuang Autonomous Region, Institute of Zoology, Chinese Academy of Sciences. (2006) *Guangxi freshwater fish annals*, 2nd edn. Guangxi People's Publishing House, Nanning (广西壮族自治区水产研究所, 中国科学院动物研究所, 广西淡水鱼类志 第2版, 广西人民出版社, 南宁, 2006).
- Fritschie K. J., Cardinale B. J., Alexandrou M. A., Oakley T. H. (2014) Evolutionary history and the strength of species interactions: Testing the phylogenetic limiting similarity hypothesis, *Ecology* **95** (5) 1407.
- Gaston K. J. (1996) Species-range-size distributions: Patterns, mechanisms and implications, *Trends Ecol. Evol.* **11**, 197.
- Gaston K. J. (2009) Geographic range limits of species, *Proc. R. Soc. B-Biol. Sci.* **276**, 1391.
- Gaston K. J., Fuller R. A. (2009) The sizes of species' geographic ranges, *J. Appl. Ecol.* **46** (1), 1.
- German D. P., Foti D. M., Heras J., *et al.* (2016) Elevated gene copy number does not always explain elevated amylase activities in fishes [J], *Physiol. Biochem. Zool.* **89** (4), 277.
- Glor R. E., Warren D. (2011) Testing ecological explanations for biogeographic boundaries, *Evolution* **65**, 673.
- Goel A., Mathupala S. P., Pedersen P. L. (2003) Glucose metabolism in cancer [J], *J. Biol. Chem.* **278** (17), 15333.
- Gómez-Ortiz Y., Monroy-Vilchis O., Mendoza-Martínez G. D. (2015) Feeding interactions in an assemblage of terrestrial carnivores in central Mexico, *Zool. Stud.* **54**, 16.
- Gotelli N. J., Engstrom R. T. (2003) Predicting species occurrences: Issues of accuracy and scale [J], *The Auk* **120** (4), 1199.
- Grange S., Duncan P. (2006) Bottom-up and top-down processes in African ungulate communities: Resources and predation acting on the relative abundance of zebra and grazing bovids, *Ecography* **29** (6), 899.
- Gravel D., Canham C. D., Beaudet M., Messier C. (2006) Reconciling niche and neutrality: The continuum hypothesis [J], *Ecol. Lett.* **9**, 399.
- Green R. H. (1971) A multivariate statistical approach to the Hutchinsonian niche: Bivalve molluscs of central Canada, *Ecology* **52**, 544.

- Grenouillet G., Pont D., Hérissé C. (2004) Within-basin fish assemblage structure: The relative influence of habitat versus stream spatial position on local species richness, *Can. J. Fish. Aquat. Sci.* **61** (1), 93.
- Gtzenberger L., Bello F., Brathen K. A., *et al.* (2012) Ecological assembly rules in plant communities—approaches, patterns and prospects, *Biol. Rev.* **87**, 111.
- Guisan A., Thuiller W. (2005) Predicting species distribution: Offering more than simple habitat models, *Ecol. Lett.* **8**, 993.
- Guisan A., Zimmermann N. E. (2000) Predictive habitat distribution models in ecology, *Ecol. Model.* **135**, 147.
- Hall A. R., Ashby B., Bascompte J., King K. C. (2020) Measuring coevolutionary dynamics in species-rich communities, *Trends Ecol. Evol.* **35** (6), 539.
- Hanski I. (1999) Habitat connectivity, habitat continuity, and metapopulations in dynamic landscapes, *Oikos* **87** (2), 209.
- Hardy C. M., Krull E. S., Hartley D. M., Oliver R. L. (2010) Carbon source accounting for fish using combined DNA and stable isotope analyses in a regulated lowland river weir pool, *Mol. Ecol.* **19**, 197.
- Heikkinen R. K., Marmion M., Luoto M. (2012) Does the interpolation accuracy of species distribution models come at the expense of transferability? [J], *Ecography* **35** (3), 276.
- Heino J., Melo A. S., Siqueira T., Soininen J., Valanko S., Bini L. M. (2015) Metacommunity organisation, spatial extent and dispersal in aquatic systems: Patterns, processes and prospects [J], *Freshw. Biol.* **60** (5), 845.
- Heino J., Soininen J., Lappalainen J., *et al.* (2005) The relationship between species richness and taxonomic distinctness in freshwater organisms, *Limnol. Oceanogr.* **50**, 978.
- Henriques-Silva R., Lindo Z., Peres-Neto P. R. (2013) A community of metacommunities: Exploring patterns in species distributions across large geographical areas, *Ecology* **94** (3), 627.
- Hidalgo M. C., Urea E., Sanz A. (1999) Comparative study of digestive enzymes in fish with different nutritional habits. Proteolytic and amylase activities [J], *Aquaculture* **170** (3–4), 267.
- Hirzel A. H., Hausser J., Chessel D., *et al.* (2002) Ecological niche factor analysis: How to compute habitat-suitability maps without absence data, *Ecology* **83**, 2027.
- Hoeinghaus D. J., Winemiller K. O., Birnbaum J. S. (2007) Local and regional determinants of stream fish assemblage structure: Inferences based on taxonomic vs. functional groups, *J. Biogeogr.* **34**, 324. <https://doi.org/10.1111/j.1365-2699.2006.01587.x>.
- Hoffmeister A., Ropolo A., Vasseur S., *et al.* (2002) The HMG-I/Y-related protein p8 binds to p300 and Pax2 trans-activation domain-interacting protein to regulate the trans-activation activity of the Pax2A and Pax2B transcription factors on the glucagon gene promoter [J], *J. Biol. Chem.* **277** (25), 22314.
- Hooper D. U., Solan M., Symstad A., Diaz S., Gessner M. O., Buchmann N., Degrange V., Grime P., Hulot F., Mermillod-Blondin F., Roy J., Spehn E., van Peer L. (2002) Species diversity, functional diversity, and ecosystem functioning, *Biodiversity and ecosystem functioning: Synthesis and perspectives* (M. Loreau, S. Naeem, P. Inchausti, Eds). Oxford University Press, Oxford, pp. 195–208.
- Hu Z., Gallo S. M. (2010) Identification of interacting transcription factors regulating tissue gene expression in human, *BMC Genomics* **11**, 49.
- Hu Z., Hu B., Collins J. F. (2007) Prediction of synergistic transcription factors by function conservation, *Genome Biol.* **8** (12), R257.

- Huang G., Guo Y., Li L., Fan S., Yu Z., Yu D. (2016) Genomic structure of the alpha amylase gene in the pearl oyster *Pinctada fucata* and its expression in response to salinity and food concentration, *Gene* **587** (1), 98.
- Hubbell S. P. (2001) *The unified neutral theory of biodiversity and biogeography*. Princeton University Press, Princeton, pp. 1–151.
- Hunan Fisheries Research Institute. (1977) *Hunan fish annals*. Human People's Publishing House, Changsha (湖南省水产科学研究所, 湖南鱼类志, 湖南人民出版社, 长沙, 1977).
- Ings T. C., Montoya J. M., Bascompte J., *et al.* (2009) Review: Ecological networks – Beyond food webs [J], *J. Anim. Ecol.* **78** (1), 253.
- Inostroza-Michael O., Hernández C. E., Rodríguez-Serrano E., *et al.* (2018) Interspecific geographic range size-body size relationship and the diversification dynamics of Neotropical Furnariid birds, *Evolution* **72**, 1124.
- Institute of Zoology, Chinese Academy of Sciences, Xinjiang Institute of Biological Desert, Chinese Academy of Sciences, Bureau of Fisheries of Xinjiang Uygur Autonomous Region. (1979) *Xinjiang fish records*. Xinjiang People's Publishing House (中国科学院动物研究所, 中国科学院新疆生物沙漠研究所, 新疆维吾尔自治区水产局等, 新疆鱼类志, 新疆人民出版社, 1979.03, 乌鲁木齐).
- Jackson D. A., Peres-Neto P. R., Olden J. D. (2001) What controls who is where in freshwater fish communities—the roles of biotic, abiotic, and spatial factors, *Can. J. Fish. Aquat. Sci.* **58** (1), 157.
- Jackson S. T., Overpeck J. T. (2000) Responses of plant populations and communities to environmental changes of the late quaternary, *Paleobiology* **26**, 194.
- Jacobson B., Peres-Neto P. R. (2010) Quantifying and disentangling dispersal in metacommunities: How close have we come? How far is there to go? *Landscape Ecol.* **25** (4), 495.
- Jennings S., Reynolds J. D., Mills S. C. (1998) Life history correlates of responses to fisheries exploitation [J], *Proc. R. Soc. London. Ser. B: Biol. Sci.* **265** (1393), 333.
- Jiang X., Zhang W. (2010) Functional diversity and its research methodology, *J. Ecol.* **30** (10), 2766 (江小雷, 张卫国. 功能多样性及其研究方法. 生态学报, 2010, **30** (10), 2766).
- Kareiva P., Washington U. (1995) Connecting landscape patterns to ecosystem and population processes, *Nature* **373** (65120), 299.
- Kawata H., Yamada K., Shou Z. F., *et al.* (2003) Zinc-fingers and homeoboxes (ZHX) 2, a novel member of the ZHX family, functions as a transcriptional repressor [J], *Biochem. J.* **373** (3), 747.
- Kéry M., Gardner B., Monnerat C. (2010) Predicting species distributions from checklist data using site-occupancy models [J], *J. Biogeogr.* **37** (10), 1851.
- Kraft N. J. B., Ackerly D. D. (2010) Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest [J], *Ecol. Monogr.* **80** (3), 401.
- Kronfeld-Schor N., Dayan T. (2003) Partitioning of time as an ecological resource, *Annu. Rev. Ecol. Evol. Syst.* **34**, 153.
- Lanszki J., Heltai M., Kövér G., Zalewski A. (2019) Non-linear relationship between body size of terrestrial carnivores and their trophic niche breadth and overlap, *Basic Appl. Ecol.* **38**, 36.
- Larson E. R., Olden J. D. (2012) Using avatar species to model the potential distribution of emerging invaders, *Glob. Ecol. Biogeogr.* **21** (11), 1114.
- Leibold M. A. (1998) Similarity and local coexistence of species in regional biotas [J], *Evol. Ecol.* **12** (1), 95.
- Leibold M. A., Holyoak M., Mouquet N., *et al.* (2004) The metacommunity concept: A framework for multi-scale community ecology, *Ecol. Lett.* **7** (7), 601.

- Leonard D., Clarke K., Somerfield P., *et al.* (2006) The application of an indicator based on taxonomic distinctness for UK marine biodiversity assessment, *J. Environ. Manage.* **78**, 52.
- Lepš J., Brown V. K., Diaz Len T. A., Gormsen D., Hedlund K., Kailová J., Korthals G. W., Mortimer S. R., Rodriguez-Barrueco C., Roy J., Santa Regina I., van Dijk C., Vander Putten W. H. (2001) Separating the chance effect from other diversity effects in the functioning of plant communities, *Oikos* **92** (1), 123.
- Leps J., deBello F., Lavori S., Berman S. (2006) Quantifying and interpreting functional diversity of natural communities: Practical considerations matter, *Preslia* **78** (4), 481.
- Lester S. E., Ruttenberg B. I., Gaines S. D., *et al.* (2007) The relationship between dispersal ability and geographic range size, *Ecol. Lett.* **10**, 745.
- Li G., Feng C., Tang Y., Zhang R., Zhao K. (2017) Indigenous fish resources, *J. Gansu Agric. Univ.* **3**, 22 (李国刚, 冯晨光, 汤永涛, 张仁意, 赵凯, 新疆内陆河土著鱼类资源调查, 甘肃农业大学学报, 2017, **3**, 22).
- Li J., Li X., *etc.*, Jia X., *et al.* (2010) Diversity and evolution of fish communities in Xijiang [J], *Chin. Aquat. Sci.* **17** (1), 298 (李捷, 李新辉等, 贾晓平等. 西江鱼类群落其多样性及演变 [J], 中国水产科学, 2010, **17** (1), 298).
- Li S. (2015) *Yellow river fish annals: Selected yellow fish specialty and fish studies*. Fisheries Publishing House (李思忠, 《黄河鱼类志: 黄河鱼类专著及鱼类学文选》, 水产出版社, 2015年).
- Li S., Jin T., Shi Y. (2000) Fish resources survey in inner Mongolia, *J. Zhelimu Anim. Husb. Coll.* **10** (3), 24 (李树国, 金天明, 石玉华, 内蒙古鱼类资源调查, 哲里木畜牧学院学报, 2000, **10** (3), 24).
- Li X., Lai Z., Li Y., Tong C. (2021a) *River ecological series – Functional research on river fish*. Science Press, Beijing (李新辉, 赖子尼, 李跃飞, 童朝锋, 河流生态丛书-江河鱼类产卵场功能研究, 科学出版社, 2021a, 北京).
- Li X., Li J., Li Y. (2020f) *River ecology series of Freshwater and estuine Fish in Hainan Island*. Science Press, Beijing (李新辉, 李捷, 李跃飞, 河流生态丛书-海南岛淡水及河口鱼类原色图鉴, 科学出版社, 2020f, 北京).
- Li X., Li Y., Wu Z. (2020e) *River ecological series – The monitoring log of drifting fish eggs and larvae in the Zhaoqing section of the Pearl River (2010)* [M]. Science Press, Beijing (李新辉, 李跃飞, 武智, 2020e, 河流生态丛书-珠江肇庆段漂流性鱼卵、仔鱼监测日志 (2010) [M]. 北京: 科学出版社).
- Li X., Li Y., Yang J. (2020b) *River ecological series – The monitoring log of drifting fish eggs and larvae in the Zhaoqing section of the Pearl River (2007)* [M]. Science Press, Beijing (李新辉, 李跃飞, 杨计平, 2020b, 河流生态丛书-珠江肇庆段漂流性鱼卵、仔鱼监测日志 (2007) [M]. 北京: 科学出版社).
- Li X., Li Y., Zhang Y. (2020a) *River ecological series – The monitoring log of drifting fish eggs and larvae in the Zhaoqing section of the Pearl River (2006)* [M]. Science Press, Beijing (李新辉, 李跃飞, 张迎秋, 2020a, 河流生态丛书-珠江肇庆段漂流性鱼卵、仔鱼监测日志 (2006). 北京: 科学出版社).
- Li X., Li Y., Zhang Y. (2020d) *River ecological series – The monitoring log of drifting fish eggs and larvae in the Zhaoqing section of the Pearl River (2009)* [M] (李新辉, 李跃飞, 张迎秋, 2020d, 河流生态丛书-珠江肇庆段漂流性鱼卵、仔鱼监测日志 (2009). 北京: 科学出版社).
- Li X., Li Y., Zhu S. (2020c) *River ecological series – The monitoring log of drifting fish eggs and larvae in the Zhaoqing section of the Pearl River (2008)* [M]. Science Press, Beijing (李新辉, 李跃飞, 朱书礼, 2020c, 河流生态丛书-珠江肇庆段漂流性鱼卵、仔鱼监测日志 (2008). 北京: 科学出版社).
- Li X., Yang J., Zhu S., Li Y., Chen W., Hu Z. (2020) Insight into the combinatorial transcriptional regulation on α -amylase gene in animal groups with different dietary nutrient content, *Genomics* **112**, 520.

- Li X.-H., Lai Z.-N., Yu Y.-M. (2022) *River ecology series – Ecology of Fish Community: Niche Modeling Based on Fish Morphological Parameters*. Science Press, Beijing (李新辉, 赖子尼, 余煜棉, 河流生态丛书-鱼类形态学模型与群落研究, 科学出版社, 2022, 北京).
- Li X.-H., Li Y.-F., Tan X.-C. (2021b) *River ecology series – Studies on early fish resources in rivers*. Science Press, Beijing (李新辉, 李跃飞, 谭细物, 河流生态丛书-江河鱼类早期资源研究, 科学出版社, 2021b, 北京).
- Li Y., Li X., Tan X., Li J., Wang C., Luo J. (2008) The current status quo and changes of fishery resources in Zhaoqing River Section of Xijiang River, *Water Conservancy Fish.* **28** (2), 80 (李跃飞, 李新辉, 谭细物, 李捷, 王超, 罗建仁, 西江肇庆江段渔业资源现状及其变化, 2008, 水利渔业, 2008, **28** (2), 80).
- Li Z., Duoolian, Li S., Wang T. (2021) Overview of competition and coexistence of terrestrial-predators [J], *Biodiversity* **1**, 81 (李治霖, 多立安, 李晟, 王天明, 陆生食肉动物竞争与共存研究概述 [J], 生物多样性, 2021, **1**, 81).
- Lin K., Mai G., Wang L., Wang X. (2020) Fish community structure and its stability in the coastalwaters of the Pearl River Estuary from 2015 to 2018 [J], *Fish. J.* **11**, 1841 (林坤, 麦广铭, 王力飞, 王学锋. 2015—2018年珠江口近岸海域鱼类群落结构及其稳定性 [J], 水产学报, 2020, **11**, 1841).
- Linder P., Bykova O., Dyke J., et al. (2012) Biotic modifiers, environmental modulation and species distribution models, *J. Biogeogr.* **39**, 2179.
- Liu J., Li W., Ye S., Wang Q., Yin Z., Yuan J., Zhang T., Lee J.-K., Gui J. (1149–2020) *Calculation method of aquaculture capacity on large water surface*. China Agricultural Press (刘家寿, 李为, 叶少文, 王齐东, 殷战, 苑晶, 张堂林, 李钟杰, 桂建芳, 大水面增养殖容量计算方法, SC/T 1149–2020, 中国农业出版社).
- Logsdon C. D., Perot K. J., McDonald A. R. (1987) Mechanism of glucocorticoid-induced increase in pancreatic amylase gene transcription, *J. Biol. Chem.* **262** (32), 15765.
- Loreau M., Naeem S., Inchausti P., Bengtsson J., Grime J. P., Hector A., Hooper D. U., Huston M. A., Raffaelli D., Schmid B., Tilman D., Wardle D. A. (2001) Biodiversity and ecosystem functioning: Current knowledge and future challenges, *Science* **294** (5543), 804.
- Lou J., Yang T., Wang T., Gao J. (2020) Structure of fish phylogenetic communities of threinternal river systems in Gansu Province [J], *J. Hubei Univ. (Nat. Sci. Ed.)* **1**, 42 (娄晋铭, 杨婷越, 王太, 高坚. 甘肃省三条内流河水系鱼类系统发育群落结构及其构建机制研究 [J], 湖北大学学报 (自然科学版), 2020, **1**, 42).
- Lu K. (1990) *Pearl river system fishery resources*. Guangdong Science and Technology Publishing House (陆奎贤, 1990, 珠江水系渔业资源, 广东科技出版社).
- Ludwig M. Z., Bergman C., Patel N. H., et al. (2000) Evidence for stabilizing selection in a eukaryotic enhancer element [J], *Nature* **403** (6769), 564.
- Luo J., Xing Y., Su X., Luo C. (1986) *Multiplex statistical analysis software for microcomputer use*. Sichuan Science and Technology Press, Chengdu (罗积玉, 邢瑛, 苏显康, 罗昌荣, 微机用多元统计分析软件, 四川科学技术出版社, 1986, 成都).
- Luo Z., Liu J., Zhou Y., Du J., Wu Q., Chai B. (2021) Soil procommunity structure and diversity insubalpine grassland, *J. Ecol.* **7**, 2783 (罗正明, 刘晋仙, 周妍英, 杜京旗, 吴强, 柴宝峰, 亚高山草地土壤原生生物群落结构和多样性海拔分布格局, 生态学报, 2021, **7**, 2783).
- Ma P., Liu Y., Reddy K. P., Chan W. K., Lam T. J. (2004a) Characterization of the seabasspancreatic alpha-amylase gene and promoter, *Gen. Comp. Endocrinol.* **137** (1), 78.
- Ma P., Sivaloganathan B., Reddy K. P., Chan W. K., Lam T. J. (2004b) Hormonal influence onamylase gene expression during Seabass (*Lates calcarifer*) larval development, *Gen. Comp. Endocrinol.* **138** (1), 14.

- Machius M., Wiegand G., Huber R. (1995) Crystal structure of calcium-depleted *Bacilluslicheniformis* alpha-amylase at 2.2 Å resolution, *J. Mol. Biol.* **246** (4), 545.
- Malizia R. W., Stigall A. L. (2011) Niche stability in Late Ordovician articulated brachiopod species before, during, and after the Richmondian Invasion [J], *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **311** (3–4), 154.
- Maron J. L., Vilá M., Bommarco R., Elmendorf S., Beardsley P. (2004) Rapid evolution of an invasive plant, *Ecol. Monogr.* **74** (2), 261.
- Mason N. W. H., Lanoiselée C., Mouillot D., *et al.* (2008) Does niche overlap control relative abundance in French lacustrine fish communities? A new method incorporating functional traits, *J. Anim. Ecol.* **77**, 661. <https://doi.org/10.1111/j.1365-2656.2008.01379.x>.
- Mason N. W. H., Lanoiselée C., Mouillot D., Irz P., Argillier C. (2007) Functional characters combined with null models reveal in consistency in mechanisms of species turnover in lacustrine fish communities, *Oecologia* **153** (2), 441.
- Matsuzaki S. S., SasaKi T., Akasaka M. (2013) Consequences of the introduction of exotic and translocated species and future extirpations on the functional diversity of fresh water fish assemblages, *Glob. Ecol. Biogeogr.* **22** (9), 1071.
- McCann K., Hastings A., Huxel G. R. (1998) Weak trophic interactions and the balance of nature [J], *Nature* **395**, 794.
- Mcnaughton S. J. (1978) Stability and diversity of ecological communities, *Nature* **274**, 251.
- Meng F., Hu A., Wang J. (2020) Microbial traits reveal species distribution patterns, community building mechanisms, and ecosystem functions [J], *J. Microbiol.* **9**, 1784 (孟凡凡, 胡鑫, 王建军. 微生物性状揭示物种分布格局、群落构建机制和生态系统功能 [J], *微生物学报*, 2020, **9**, 1784).
- Miller J. (2010) Species distribution modeling, *Geogr. Compass* **4**, 490.
- Miller J. A. (2013) Species distribution models: Spatial autocorrelation and non-stationarity, *Prog. Phys. Geog.* **37**, 161.
- Miller T. J. (2007) Contribution of individual-based coupled physical-biological models to understanding recruitment in marine fish populations [J], *Mar. Ecol. Prog. Ser.* **347**, 127.
- Moal J., Daniel J. Y., Sello D., *et al.* (2000) Amylase mRNA expression in *Crassostrea gigas* during feeding cycles [J], *J. Comp. Physiol. B: Biochem. Syst. Environ. Physiol.* **170** (1), 21.
- Moore J. C., Hunt H. W. (1988) Resource compartmentation and the stability of real ecosystems [J], *Nature* **333**, 261.
- Mouillot D., Dumay O., Tomasini J. A. (2007) Limiting similarity, niche filtering and functional diversity in coastal lagoon fish communities, *Estuarine Coastal Shelf Sci.* **71** (3/4), 443.
- Mu Y., Liu Y., Wang J., *et al.* (1999) Effect of water temperature and light on gonad maturation and spawning in flounder [J], *J. Dalian Ocean Univ.* **14** (2), 62 (木云雷, 刘悦, 王鉴, 等. 水温和光照对牙鲆亲鱼性腺成熟和产卵的影响 [J], *大连海洋大学学报*, 1999, **14** (2), 62).
- Mu-Lian R., *et al.* (1998) Yili River Fish, *J. Fish.* **11** (1), 7 (任慕莲 等, 伊犁河鱼类, *水产学杂志*, 1998, **11**, 1, 7).
- Murdoch W. W. (1975) Diversity, complexity, stability and pest control, *J. Appl. Ecol.* **12**, 795.
- Naem S., Wright J. P. (2003) Disentangling biodiversity effects on ecosystem functioning: Deriving solutions to a seemingly insurmountable problem, *Ecol. Lett.* **6**, 567. <https://doi.org/10.1046/j.1461-0248.2003.00471.x>.
- Nelson J. S., Grande T. C., Wilson M. V. H. (2016) *Fishes of the world* [M]. John Wiley & Sons, Hoboken.

- Nic Lughadha E., Walker B. E., Canteiro C., *et al.* (2018) The use and misuse of herbarium specimens in evaluating plant extinction risks, *Phil. Trans. R. Soc. B* **374**, 20170402. <https://doi.org/10.1098/rstb.2017.0402>.
- Nyström M., Folke C. (2001) Spatial resilience of coral reefs, *Ecosystems* **4** (5), 406.
- Odom D. T., Dowell R. D., Jacobsen E. S., Nekludova L., Rolfe P. A., Danford T. W., Gifford D. K., Fraenkel E., Bell G. I., Young R. A. (2006) Core transcriptional regulatory circuitry in human hepatocytes, *Mol. Syst. Biol.* **2**, 2006.
- Ostrand K. G., Wilde G. R. (2002) Seasonal and spatial variation in a prairie stream – Fish assemblage, *Ecol. Freshw. Fish* **11** (3), 137.
- Ovaskainen O., Roy D. B., Fox R., Anderson B. J. (2016) Uncovering hidden spatial structure in species communities with spatially explicit joint species distribution models [J], *Methods Ecol. Evol.* **7** (4), 428.
- Pandey A., Nigam P., Soccol C. R., Soccol V. T., Singh D., Mohan R. (2000) Advances in microbial amylases, *Biotechnol. Appl. Biochem.* **31**, 132.
- Pearl River Fisheries Research Institute, Chinese Academy of Fishery Sciences, South China Normal University, Jinan University, Zhanjiang Fisheries College, Shanghai Fisheries University. (1991) *Guangdong freshwater fish annals*. Guangdong Science and Technology Press, Guangzhou (中国水产科学研究院珠江水产研究所, 华南师范大学, 暨南大学, 湛江水产学院, 上海水产大学等, 广东淡水鱼类志, 广东科技出版社, 广州, 1991).
- Pearl River Fisheries Research Institute, Chinese Academy of Fishery Sciences, Shanghai Fisheries University, East China Sea Fisheries Research Institute, Chinese Academy of Fishery Sciences, Guangdong Fisheries School. (1986) *Annals of freshwater and estuary in Hainan Island*. Guangdong Science and Technology Publishing House, Guangzhou (中国水产科学研究院珠江水产研究所, 上海水产大学, 中国水产科学研究院东海水产研究所, 广东省水产学校等, 海南岛淡水及河口鱼类志, 广东科技出版社, 广州, 1986).
- Peres-Neto P., Legendre P., Dray S., Borcard D. (2006) Variation partitioning of species data matrices: Estimation and comparison of fractions, *Ecology* **87** (10), 2614.
- Petchey O. L., Gaston K. J. (2006) Functional diversity: Back to basics and looking forward, *Ecol. Lett.* **9** (6), 741.
- Peterson A., Soberon J., Sanchez-Cordero V. (1999) Conservatism of ecological niches in evolutionary time, *Science* **285** (5431), 1265.
- Peterson A. T., Papes M., Eaton M. (2007) Transferability and model evaluation in ecological niche modeling: A comparison of GARP and Maxent, *Ecography* **30**, 550.
- Peterson B. J. (1986) Scientific-information – Opportunities, requirements, rewards, *Abstracts Papers Am. Chem. Soc.* **191**, 174.
- Peterson B. J., Fry B. (1987) Stable isotope in ecosystem studies, *Annu. Rev. Ecol. Syst.* **18**, 293.
- Peterson B. J., Howarth R. W. (1985) Sulfur, carbon and nitrogen isotopes as tracers of organic-matter flow in Georgia Estuaries, *Estuaries* **8** (2B), A88.
- Peterson B. J., Howarth R. W., Garritt R. H. (1985) Multiple stable isotopes used to trace the flow of organic matter in estuarine food webs, *Science* **227**, 1361.
- Peterson B. J., Howarth R. W., Garritt R. H. (1986) Sulfur and carbon isotopes as tracers of salt-marsh organic matter flow, *Ecology* **67**, 865.
- Pimm S. L. (1984) The complexity and stability of ecosystems [J], *Nature* **307**, 321.

- Pinnegar J. K., Jennings S., O'Brien C. M., Polunin N. V. C. (2002) Long-term changes in the trophic level of the Celtic Sea fish community and fish market price distribution, *J. Appl. Ecol.* **39** (3), 377. <https://doi.org/10.1046/j.1365-2664.2002.00723.x>.
- Pof N. L., Allan J. D. (1995) Functional organization of stream fish assemblages in relation to hydrological variability, *Ecology* **76** (2), 606.
- Polis G. A., Strong D. R. (1996a) Food web complexity and community dynamics [J], *Nature* **147**, 813.
- Polis G. A., Strong D. R. (1996b) Food web complexity and community dynamics, *Am. Nat.: Devoted Concept. Unification Biol. Sci.* **147** (5), 813.
- Post D. M. (2002) Using stable isotopes to estimate trophic position: Models, methods, and assumptions, *Ecology* **83**, 703.
- Prieur-Richard A. H., Lavorel S. (2000) Invasions: The perspective of diverse plant communities, *Aust. Ecol.* **25** (1), 1.
- Pulliam H. R. (2000) On the relationship between niche and distribution, *Ecol. Lett.* **3**, 349.
- Qin B., Evergreen, Yu C., Liu S., Ma Q., Zheng K. K. (2013) Cloning of alpha-amylase genes of *Cynoglossus semilaevis* Günther and effect of taurine on its expression [J], *Ocean Lake* **4**, 988 [秦帮勇, 常青, 于朝磊, 柳淑芳, 马零, 郑珂珂, 半滑舌鲷(*Cynoglossus semilaevis* Günther) α -淀粉酶基因的克隆及牛磺酸对其表达的影响 [J], *海洋与湖沼*, 2013, **4**, 988].
- Radler P. D., Wehde B. L., Wagner K. U. (2017) Crosstalk between STAT5 activation and PI3K/AKT functions in normal and transformed mammary epithelial cells, *Mol. Cell. Endocrinol.* **451**, 31.
- Randin C. F., Dirnböck T., Dullinger S., *et al.* (2006) Are niche-based species distribution models transferable in space? *J Biogeogr.* **33**, 1689.
- Rangel T. F., Loyola R. D. (2012) Labeling ecological niche models, *Nat. Conservacao* **10** (2), 119.
- Reeve A. H., Borregaard M. K., Fjeldså J. (2016) Negative range size-abundance relationships in Indo-Pacific bird communities, *Ecography* **39**, 990.
- Rejmanek M., Stary P. (1979) Connectance in real biotic communities and critical values for stability of model ecosystems [J], *Nature* **280**, 311.
- Ren M., Guo Y., Zhang R., Cai L., Adk, Li H., Fu Y., Liu K., *et al.* (2002) Composition of fish and fish flora of Irzise River in China, arid zone research **19** (2), 62 [任慕莲, 郭焱, 张人铭, 蔡林刚, 阿达克, 李红, 付雅丽, 刘昆仑等, 我国额尔齐斯河鱼类及鱼类区系组成, 干旱区研究, 2002, **19** (2), 62].
- Rochet M. J., Trenkel V. M. (2003) Which community indicators can measure the impact of fishing? A review and proposals [J], *Can. J. Fish. Aquat. Sci.* **60**, 86.
- Sæbø J. S. (2016) *Spatial and temporal distributions and interactions in a neotropical ground-dwelling animal community*. Master dissertation, Norwegian University, Ås.
- Schoener T. W. (1974) Resource partitioning in ecological communities, *Science* **185**, 27.
- Schurr F. M., Pagel J., Cabral J. S., Groeneveld J., Bykova O., O'Hara R. B., Hartig F., Kissling W. D., Linder H. P., Midgley G. F., Schröder B., Singer A., Zimmermann N. E. (2012) How to understand species' niches and range dynamics: A demographic research agenda for biogeography [J], *J. Biogeogr.* **39** (12), 2146.
- Sexton J. P., McIntyre P. J., Angert A. L., Rice K. J. (2009) Evolution and ecology of species range limits, *Annu. Rev. Ecol. Evol. Syst.* **40** (1), 415.
- Shapiro M. D., Marks M. E., Peichel C. L., *et al.* (2004) Genetic and developmental basis of evolutionary pelvic reduction in three espine sticklebacks [J], *Nature* **428** (6984), 717.

- Sharma S., Legendre P., De Cdceres M., Boisclair D. (2011) The role of environmental and spatial processes in structuring native and non-native fish communities across thousands of lakes, *Ecography* **34** (5), 762.
- Shen Z. H., Fang J. Y., Chiu C. A., *et al.* (2015) The geographical distribution and differentiation of Chinese beech forests and the association with *Quercus*, *Appl. Veg. Sci.* **18**, 23.
- Shuai F., Lek S., Li X. (2018a) Biological invasions undermine the functional diversity of fish communities in a large subtropical river, *Biol. Invasions* **4**, 1.
- Shuai F., Li X., Chen F., Li Y., Lek S. (2017a) Spatial patterns of fish assemblages in the Pearl River, China: Environmental correlates, *Fundam. Appl. Limnol.* **189** (4), 329.
- Shuai F., Li X., He A., Liu Q., Zhang Y., Wu Z., Zhu S. (2020) Characteristics of fish diversity in Guangxi Pearl River System, **4**, 819 (帅方敏, 李新辉, 何安尤, 刘乾甫, 张迎秋, 武智, 朱书礼, 珠江水系广西江段鱼类多样性空间分布特征, 水生生物学报, 2020, **4**, 819).
- Shuai F., Li X., Li Y., Jie L., Jiping Y., Lek S. (2015) Forecasting the invasive potential of Nile tilapia (*Oreochromis niloticus*) in a large subtropical river using a univariate approach, *Fundam. Appl. Limnol.* **187**, 165.
- Shuai F., Li X., Li Y., Li J., Yang J., Lek S. (2016) Temporal patterns of larval fish occurrence in a large subtropical river, *PLoS ONE* **11** (5), 1.
- Shuai F., Li X., Liu C., *et al.* (2017c) Research on the spatial distribution pattern of fish community diversity in the Pearl River system, *J. Ecol.* **37** (19), 3182 (帅方敏, 李新辉, 刘乾甫等, 珠江水系鱼类群落多样性空间分布格局研究, 生态学报, 2017, **37** (19), 3182).
- Shuai F., Li X., Liu Q., Zhu S., Wu Z., Zhang Y. (2019) Nile tilapia (*Oreochromis niloticus*) invasions disrupt the functional patterns of fish community in a large subtropical river in China, *Fish. Manage. Ecol.* **26** (6), 578.
- Shuai F., Yu S., Lek S., Li X. (2018b) Habitat effects on intra-species variation in functional morphology: Evidence from freshwater fish, *Ecol. Evol.* **8** (22), 10902.
- Shuai F. M., Li X., Chen F., Li Y., Yang J., Li J., Wu Z. (2017b) Functional diversity of freshwater fish, *J. Ecol.* **37** (15), 1 (帅方敏, 李新辉, 陈方灿, 李跃飞, 杨计平, 李捷, 武智. 淡水鱼类功能多样性及其研究方法. 生态学报, 2017, **37** (15), 1).
- Shurin J. B., Amarasekare P., Chase J. M., *et al.* (2004) Alternative stable states and regional community structure, *J. Theor. Biol.* **227** (3), 359.
- Slater E. P., Hesse H., Muller J. M., Beato M. (1993) Glucocorticoid receptor binding site in the mouse alpha-amylase 2 gene mediates response to the hormone, *Mol. Endocrinol.* **7** (7), 907.
- Soberón J., Peterson A. T. (2005) Interpretation of models of fundamental ecological niches and species' distributional areas, *Biodivers. Inf.* **2**, 1.
- Stachowicz J. J. (2001) Mutualism, facilitation, and the structure of ecological communities: Positive interactions play a critical, but underappreciated, role in ecological communities by reducing physical or biotic stresses in existing habitats and by creating new habitats on which many species depend, *BioScience* **51** (3), 235.
- Stanley T. R., Royle J. A. (2005) Estimating site occupancy and abundance using indirect detection indices [J], *J. Wildlife Manage.* **69** (3), 874.
- Stigall A. L. (2012) Using ecological niche modelling to evaluate niche stability in deep time [J], *J. Biogeogr.* **39** (4), 772.
- Stockwell D., Peters D. (1998) The GARP modelling system: Problems and solutions to automated spatial prediction, *Int. J. Geogr. Inf. Sci.* **13**, 143.

- Stockwell D. R. B., Peterson A. T. (2002) Effects of sample size on accuracy of species distribution models [J], *Ecol. Model.* **148** (1), 1.
- Strecker A., Olden J., Whittier J., *et al.* (2011) Defining conservation priorities for freshwater fishes according to taxonomic, functional, and phylogenetic diversity, *Ecol. App.* **21**, 3002.
- Sun D., Liu X., Zhou S. (2018) The restoration process and community construction of alpine meadow diversity after artificial removal of plant functional groups was stopped [J], *Biodiversity* **7**, 655 (孙德鑫, 刘向, 周淑荣. 停止人为去除植物功能群后的高寒草甸多样性恢复过程与群落构建 [J], *生物多样性*, 2018, **7**, 655).
- Svenning J.-C., Fløjgaard C., Marske K. A., Nógues-Bravo D., Normand S. (2011) Applications of species distribution modeling to paleobiology [J], *Quat. Sci. Rev.* **30** (21–22), 2930.
- Swenson N. G., Enquist B. J., Pither J., *et al.* (2006) The problem and promise of scale dependency in community phylogenetics [J], *Ecology* **87** (10), 2418.
- Takahiro N., Yusuke N., Mitsuru E., *et al.* (1986) Primary structure of human salivary α -amylase gene [J], *Gene* **41** (2–3), 299.
- Takimoto G., Post D. M., Spiller D. A., *et al.* (2012) Effects of productivity, disturbance, and ecosystem size on food-chain length: Insights from a metacommunity model of intraguild predation, *Ecol. Res.* **27** (3), 481.
- Tan X., Li X., Li Y., Li J. (2012) Nile tilapia and its spatial distribution in the Pearl River system, *J. Biosaf.* **21** (4), 295 (谭细物, 李新辉, 李跃飞, 李捷, 尼罗罗非鱼早期发育形态及其在珠江水系的分布, *生物安全学报*, 2012, **21** (4), 295).
- Tan X., Li Y., Lai Z., Luo J., Li X., *et al.* (2010) Study on community structure and annual change of fish fry in Zhaoqing section of Xijiang River, **3** (5), 27 (谭细物, 李跃飞, 赖子尼, 罗建仁, 李新辉等. 西江肇庆段鱼苗群落结构组成及其周年变化研究, *水生态学杂志*, 2010, **3** (5), 27).
- Tan Z., Niu B., Tsang K. Y., Melhado I. G., Ohba S., He X., Huang Y., Wang C., McMahon A. P., Jauch R., *et al.* (2018) Synergistic co-regulation and competition by a SOX9-GLI-FOXA phasic transcriptional network coordinate chondrocyte differentiation transitions, *PLoS Genet.* **14** (4), e1007346.
- Tanner J. E., Hughes T. P., Connell J. H. (1995) Species coexistence, keystone species, and succession: A sensitivity analysis, *Oceanogr. Lit. Rev.* **42** (5), 378.
- Tedesco, *et al.* (2017) Data descriptor: A global database on freshwater fish species occurrence in drainage basins, *Sci. Data* **4**, 170141. <https://doi.org/10.1038/sdata.2017.141>.
- Tilman D., Knops J., Wedin D., Reich P., Ritchie M., Siemann E. (1997) The influence of functional diversity and composition on ecosystem processes, *Science* **277** (5330), 1300.
- Ulrich W., Hajdamowicz I., Zalewski M., *et al.* (2010) Species assortment or habitat filtering: A case study of spider communities on lake islands [J], *Ecol. Res.* **25**, 375.
- Urban M. C. (2004) Disturbance heterogeneity determines freshwater metacommunity structure, *Ecology* **85** (11), 2971.
- Václavík T., Meentemeyer R. K. (2009) Invasive species distribution modeling (iSDM): Are absence data and dispersal constraints needed to predict actual distributions? *Ecol. Model.* **220**, 3248.
- Vamosi S. M., Heard S. B., Vamosi J. C., *et al.* (2009) Emerging patterns in the comparative analysis of phylogenetic community structure [J], *Mol. Ecol.* **18** (4), 572.
- van der Laan R. (2017) *Almere*. The Netherlands. Freshwater fish list (Online). ISSN: 2468-9157.
- Vandermeer J. H. (1972) Niche theory [J], *Ann. Rev. Ecol. Syst.* **3** (1), 107.
- Vanni M. J., Arend K. K., Bremigan M. T., *et al.* (2005) Linking landscapes and food webs: Effects of omnivorous fish and watersheds on reservoir ecosystems, *BioScience* **55** (2), 155.

- Vanschoenwinkel B., Waterkeyn A., Jocqué M., *et al.* (2010) Species sorting in space and time – The impact of disturbance regime on community assembly in a temporary pool metacommunity, *J. North Am. Benthol. Soc.* **29** (4), 1267.
- Vellend M. (2010) Conceptual synthesis in community ecology [J], *Q. Rev. Biol.* **85** (2), 183.
- Villéger S., Moullot D. (2008) Additive partitioning of diversity including species differences: A comment on Hardy & Senterre (2007), *J. Ecol.* **96** (5), 845.
- Vilmi A., Karjalainen S. M., Hellsten S., Heino J. (2016) Bioassessment in a metacommunity context: Are diatom communities structured solely by species sorting? [J], *Ecol. Indic.* **62**, 86.
- Walker P. A., Cocks K. D. (1991) HABITAT: A procedure for modelling a disjoint environmental envelope for a plant or animal species, *Glob. Ecol. Biogeogr. Lett.* **1**, 108.
- Walls B. J., Stigall A. L. (2011) Analyzing niche stability and biogeography of Late Ordovician brachiopod species using ecological niche modeling [J], *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **299** (1–2), 15.
- Wang Q., Lu Z. X., Zhao J. W., Chen Y. Q. (2020) The impact path and intensity of mutualism and plant diversity on different trophic levels of arthropod community, *Acta Ecol. Sinica* **40** (1), 51.
- Wang R., Li J. (2010) Analysis of the influence of water temperature, flow velocity and water depth of diversion hydropower station [J], *Sichuan Hydropower Gener.* **29** (2), 76 (王锐, 李嘉. 引水式水电站减水河段的水温、流速及水深变化对鱼类产卵的影响分析 [J], *四川水力发电*, 2010, **29** (2), 76).
- Wang R. W., Shi L., Ai S. M., Zheng Q. (2008) Trade-off between reciprocal mutualists: Local resource availability-oriented interaction in fig/fig wasp mutualism, *J. Anim. Ecol.* **77** (3), 616.
- Wang Y. P., Lu Y., Zhang Y., *et al.* (2015) The draft genome of the grass carp (*Ctenopharyngodon idellus*) provides insights into its evolution and vegetarian adaptation [J], *Nat. Genet.* **47** (6), 625.
- Wang Z., Chen M., Lv S., Xu C., Lei G. (2006) Species diversity and spatio-temporal distribution patterns of icefishes (Salangidae) in Poyang Lake, *J. Ecol.* **5**, 1337 (王忠锁, 陈明华, 吕偲, 许崇任, 雷光春, 鄱阳湖银鱼多样性及其时空格局, *生态学报*, 2006, **5**, 1337).
- Wang Z., Mehra V., Simpson M. T., Maunze B., Chakraborty A., Holan L., Eastwood E., Blackmore M. G., Venkatesh I. (2018) KLF6 and STAT3 co-occupy regulatory DNA and functionally synergize to promote axon growth in CNS neurons, *Sci. Rep.* **8** (1), 12565.
- Warwick R., Clarke K. (1998) Taxonomic distinctness and environmental assessment, *J. Appl. Ecol.* **35**, 532.
- Webb C. O., Ackerly D. D., Kembel S. W. (2008) Phylocom: Software for the analysis of phylogenetic community structure and trait evolution [J], *Bioinformatics* **24**, 2098.
- Webb C. O., Ackerly D. D., McPeck M. A., Donoghue M. J. (2002) Phylogenies and community ecology [J], *Annu. Rev. Ecol. Syst.* **33**, 475.
- Weinrich L., Meister A., Rutter W. J. (1991) Exocrine pancreas transcription factor 1 binds to a bipartite enhancer element and activates transcription of acinar genes, *Mol. Cell. Biol.* **11** (10), 4985.
- Wiens J. A. (1989) Spatial scaling in ecology [J], *Funct. Ecol.* **3**, 385.
- Winemiller K. O., Polis G. A. (1996) Food webs: What can they tell us about the world? *Food Webs*, 1.
- Wisz M. S., Pottier J., Kissling W. D., Pellissier L., Lenoir J., Damgaard C. F., Dormann C. F., Forchhammer M. C., Grytnes J.-A., Guisan A., Heikkinen R. K., Høye T. T., Kühn I., Luoto M., Maiorano L., Nilsson M.-C., Normand S., Öckinger E., Schmidt N. M., Termansen M., Timmermann A., Wardle D. A., Aastrup P., Svenning J.-C. (2013) The role of biotic interactions in shaping distributions and realised assemblages of species: Implications for species distribution modelling [J], *Biol. Rev.* **88** (1), 15.

- Wright J. W., Davies K. F., Lau J. A., McCall A. C., McKay J. K. (2006) Experimental verification of ecological niche modeling in a heterogeneous environment, *Ecology* **87** (10), 2433.
- Wu L., et al. (1989) *Guizhou fish annals*. Guizhou People's Publishing House, Guiyang (伍律等, 贵州鱼类志, 贵州人民出版社, 贵阳, 1989).
- Wu X., et al. (1964) *Chinese carp Cyprinus carpio Linnaeus, Cyprininae, 1758 families*. Shanghai Science and Technology Press, Shanghai (伍献文等, 中国鲤 Cyprinus carpio Linnaeus, 1758科 Cyprininae鱼类志 上, 上海科学技术出版社, 上海, 1964).
- Wu X., et al. (1982) *Chinese carp Cyprinus carpio Linnaeus, 1758 family Cyprininae fish records*. Shanghai Science and Technology Press, Shanghai (伍献文等, 中国鲤 Cyprinus carpio Linnaeus, 1758科 Cyprininae鱼类志 下, 上海科学技术出版社, 上海, 1982).
- Wu X., Yang G., Le P., et al. (1963) *China*. Science Press, Beijing (伍献文, 杨干荣, 乐佩琦等, 中国经济动物志淡水鱼类, 科学出版社, 北京, 1963).
- Xia Y., Li Y., Zhu S., Li J., Li S., Li X. (2020) Individual dietary specialization reduces intraspecific competition, rather than feeding activity, in black amur bream (*Megalobrama terminalis*), *Sci. Rep.* **10** (1), 17961. <https://doi.org/10.1038/s41598-020-74997-8>.
- Xia Y., Zhao W., Xie Y., Xue H., Li J., Li Y., Chen W., Huang Y., Li X. (2019) Ecological and economic impacts of exotic fish species on fisheries in the Pearl River basin, *Manage. Biol. Invasions* **10** (1), 127.
- Xia Y. G., Li J., Li Y. F., Zhu S. L., Huang Y. F., Wu Z., Liu Q. H., Li X. H. (2017) Small-subunit ribosomal DNA sequencing analysis of dietary shifts during gonad maturation in wild black Amur bream (*Megalobrama terminalis*) in the lower reaches of the Pearl River, *Fish. Sci.* **83** (6), 955.
- Xing Y., Zhang C., Fan E., Zhao Y. (2016) Freshwater fishes of China: Species richness, endemism, threatened species and conservation, *Diversity Distrib.* **22** (3), 358.
- Xiong Y., Zhang M., Zhang H., Su G., Sa Y., Xu J. (2015) A preliminary study on the relationship between fish morphological characteristics and trophic level location? *J. Lake Sci.* **27** (3), 466 (熊鹰, 张敏, 张欢, 苏国欢, 沙泳翠, 徐军, 鱼类形态特征与营养级位置之间关系初探? 湖泊科学, 2015, **27** (3), 466).
- Xu C., Wang S., Zhao F., Yang G., Zhuang P. (2019) The nutritional structure and changes of the Aquatic Animal Food Network in the Yangtze Estuary, *Aquat. Biol. J.* **1**, 155 (徐超, 王思凯, 赵峰, 杨刚, 庄平, 长江口水生动物食物网营养结构及其变化, 水生生物学报, 2019, **1**, 155).
- Xu T., Li X., Li Y., Yang J., Wu Z., Zhu S., Li J. (2018) Early status of fish resources in Jinling River in the middle reaches of Yujiang River, *South. Fish. Sci.* **14** (2), 19 (徐田振, 李新辉, 李跃飞, 杨计平, 武智, 朱书礼, 李捷, 郁江中游金陵江段鱼类早期资源现状, 南方水产科学, 2018, **14** (2), 19).
- Xu Y., Dong S., Li S., Shen H. (2019) Ecological filtration mechanism of plant community construction, *J. Ecol.* **7**, 2267 (许驭丹, 董世魁, 李帅, 沈豪, 植物群落构建的生态过滤机制研究进展, 生态学报, 2019, **7**, 2267).
- Xu Y., Peng J. F., Qu J. H., et al. (2020) Assessing food web health with network topology and stability analysis in aquatic ecosystem [J], *Ecol. Indic.* **109**, 105820.
- Yang T., Yu D., Gao X., Liu H. (2020) Mechanism analysis of fish community construction in the middle reaches of the Yangtze River, *Hydrobiol. J.* **44** (5), 1045 (杨婷越, 俞丹, 高欣, 刘焕章, 长江中游干流鱼类群落构建机制分析, 水生生物学报, 2020, **44** (5), 1045).
- Ye X., Ren L., Kuang Z., Wang Y., Xu D. (2021) Study on the nutrient structure of Yangcheng Lake fish community based on stable isotope technology [J], *Aquat. Sci. Chin.* (6), 703 (叶学瑶, 任泷, 匡箴, 王媛, 徐东坡. 基于稳定同位素技术的阳澄湖鱼类群落营养结构研究 [J], 中国水产科学, 2021, (6), 703).
- Yodzis P. (1980) The connectance of real ecosystems [J], *Nature* **284**, 544.

- Yodzis P. (1981) The stability of real ecosystems [J], *Nature* **289**, 674.
- Yosiaki Ito, Yoh Iwasa. (1981) Evolution of litter size, *Popul. Ecol.* **23** (2), 344.
- Yu X., Ren X. (1999) *Multiple statistical analysis*. China Statistics Press, Beijing (于秀林, 任雪松, 多元统计分析, 中国统计出版社出版, 1999, 北京).
- Yu Z., Zhou C., Deng Z., et al. (1985.2~5) The spawning field of the Yangtze River after the closure of Gezhouba Water Conservancy Project [A], *Collection of ichthyology* (Series 4). Science Press, Beijing (余志堂, 周春生, 邓中森等. 葛洲坝水利枢纽工程截流后的长江四大家鱼产卵场[A], 中国鱼类学会, 鱼类学论文集(第四辑). 北京: 科学出版社, 1985.2~5).
- Zhang C. (1960) *Fish records of China*. People's Education Press, Beijing (张春霖, 中国鱼类志, 人民教育出版社, 北京, 1960).
- Zhang C., et al. (2020) *Chinese mainland fish species and distribution*. Science Press, Beijing (张春光等, 中国内陆鱼类物种与分布, 科学出版社, 2020, 北京).
- Zhang C., Zhao Y. (2016) *Fish species and distribution in inland China*. Science Press, Beijing (张春光, 赵亚辉, 中国内陆鱼类物种与分布. 北京: 科学出版社, 2016).
- Zhang H., Wu G., Zhang P., et al. (2013) Trophic fingerprint of fish communities in subtropical floodplain lakes, *Ecol. Freshw. Fish* **22** (2), 1.
- Zhang J., Fan L. (2011) Functional diversity of species and their research methods, *J. Mount. Stud.* **29** (5), 513 (张金屯, 范丽宏. 物种功能多样性及其研究方法. 山地学报, 2011, **29** (5), 513).
- Zhang T. (2005) *Life history strategies, nutritional characteristics and community structure of fish in Pingdan Pond* [PhD thesis]. Institute of Hydrobiology, Chinese Academy of Sciences, Wuhan (张堂林, 扁担塘鱼类生活史策略, 营养特征及群落结构研究[博士学位论文]. 武汉: 中国科学院水生生物研究所, 2005).
- Zhang Y., Yellow R. F., Li X., Liu C., Li J., Li Y., Yang J., Zhu S. (2020) Fish community structure and environmental impact analysis of Xijiang River, *South. Aquat. Sci.* **16** (1), 42 (张迎秋, 黄稻田, 李新辉, 刘乾甫, 李捷, 李跃飞, 杨计平, 朱书礼, 西江鱼类群落结构和环境影响分析, 南方水产科学, 2020, **16** (1), 42).
- Zhao T., Villéger S., Lek S., Cucherousset J. (2014) High intraspecific variability in the functional niche of a predator is associated with onto genetics shift and individual specialization, *Ecol. Evol.* **4** (24), 4649.
- Zheng C. (Ed.). (1989) *Fish annals of the Pearl River*. Science Press, Beijing (郑慈英, 珠江鱼类志, 科学出版社, 北京, 1989).
- Zhou L. (1986) How much carbon can terrestrial and water plants each fix by photosynthesis? [J], *Teach. Biol.* **2**, 47 (周良骏. 地球上陆生和水体植物光合作用各能固定多少碳? [J], 生物学教学, 1986, **2**, 47).
- Zhou W., Ding D., Suo A., He W., Tian T. (2021) Selection method of key function group of fishery resources in the Pearl River Estuary, *J. Fish.* **45** (03), 433 (周卫国, 丁德文, 索安宁, 何伟宏, 田涛, 珠江口海洋牧场渔业资源关键功能群的遴选方法, 水产学报, 2021, **45** (03), 433).
- Zhu S., Zhang Y., Chen W., Yang J., Li J., Wu Z., Li X. (2020, March) Analysis of the relationship between the pancreatic alpha amylase gene 5' flanking sequence and the feeding habits of fish, *Chin. Aquat. Sci.* **27** (3), 277 (朱书礼, 张迎秋, 陈蔚涛, 杨计平, 李捷, 武智, 李新辉, 胰 α -淀粉酶基因 5'端调控序列与鱼类食性的关系, 中国水产科学 2020 年 3 月, **27** (3), 277).
- Zintzen V., Anderson M., Roberts C., et al. (2011) Increasing variation in taxonomic distinctness reveals clusters of specialists in the deep sea, *Ecography* **34**, 306.
- Zobel M. (1992) Plant species co-existence: The role of historical, evolutionary and ecological factors [J], *Oikos* **65** (2), 314.

